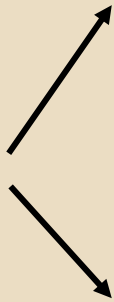
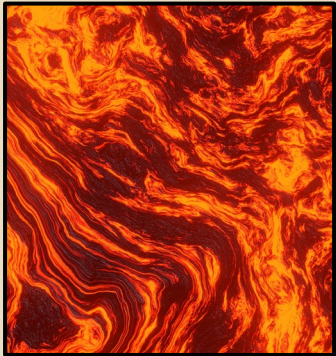


# The Structure and Dynamics of Mitotic Waves forming the *Drosophila* Blastoderm

Harrison Oatman

# Many natural phenomena feature changes in density



# Increasing the density of a system is simple

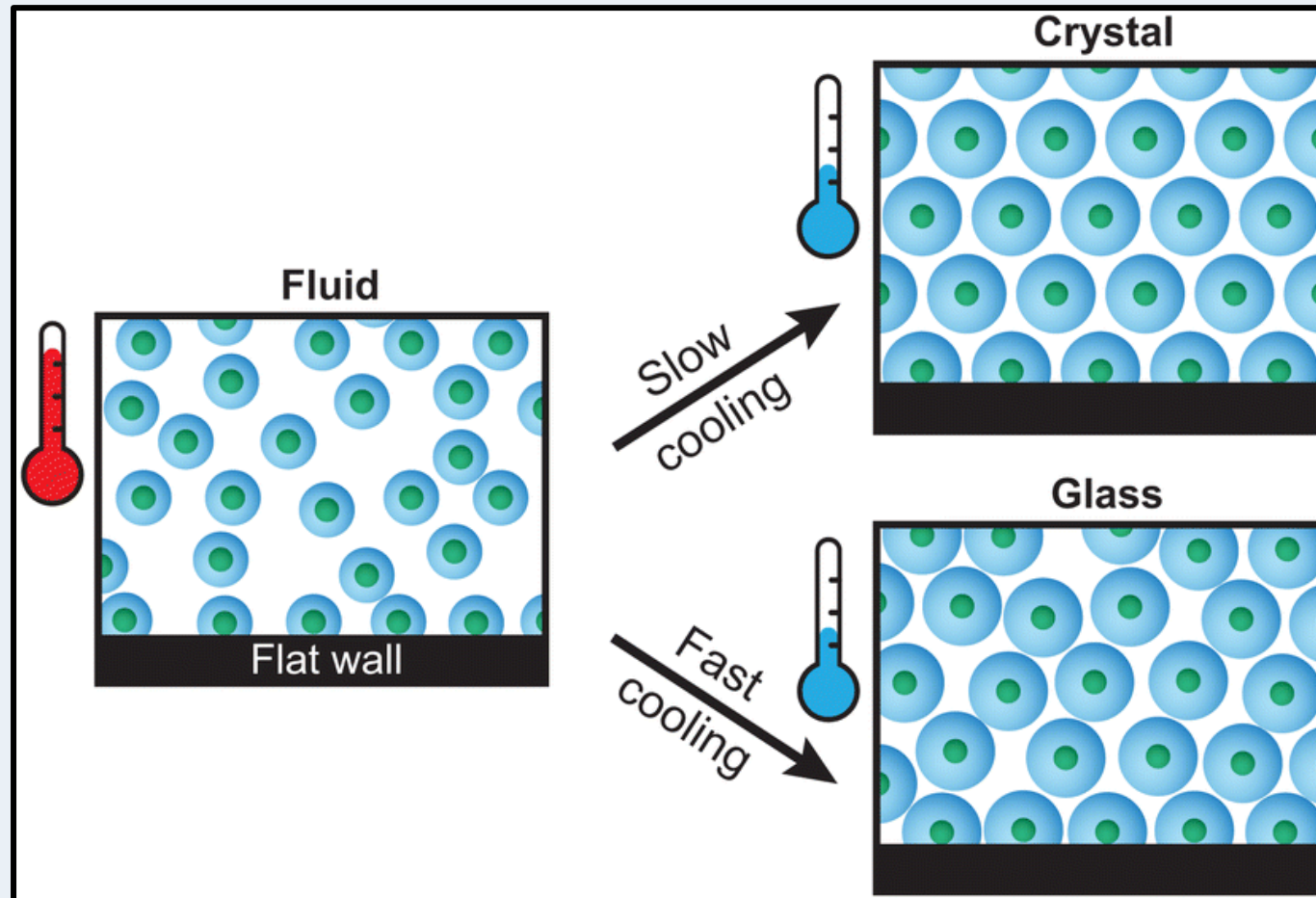
$$\rho = \frac{N}{V}$$

Decrease  $V$

Increase  $N$

...but the approach matters!

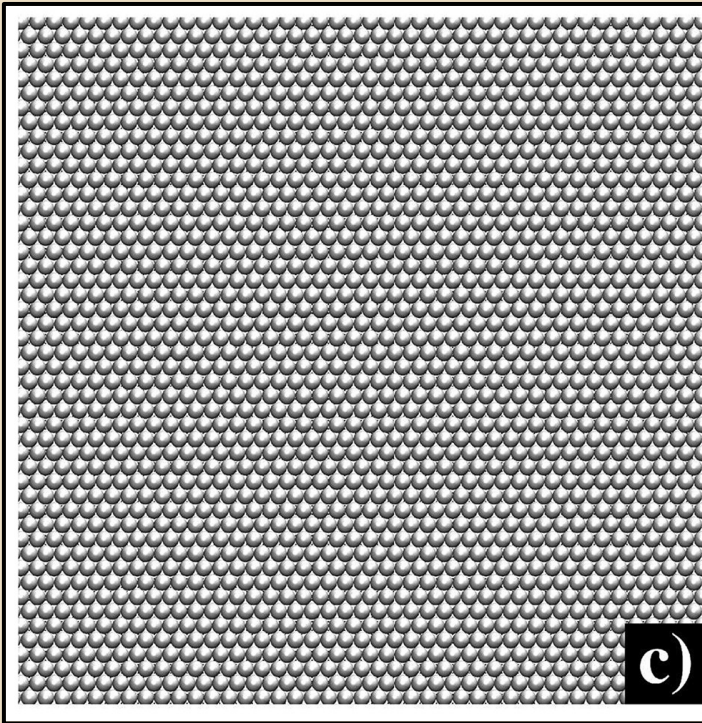
# Decreasing $V$ : rate of cooling affects the resulting structure



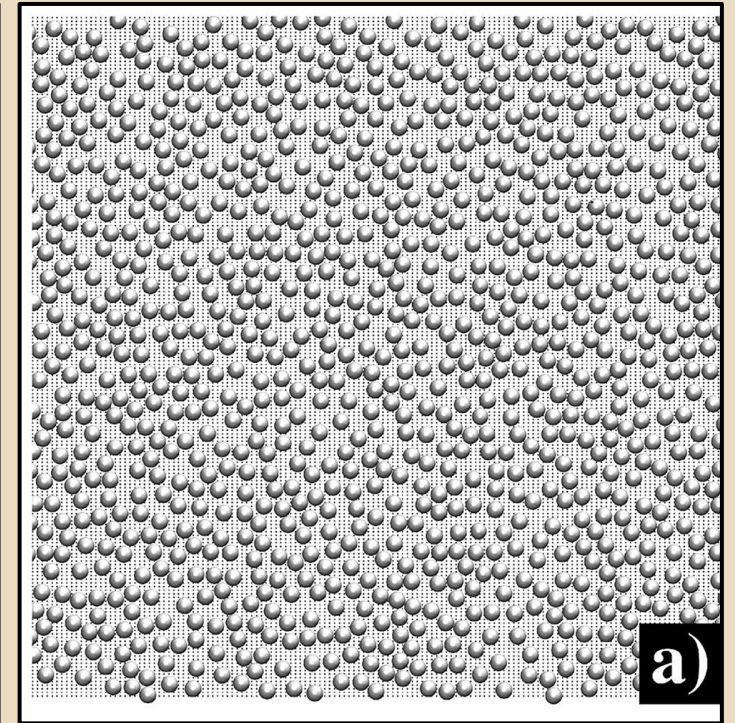
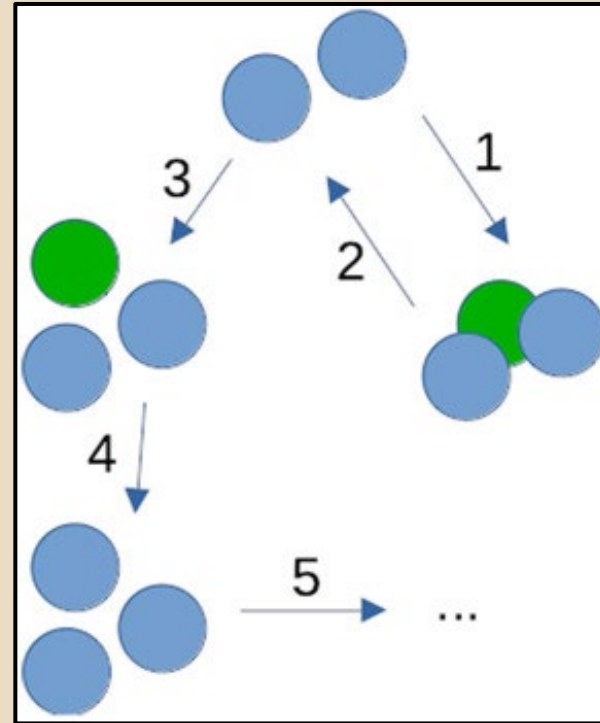


# Increasing N: random sequential adsorption produces a less than maximal coverage

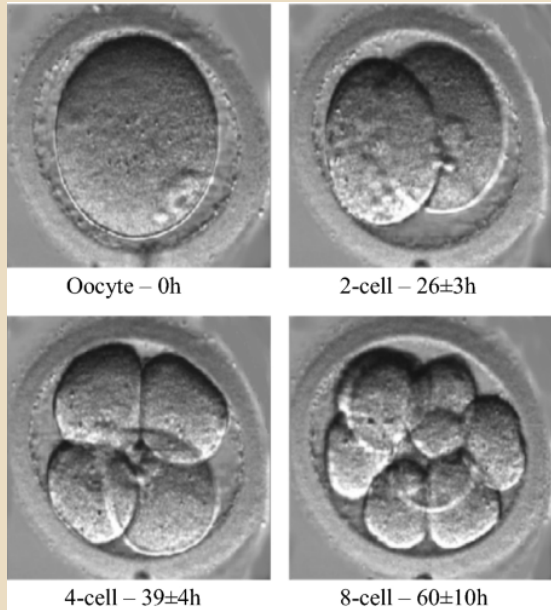
coverage  $\approx 0.909$



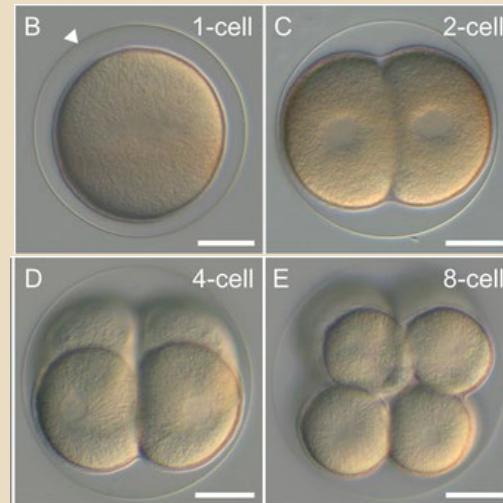
coverage  $\approx 0.547$



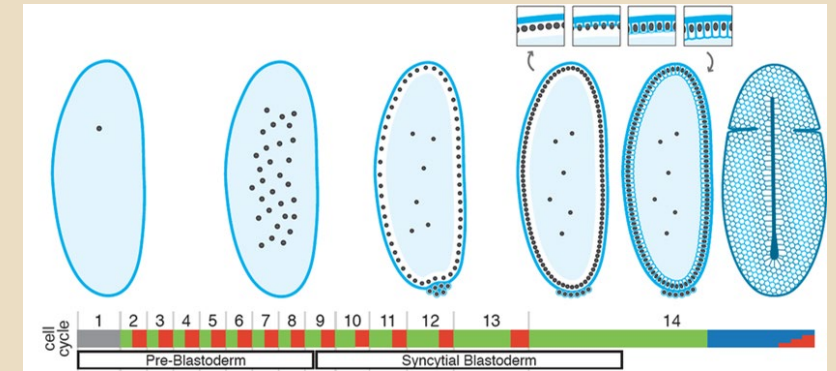
# Densification of nuclei is a universal task of multicellular embryogenesis



*Malmsten et al. (2020)*



*Formery et al. (2022)*



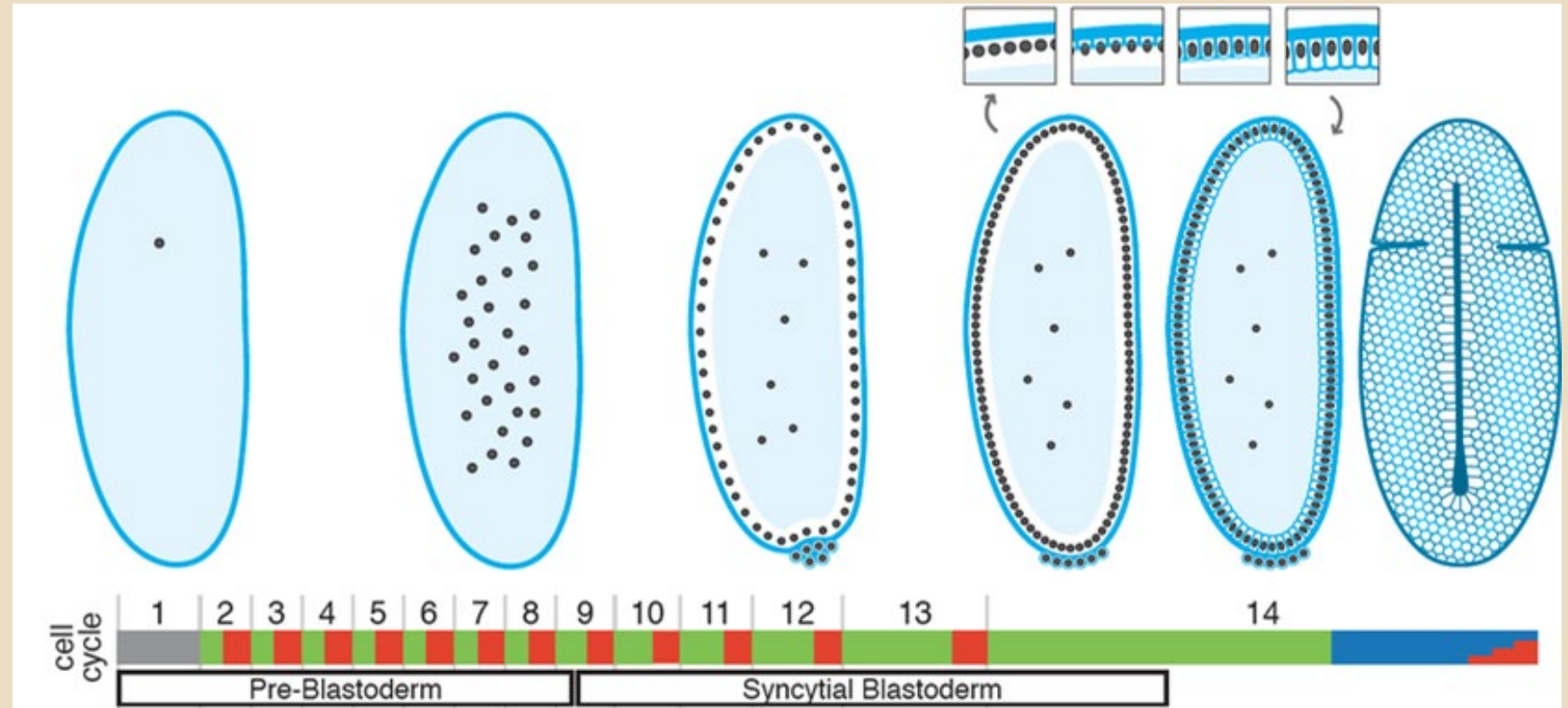
*Farrell and O'Farrell (2014)*

How does the densification approach affect later embryonic structure?



# *Drosophila* development begins with 13 nuclear divisions

- 9 divisions take place in the yolk
- Nuclei migrate to the surface of the egg, and undergo 4 more divisions
- These divisions form a blastoderm containing ~6000 nuclei



Farrell and O'Farrell (2014)

1941

# STUDIES ON THE CYTOLOGY AND EARLY EMBRYOLOGY OF THE EGG OF *DROSOPHILA MELANOGASTER*<sup>1</sup>

MORRIS RABINOWITZ

*Washington Square College, New York University*

1983

# STUDIES OF NUCLEAR AND CYTOPLASMIC BEHAVIOUR DURING THE FIVE MITOTIC CYCLES THAT PRECEDE GASTRULATION IN *DROSOPHILA* EMBRYOGENESIS

VICTORIA E. FOE AND BRUCE M. ALBERTS

*Department of Biochemistry and Biophysics, University of California at San Francisco, San Francisco, California 94143, U.S.A.*

TABLE 2

*Temperature: 24°C. Total number of eggs: 354*

STAGE OF DEVELOPMENT	NUMBER OF EGGS	MEAN AGE (IN MINUTES)
Telophase of 2nd maturation		
division to conjugation of pronuclei	13	15 ± 1.21
1st division of cleavage nuclei	10	23 ± 1.72
2nd division of cleavage nuclei	14	34 ± 1.72
3rd division of cleavage nuclei	17	47 ± 1.47
4th division of cleavage nuclei	10	53 ± 1.05
5th division of cleavage nuclei	18	60 ± 0.57
6th division of cleavage nuclei	22	70 ± 0.58
7th division of cleavage nuclei	14	78 ± 1.28
8th division of cleavage nuclei	31	93 ± 1.13
1st division of blasteme nuclei	82	99 ± 0.60
2nd division of blasteme nuclei	97	109 ± 0.51
3rd division of blasteme nuclei	24	120 ± 0.90

V. E. Foe and B. M. Alberts

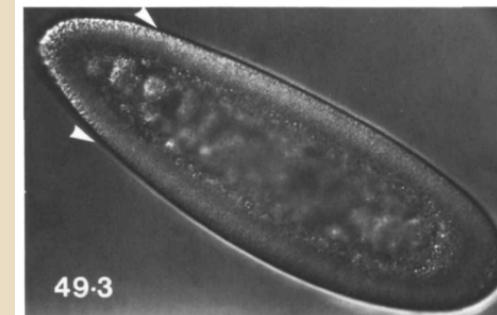


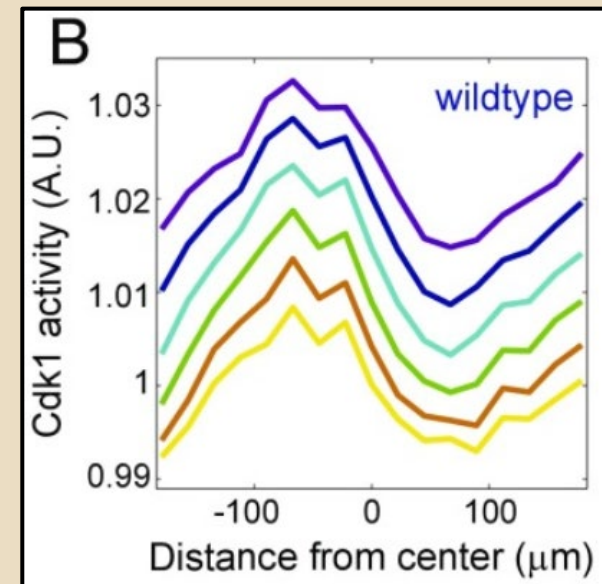
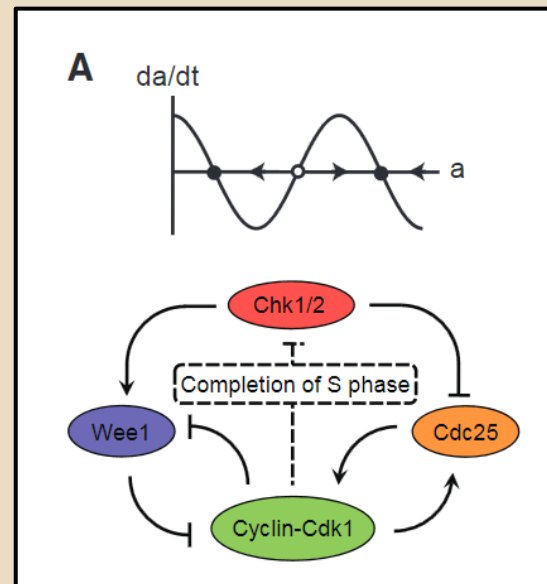
Fig. 7

# Previous works have investigated the chemical interactions underlying the mitotic waves

► Proc Natl Acad Sci U S A. 2018 Feb 15;115(10):E2165–E2174. doi: [10.1073/pnas.1714873115](https://doi.org/10.1073/pnas.1714873115)

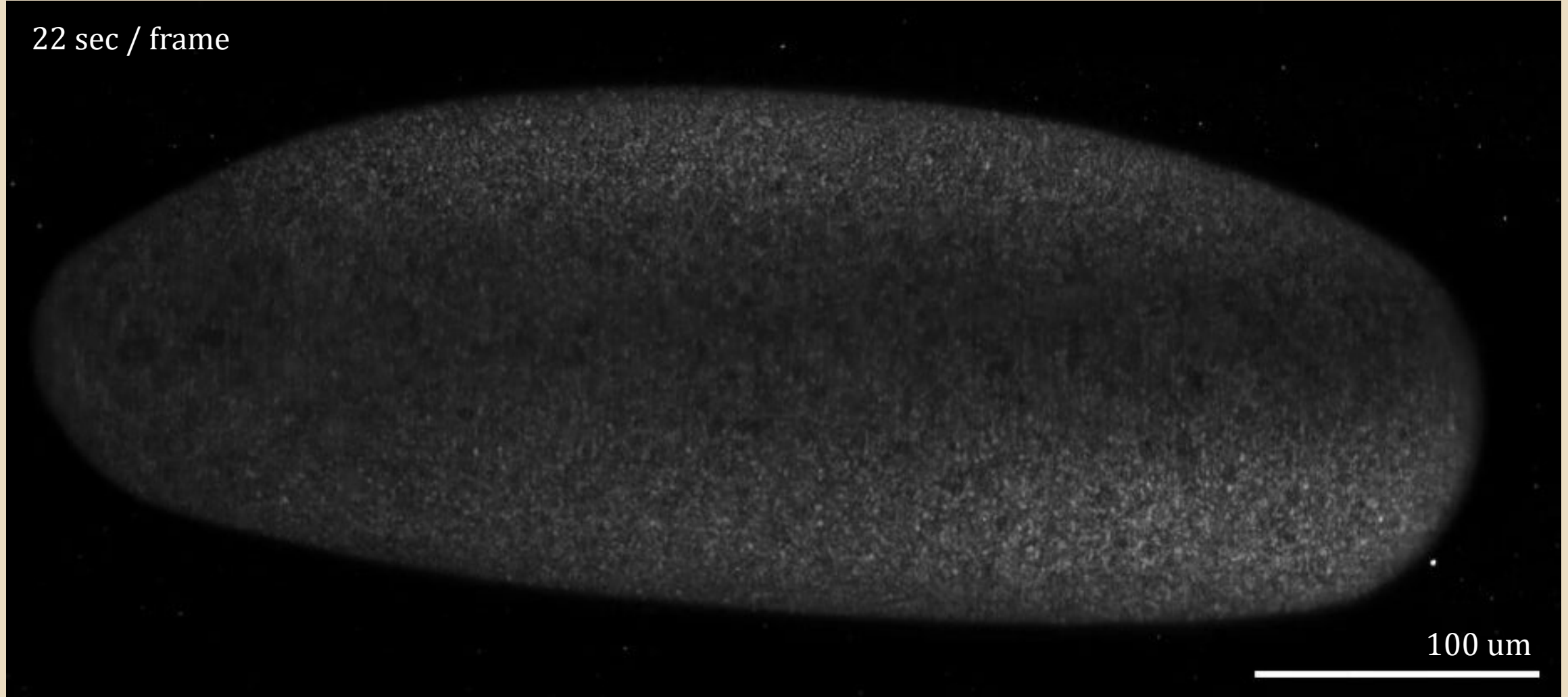
## Mitotic waves in the early embryogenesis of *Drosophila*: Bistability traded for speed

Massimo Vergassola <sup>a,1</sup>, Victoria E Deneke <sup>b</sup>, Stefano Di Talia <sup>b,1</sup>



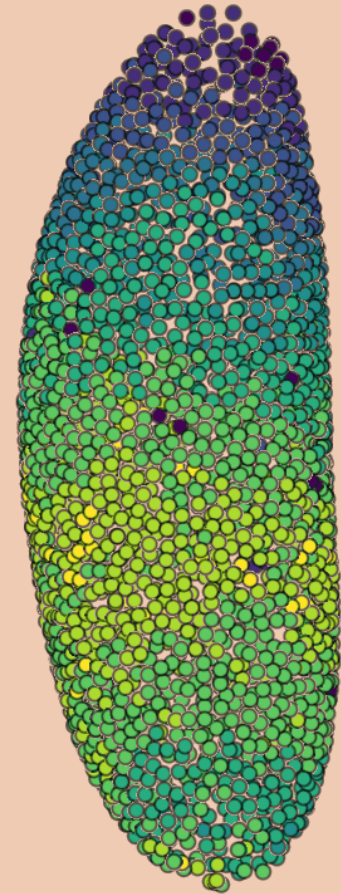


# Mitotic waves are an embryonic-scale phenomenon that takes place at the per-nucleus level



# How do mitotic waves direct *Drosophila* blastoderm formation?

- I. What is the trajectory by which mitotic waves traverse the *Drosophila* embryo?
- II. How do mitotic waves affect the final packing of nuclei?
- III. How do the waves respond to genetic perturbation?

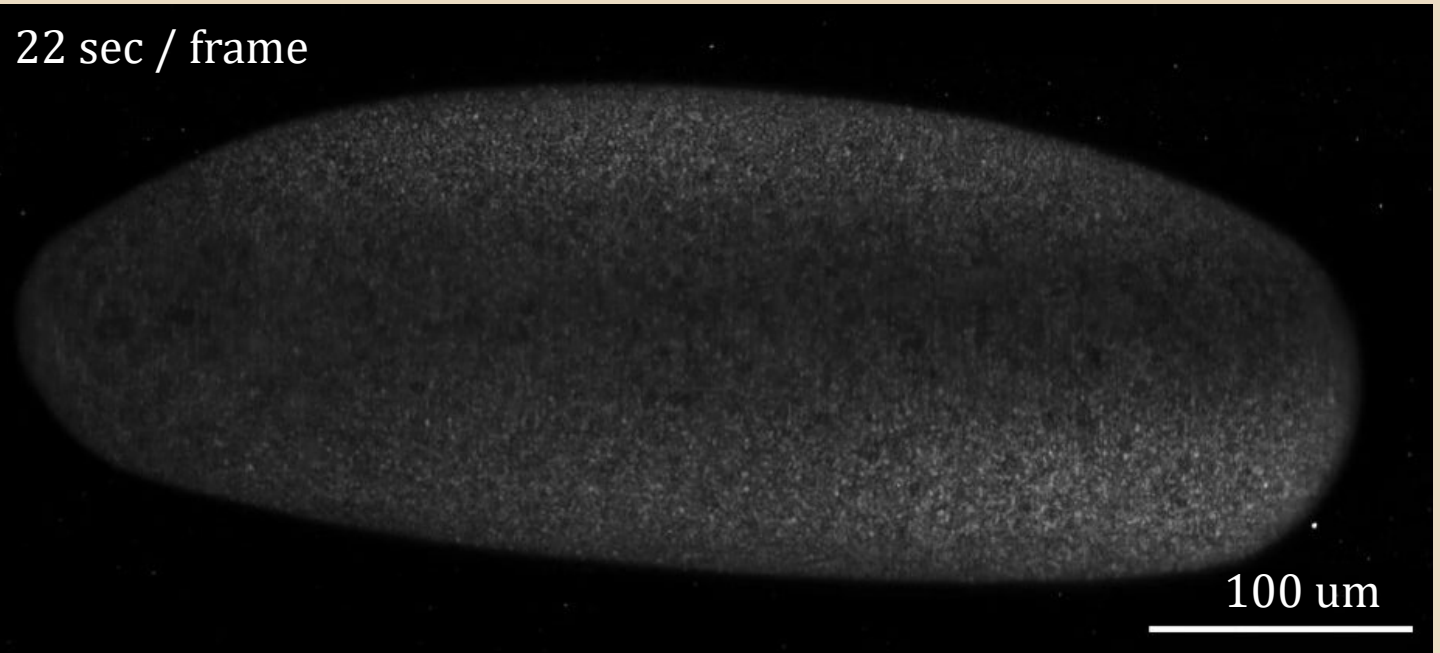


*Division times*  
(darker = earlier)

Light sheet imaging  
produces highly  
resolved, isotropic,  
*in toto* data

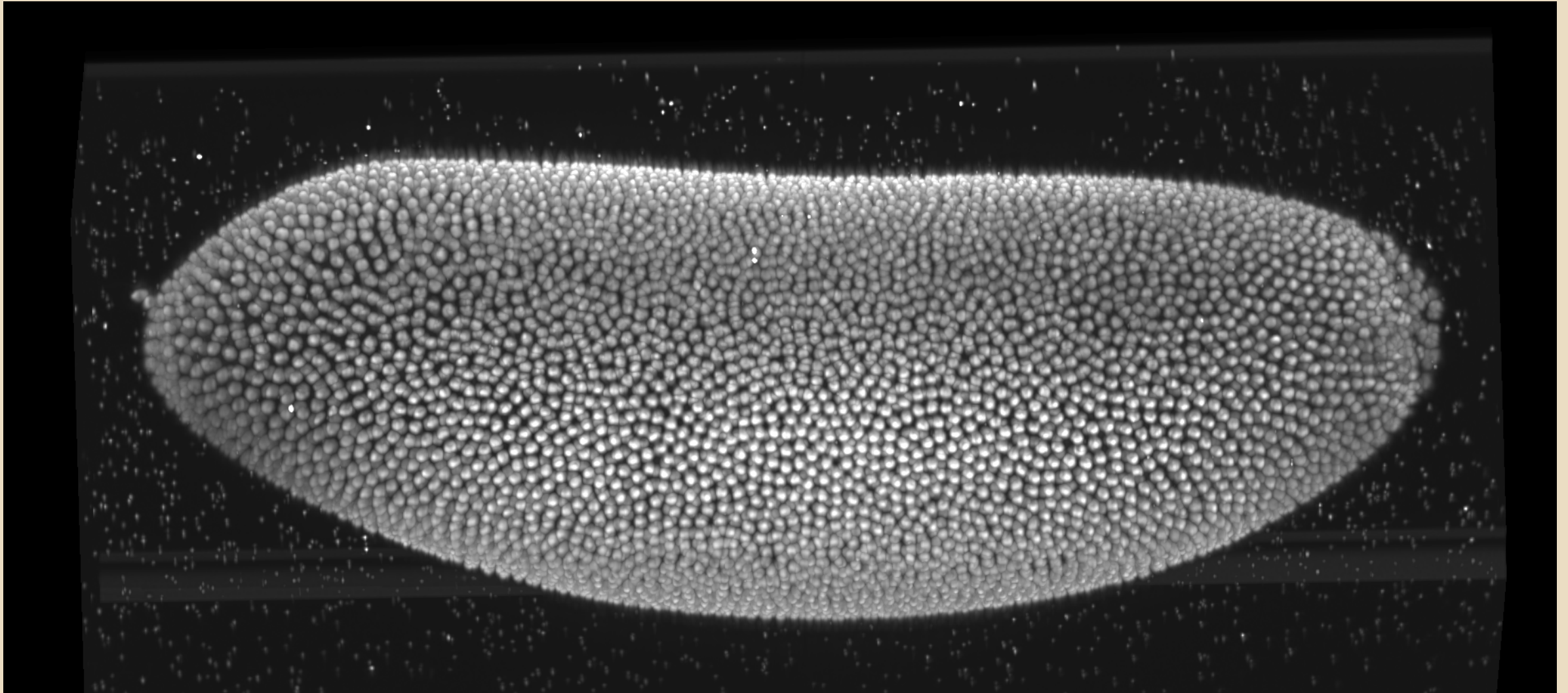


Liu Yang



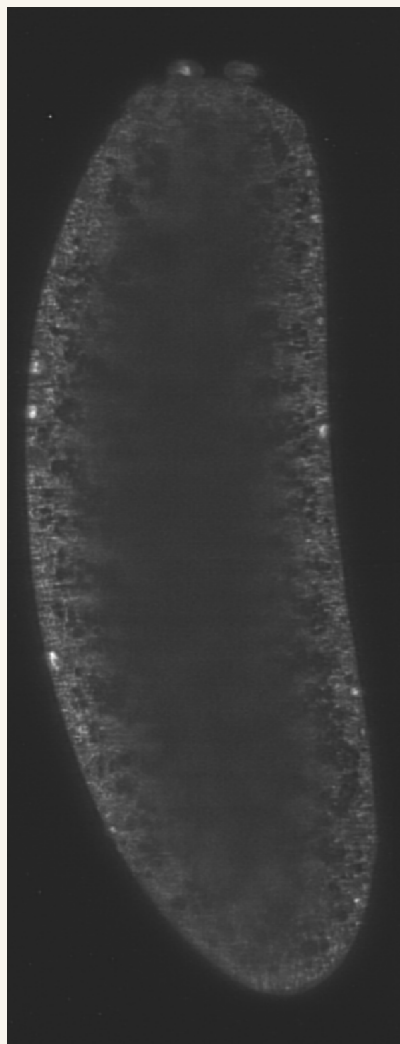
- Light sheet microscopy
  - Two cameras x two angles per time point = 4 views
  - 20-22 seconds / time point (about as fast as possible)
- Embryos expressing h2a-GFP
  - Start when nuclei first visible (NC8) through NC14
  - Cooled to 18 C to slow down development

**Raw data (final frame)**

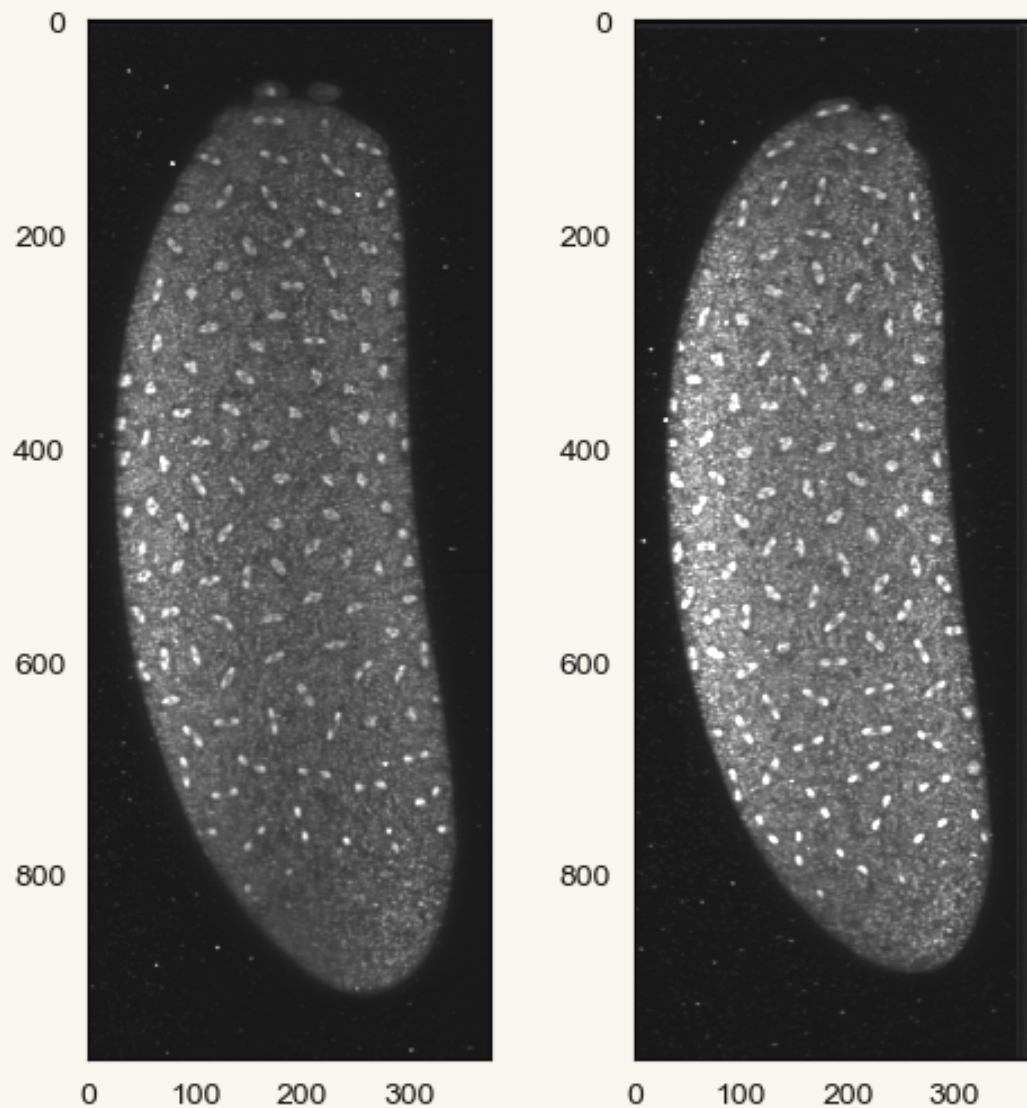




Single slice



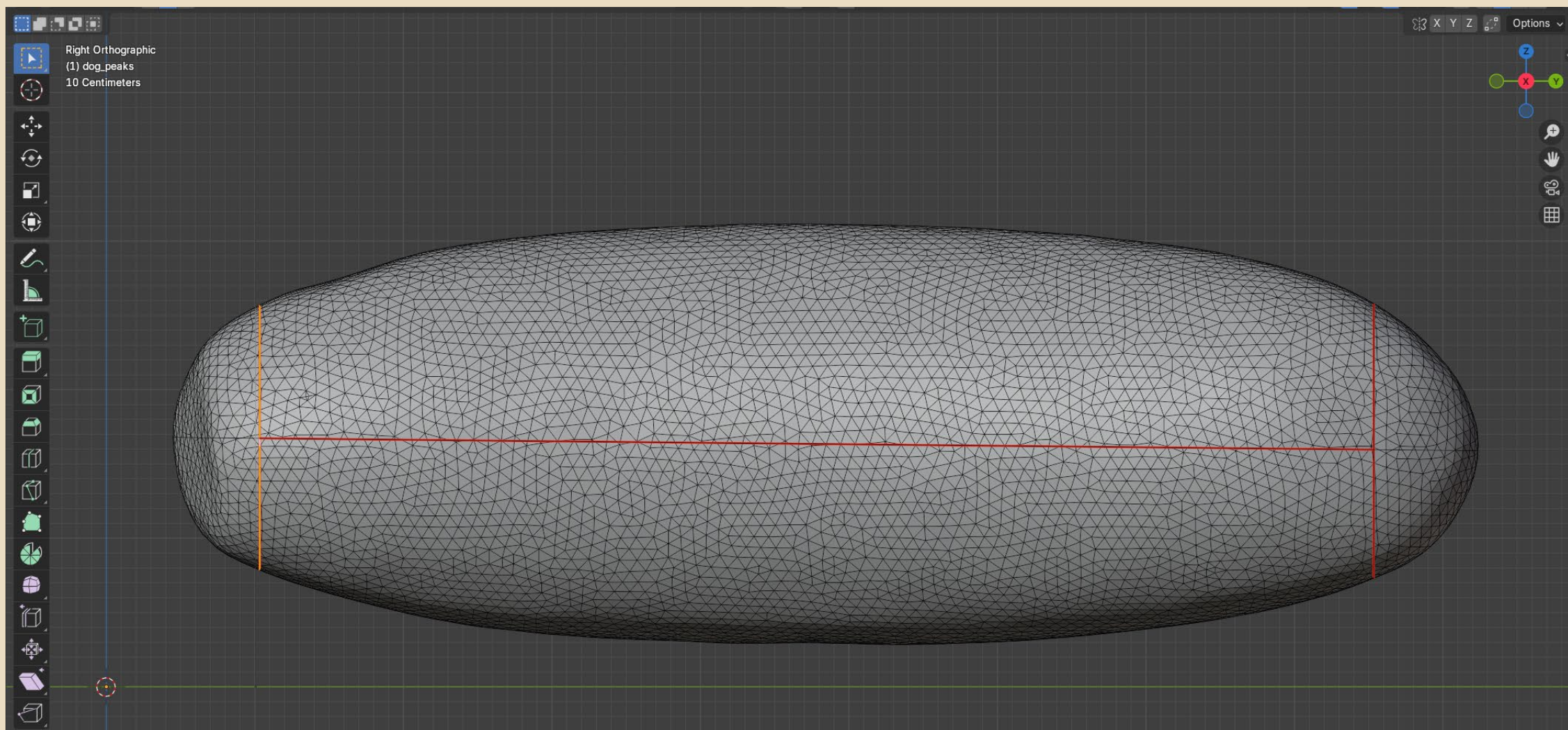
Max projected



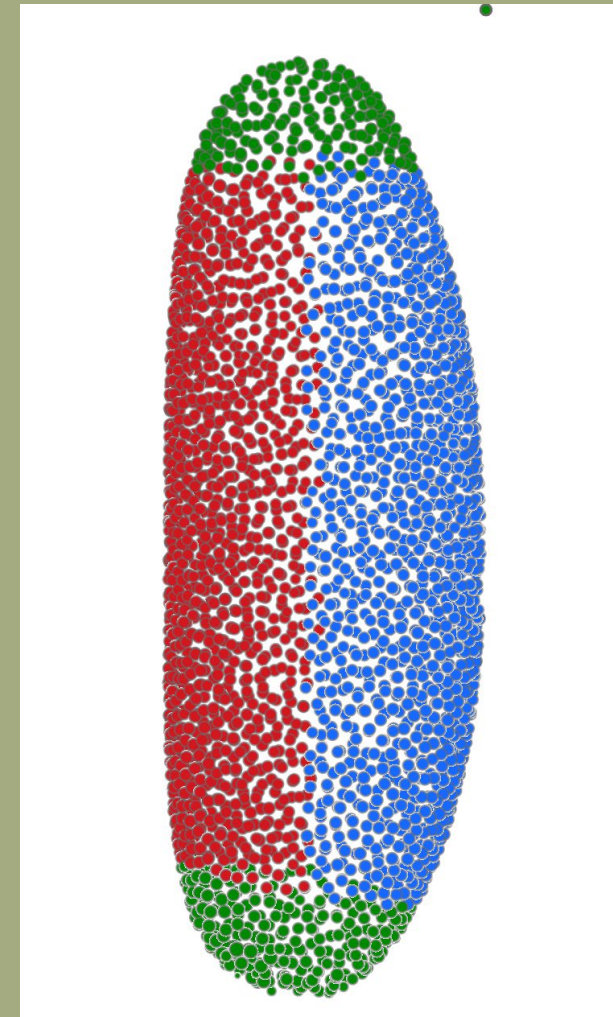
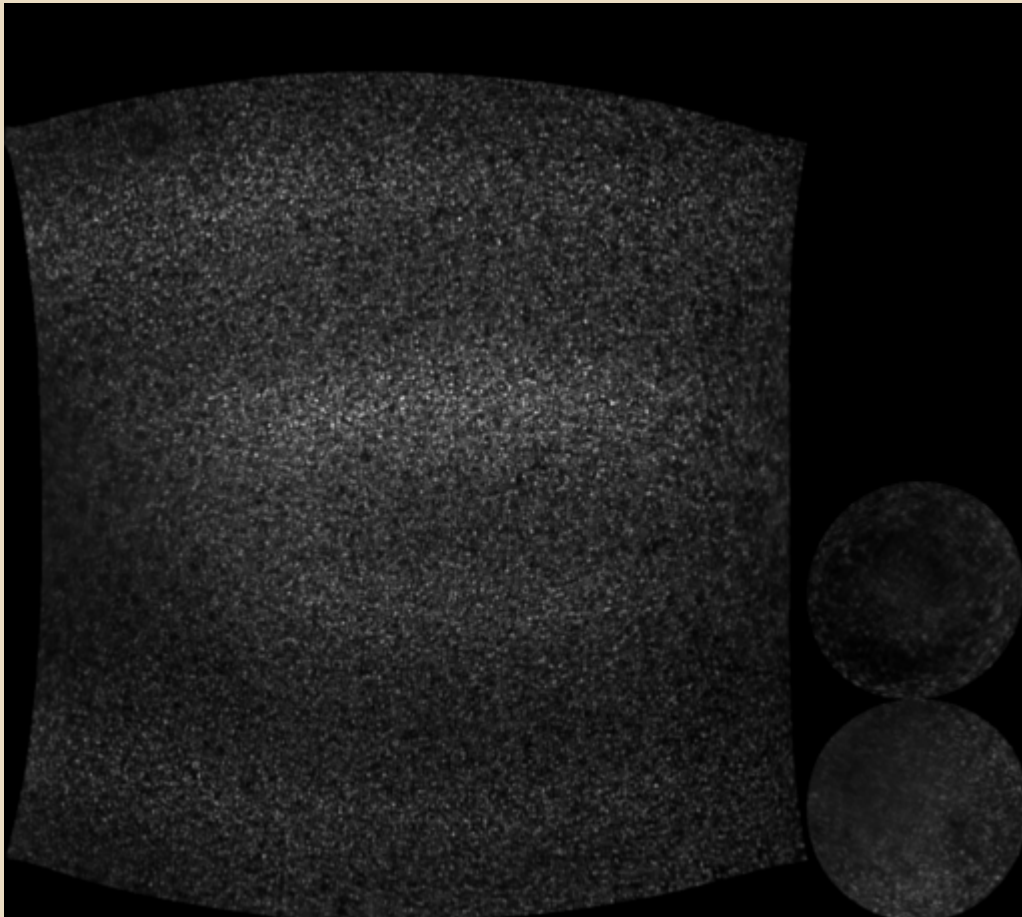
**Choosing the  
best data  
transformation**



## Embryos are converted to meshes (blender)

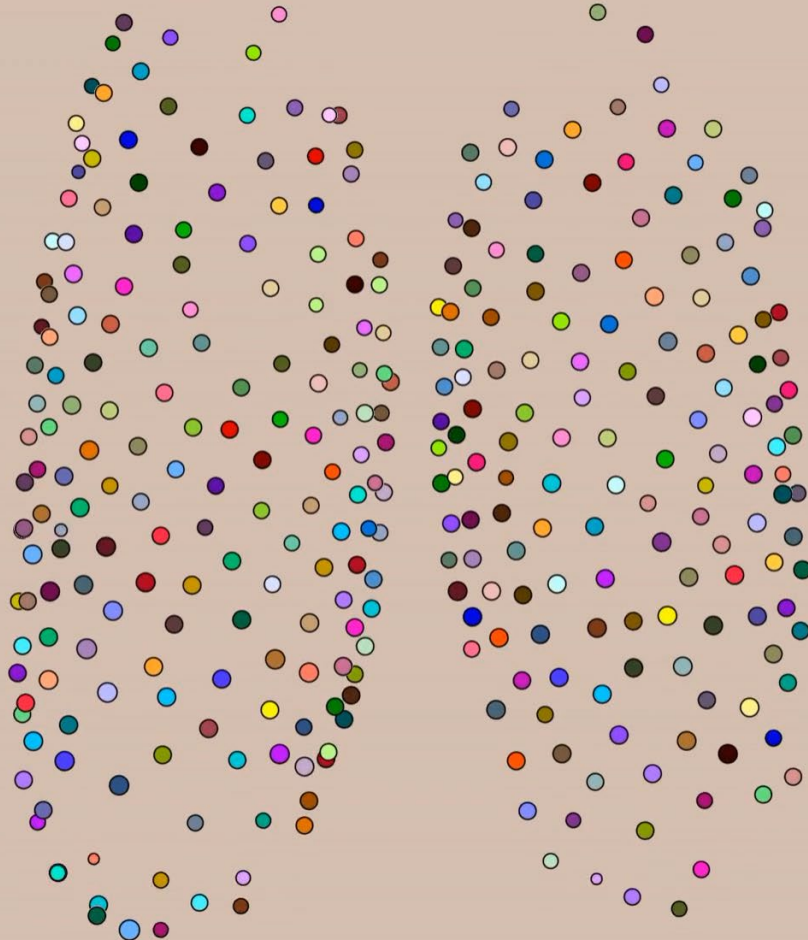


# Segmentations are carried out in 2d using Cellpose-SAM





# Nuclei are tracked via a two-step algorithm



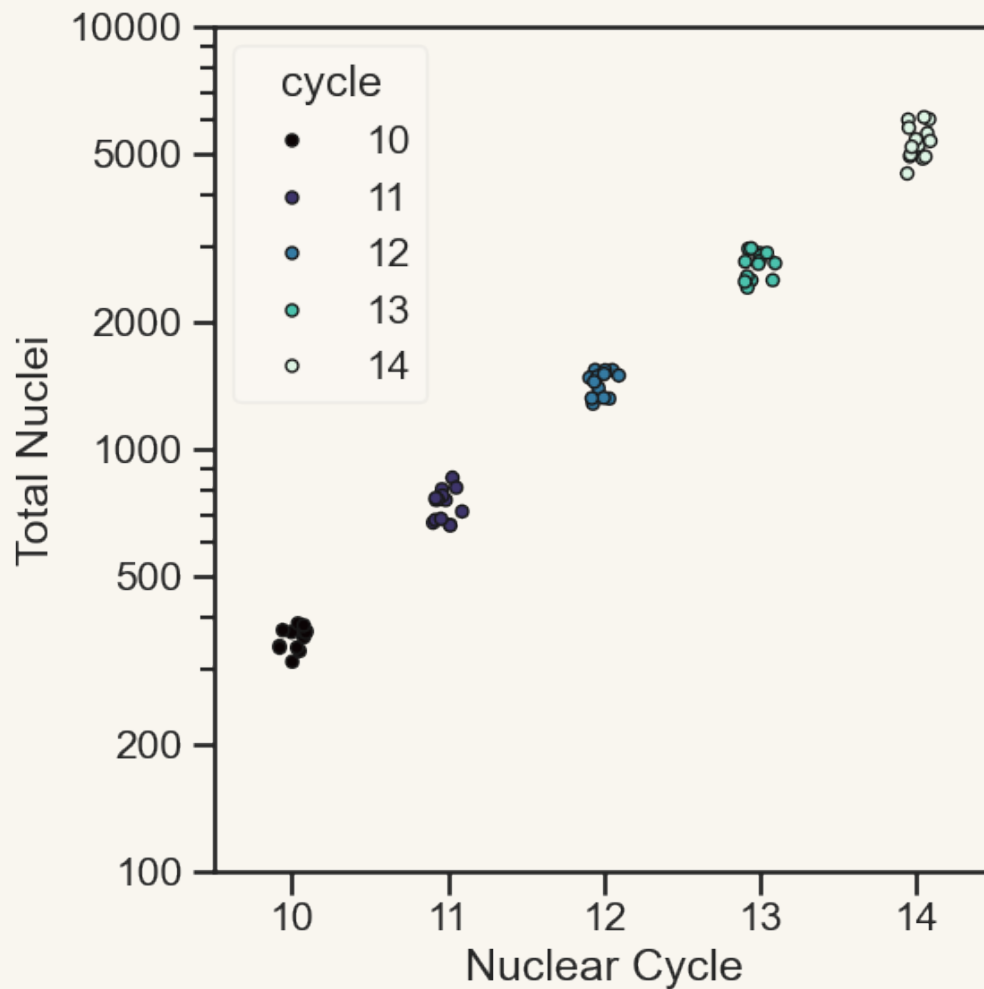
## Step 1: Tracking without divisions

- LAP tracking using 3D coordinates (implemented by Trackmate)

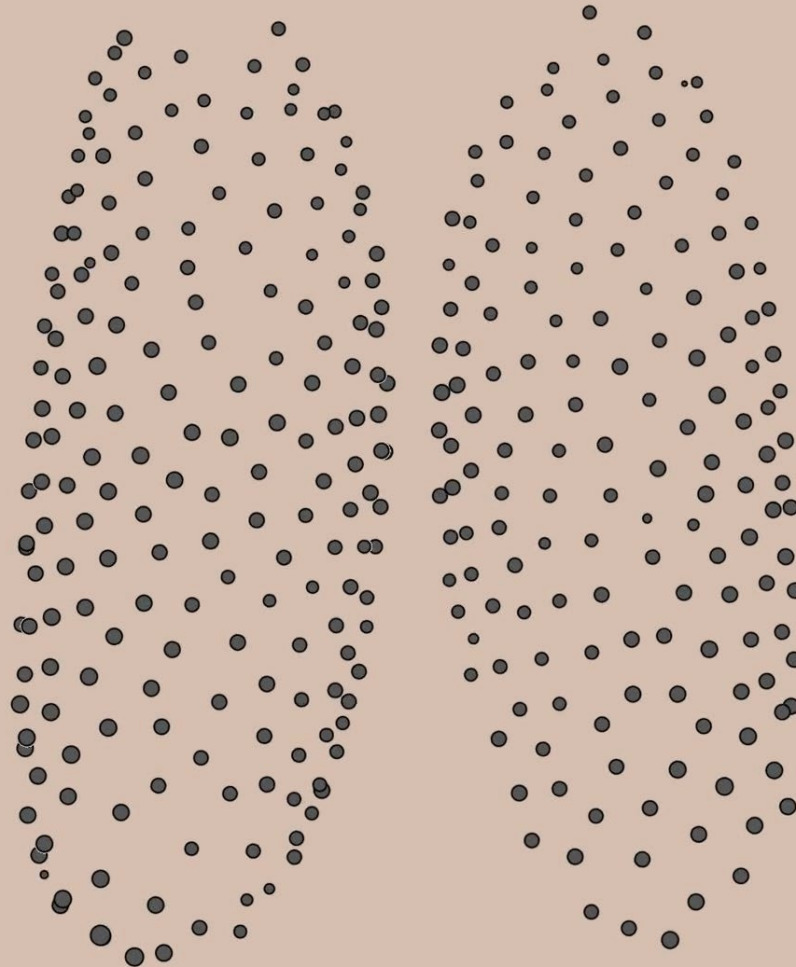
## Step 2: Assign daughter nuclei to parents

- Split the problem up into one problem per M phase
- Assume that daughter nuclei move in opposite directions during anaphase

# Thousands of nuclei are tracked per embryo



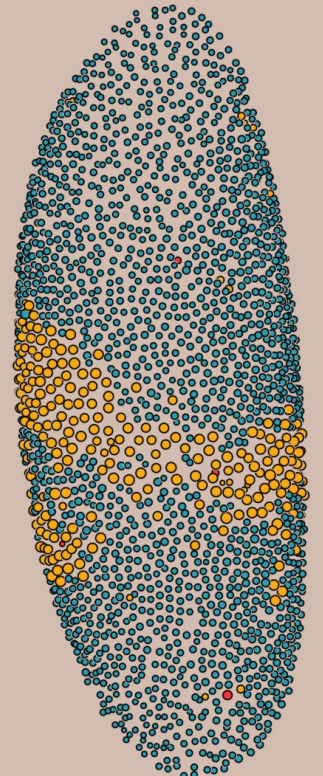
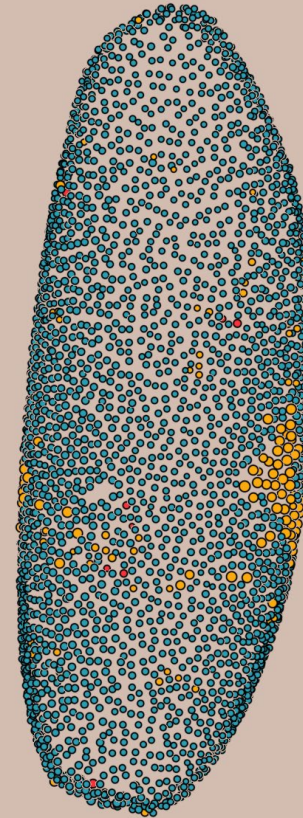
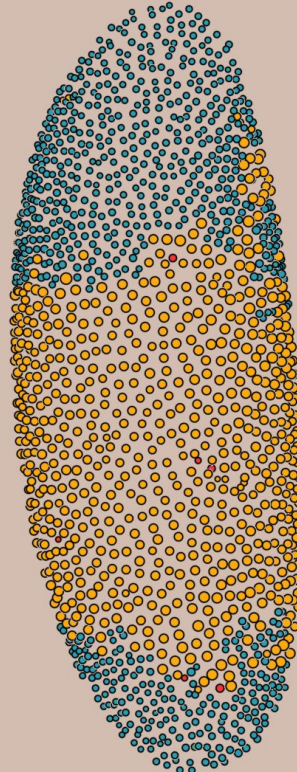
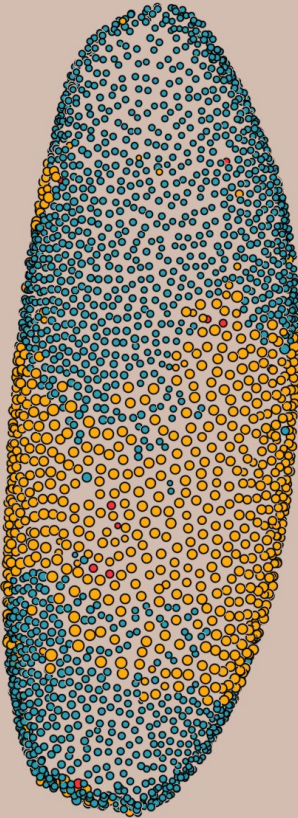
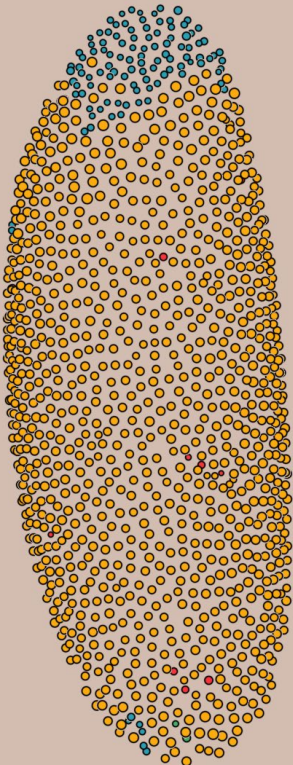
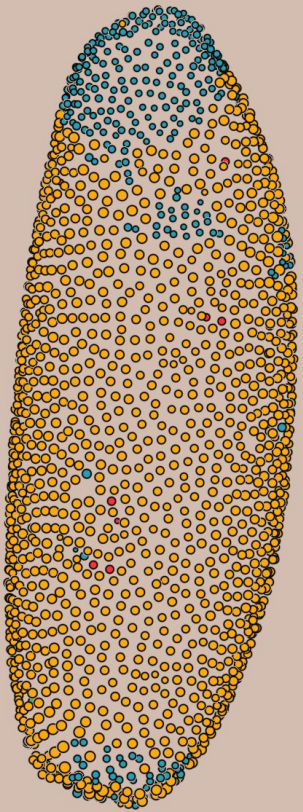
# Individual nuclear divisions can be tracked across the embryo



Gray: NC 10  
Green: NC 11  
Red: NC 12  
Yellow: NC 13  
Blue: NC 14

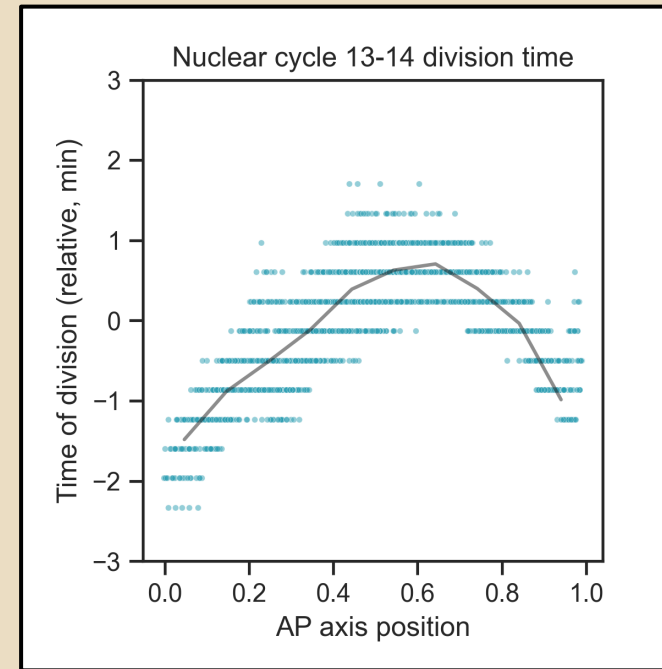
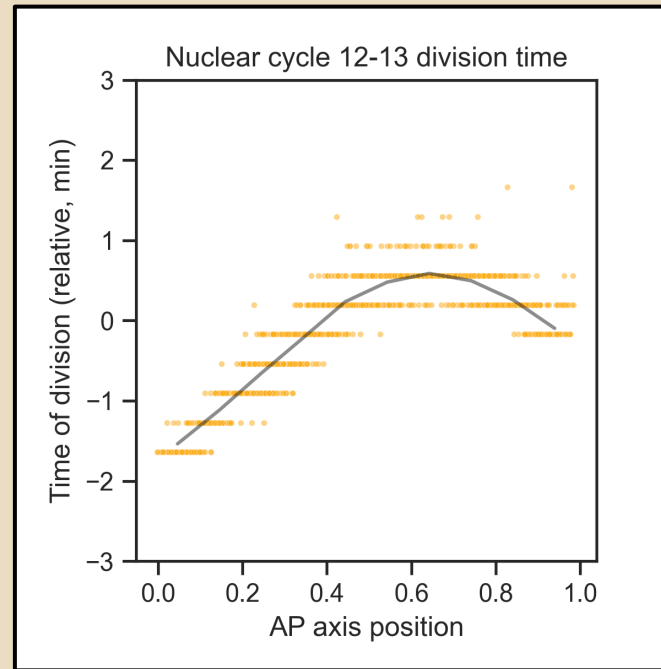
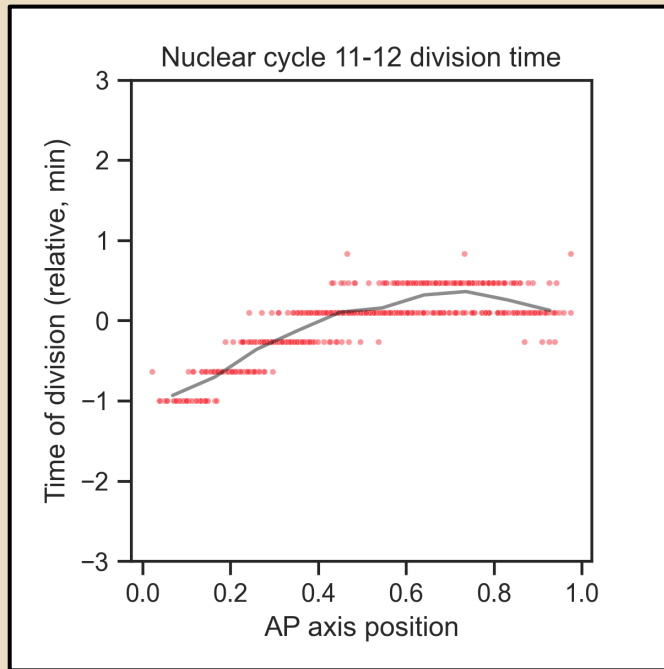


# Individual nuclear divisions can be tracked across the embryo

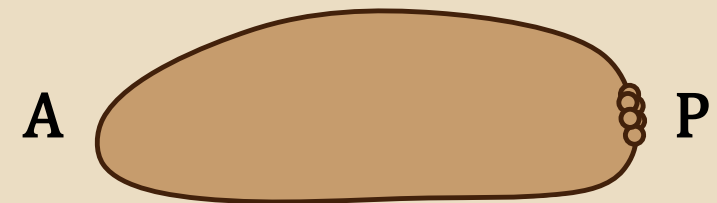


Yellow: NC 13 (pre-division)  
Blue: NC 14 (post-division)

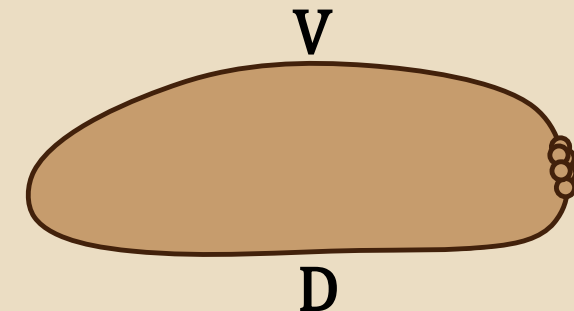
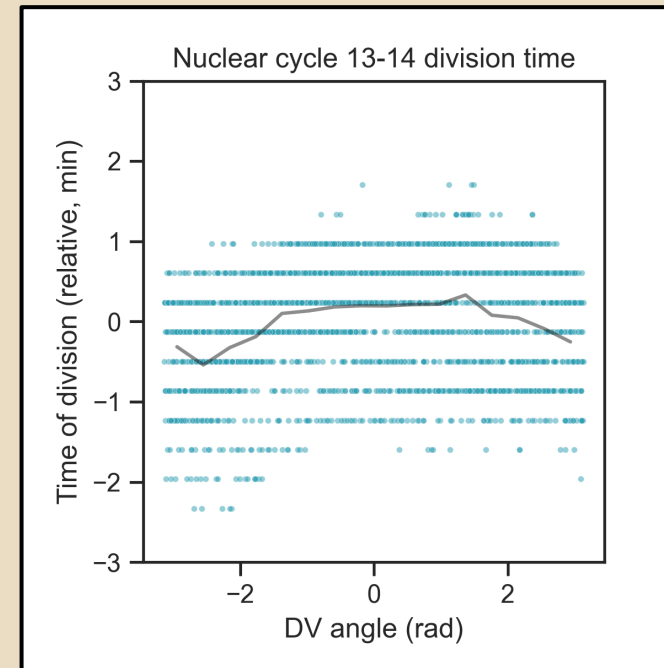
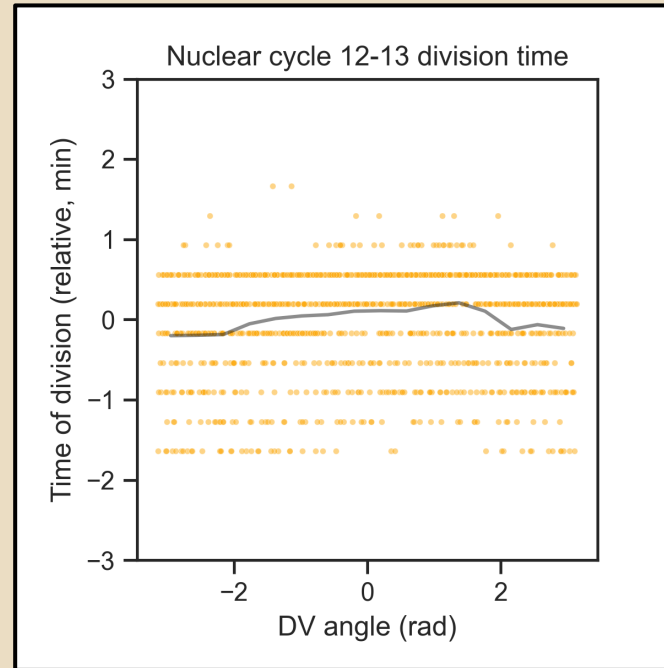
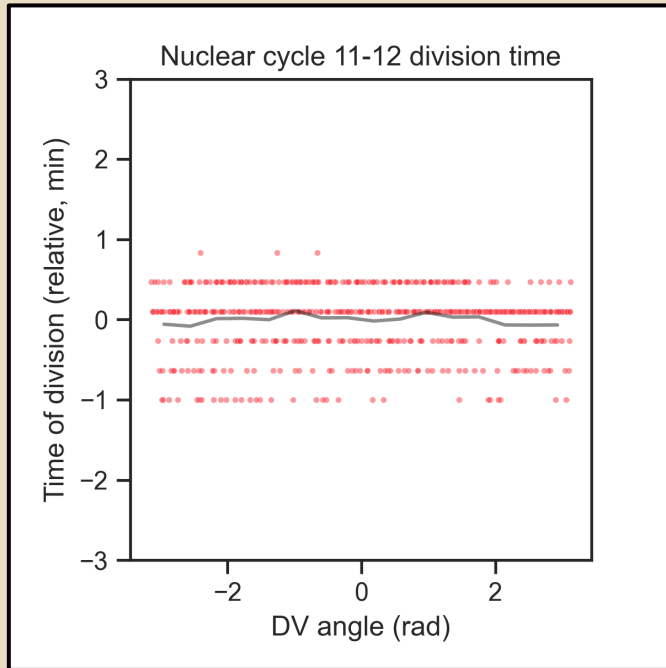
# Nuclear division times vary across the AP axis



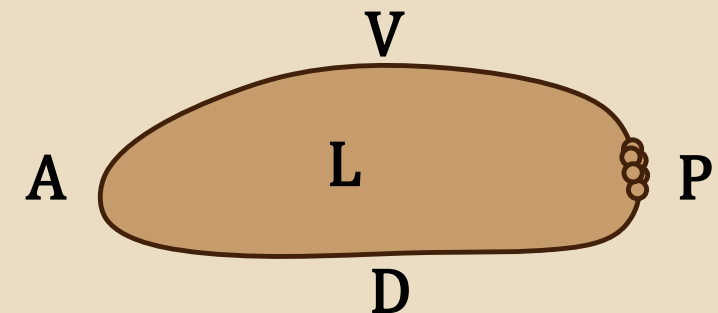
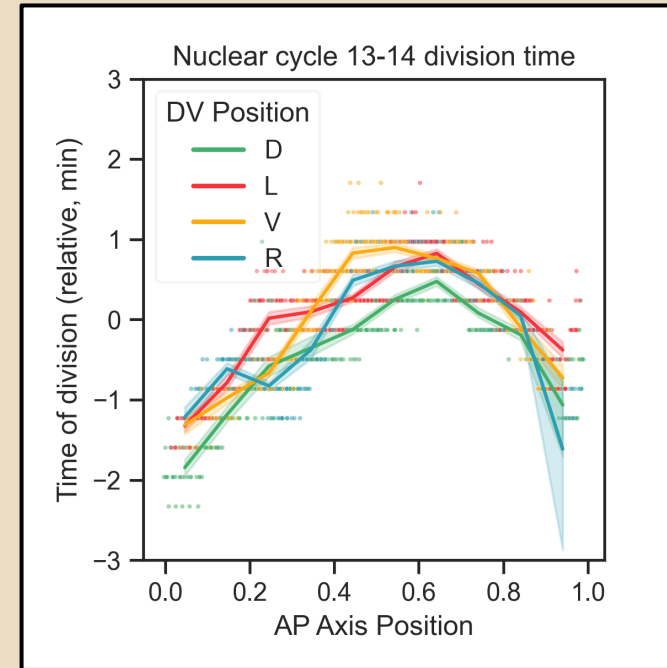
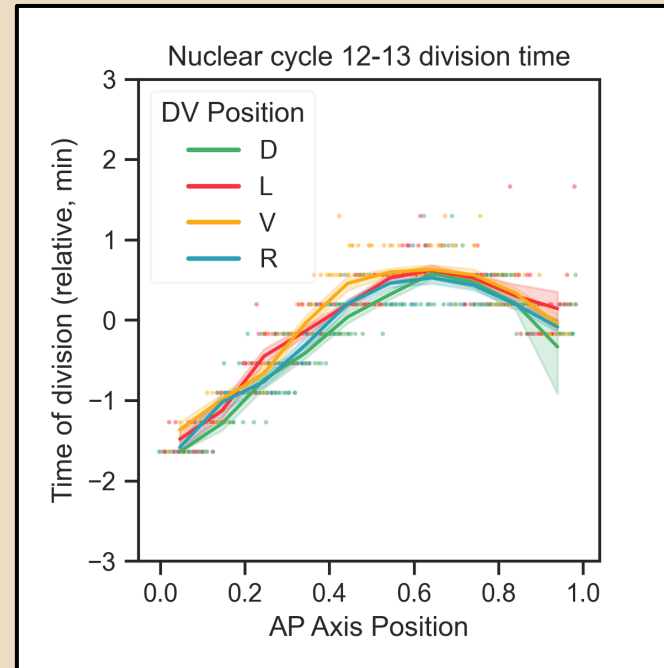
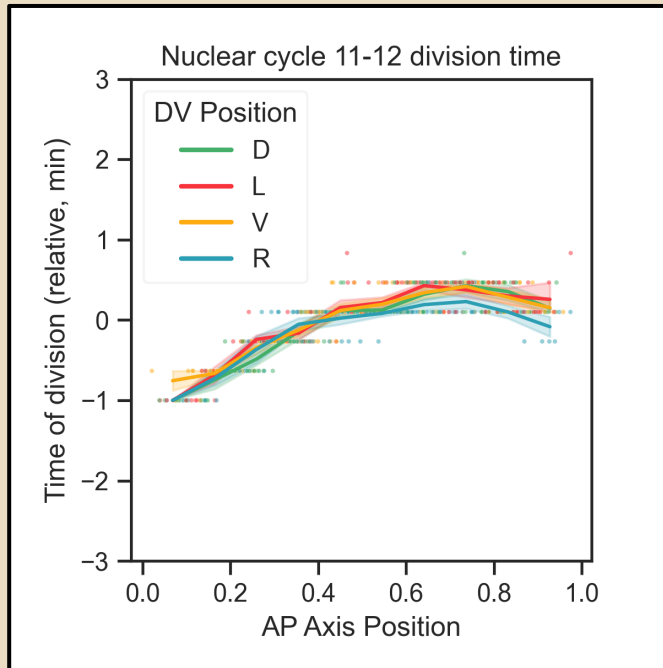
Each dot: one nucleus dividing



# Nuclear division times vary less across the DV axis

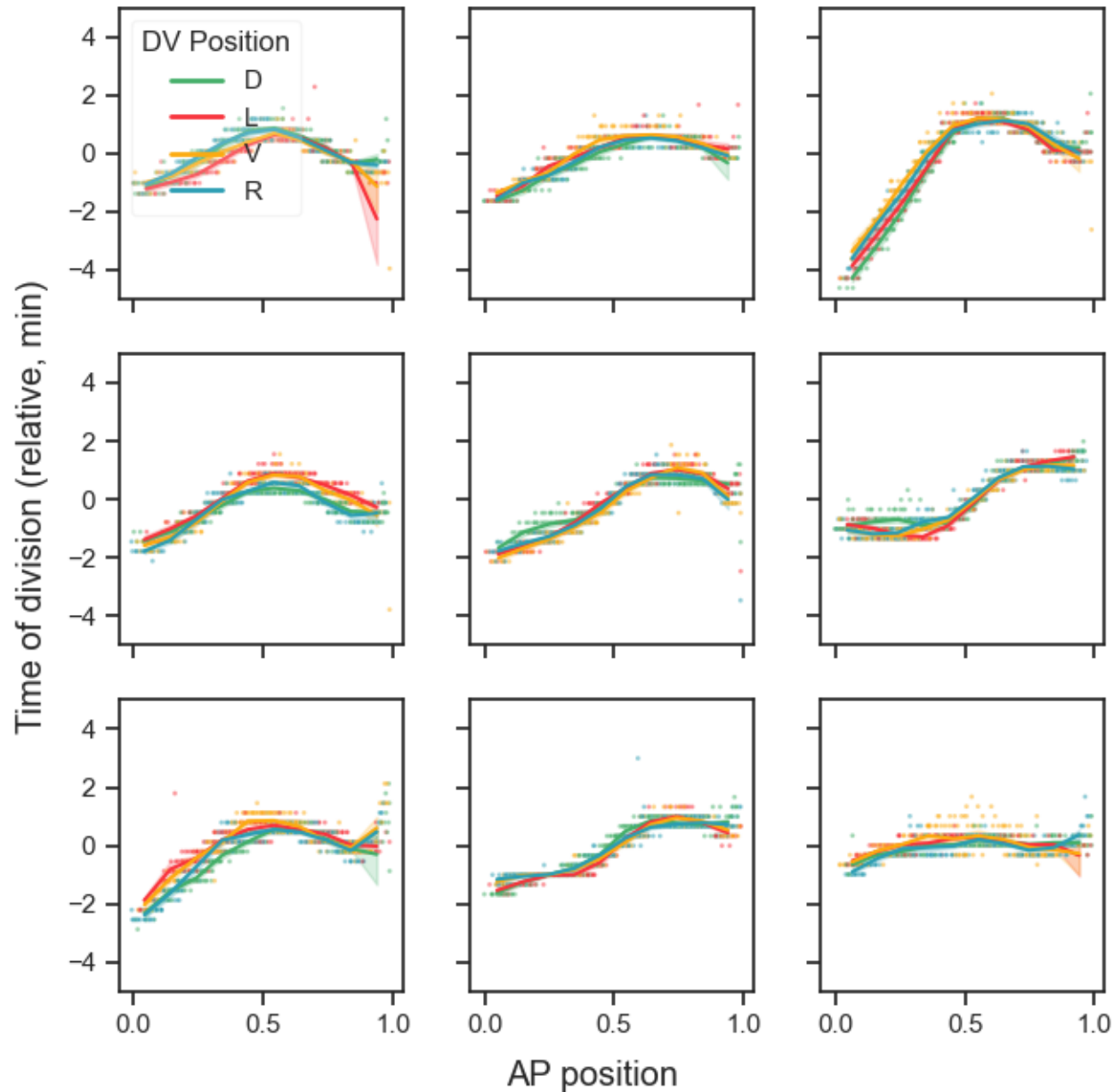


# All sides of the embryo show similar mitotic waves

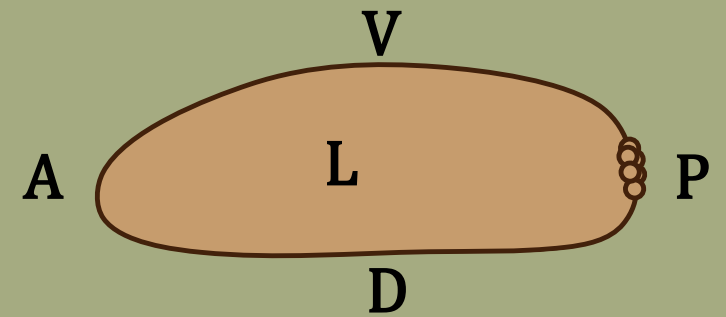




### NC12 Division



**This strong AP axis  
relationship holds over  
many embryos**





# Properties of mitotic waves

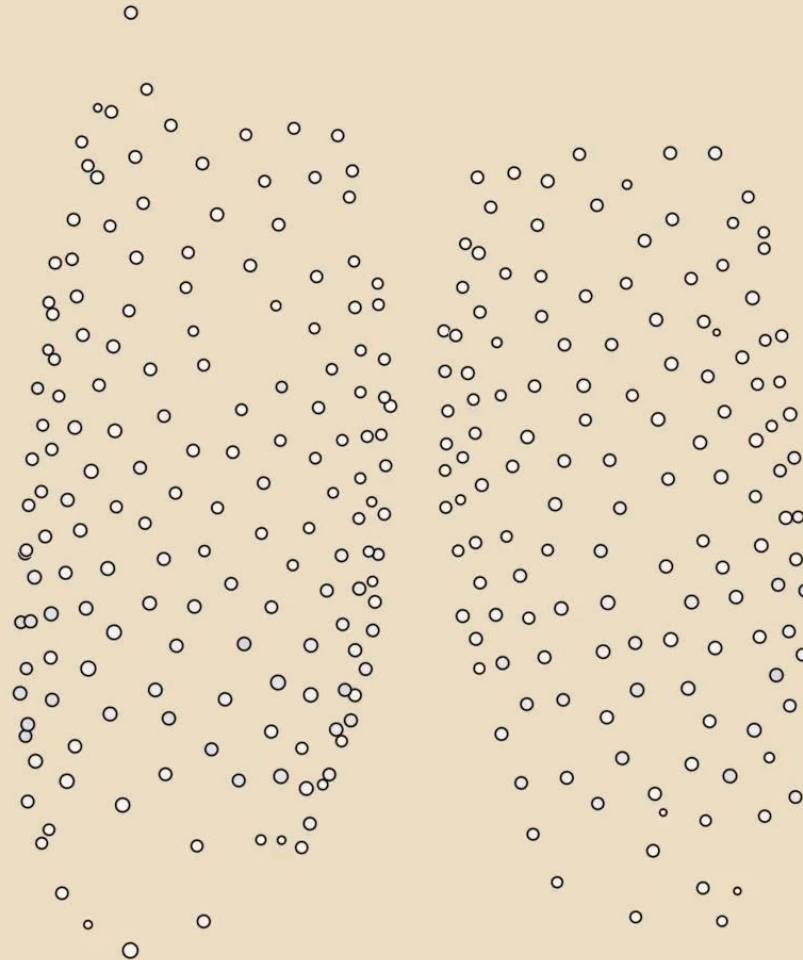
- I. Mitotic waves traverse the embryo primarily along the AP axis.**
- II. Mitotic waves begin at the poles and move towards the center of the embryo**

# Nuclei spread out along the AP axis over divisions

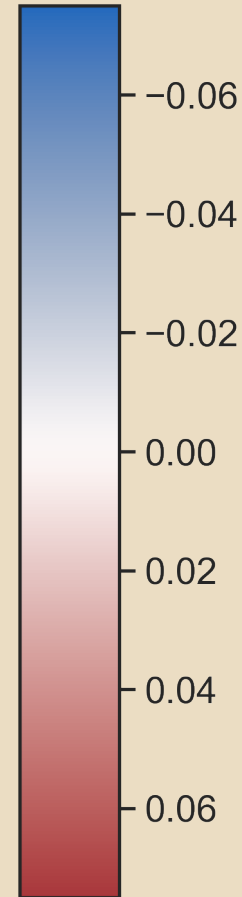
Anterior



Posterior

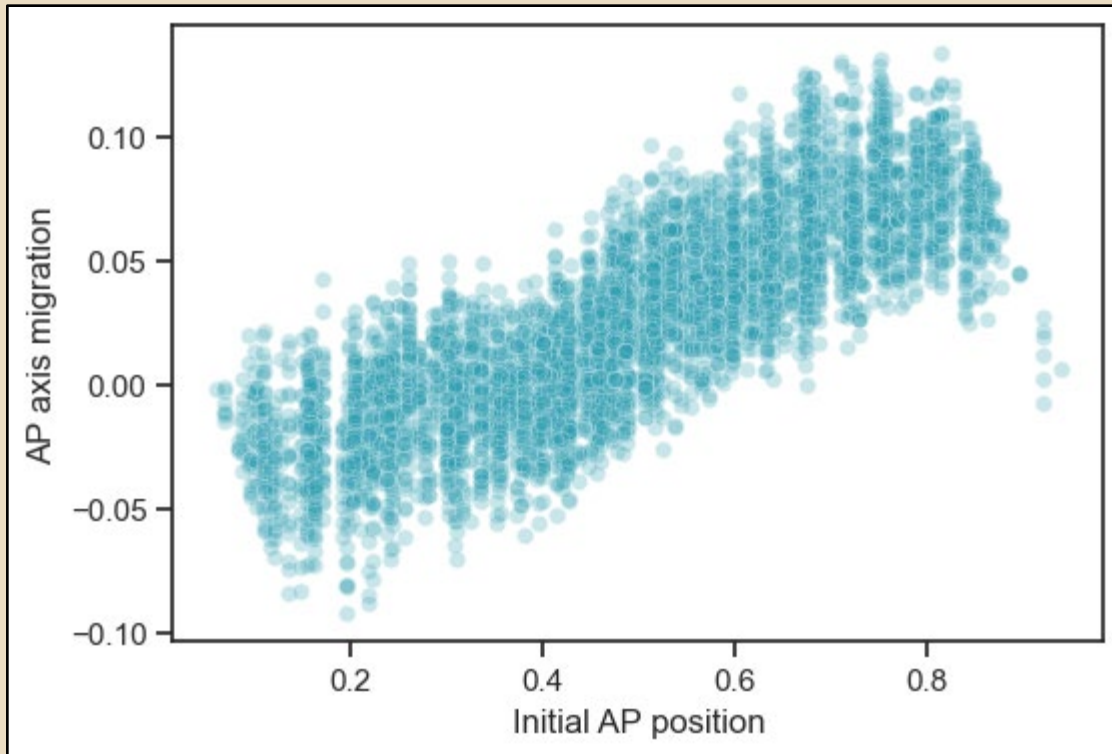


Net AP axis  
movement since start

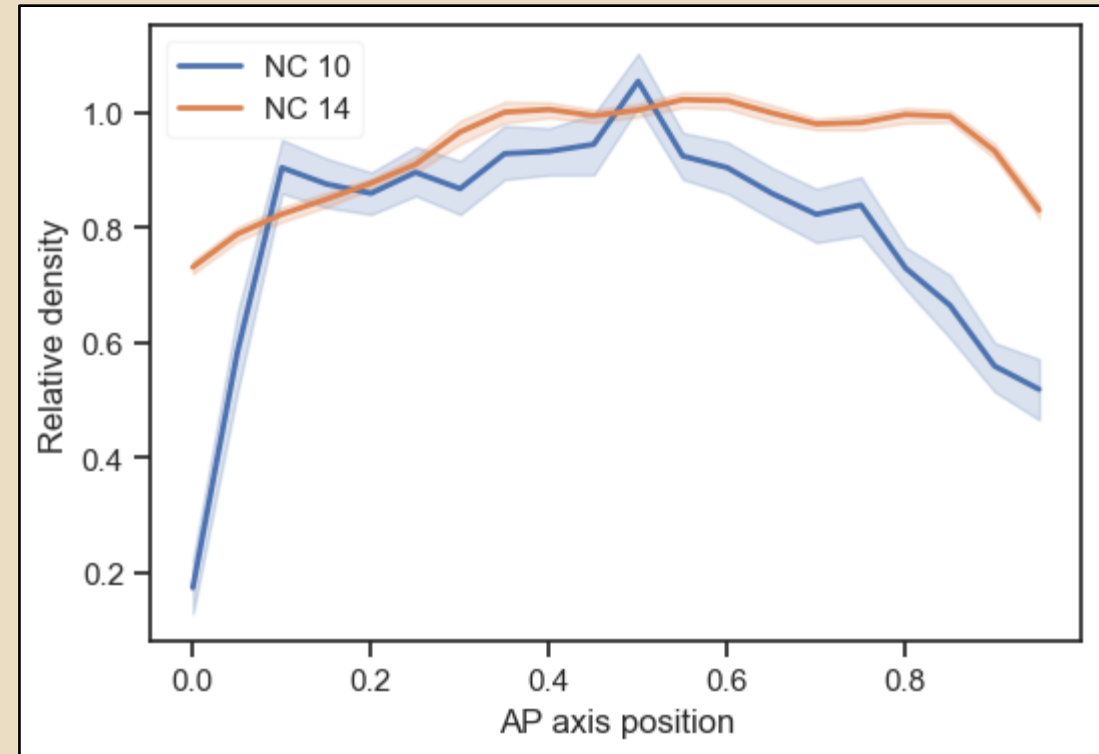


# Nuclear spreading generates a more uniform density across the embryo

Initial position vs. net migration

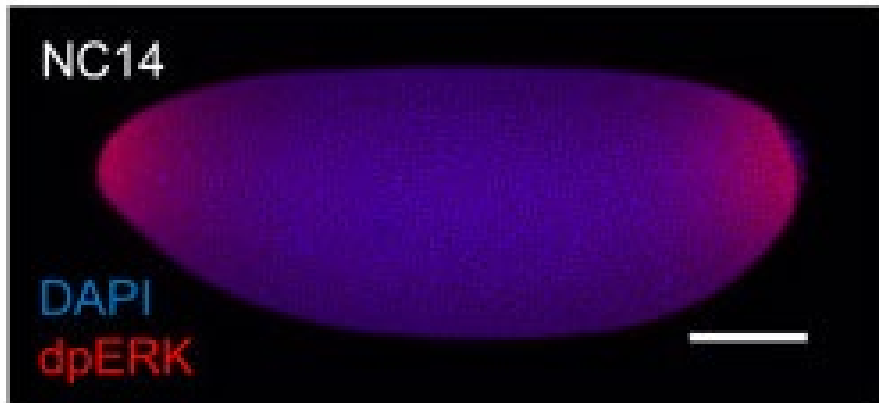


Initial vs. final density profile



# ERK activity influences timing of mitotic entry

Erk is active at the poles of the embryo



## Developmental Cell

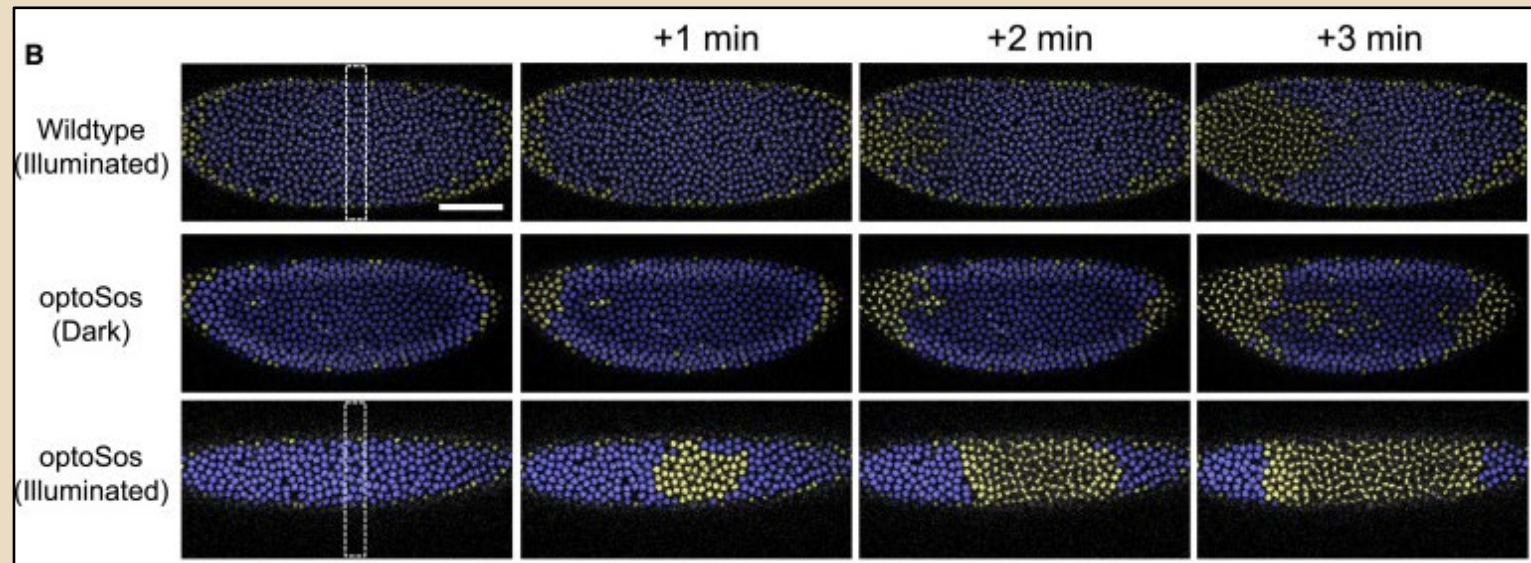


Volume 59, Issue 23, 2 December 2024, Pages 3061-3071.e6

Short article

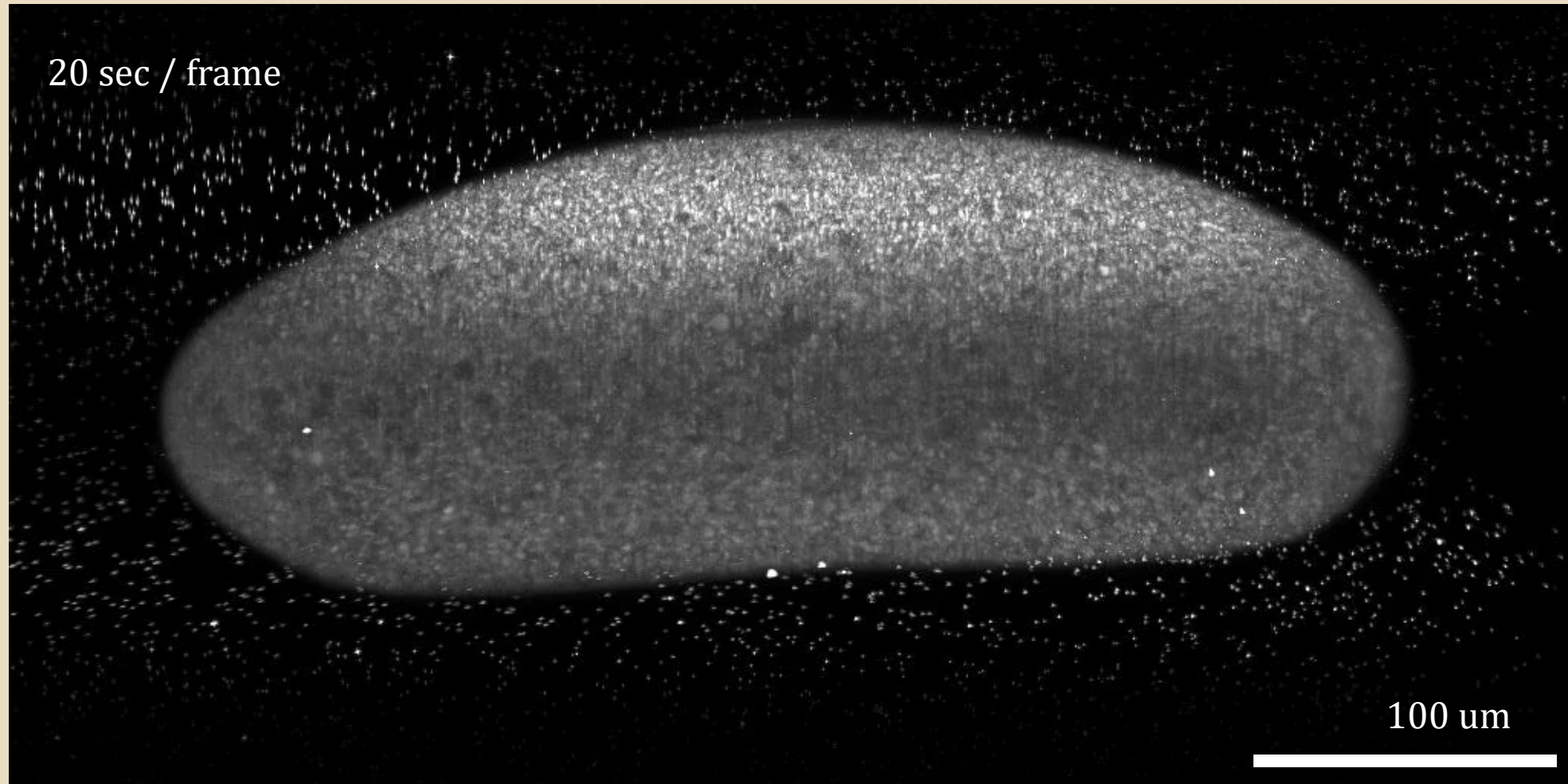
### ERK synchronizes embryonic cleavages in *Drosophila*

Liu Yang<sup>1,8</sup>, Audrey Zhu<sup>1,2,8</sup>, Javed M. Aman<sup>1,3</sup>, David Denberg<sup>1,4</sup>, Marcus D. Kilwein<sup>5</sup>, Robert A. Marmion<sup>1</sup>, Alex N.T. Johnson<sup>1,2,5</sup>, Alexey Veraksa<sup>6</sup>, Mona Singh<sup>1,3</sup>, Martin Wühr<sup>1,2,5</sup> ✉, Stanislav Y. Shvartsman<sup>1,4,5,7,9</sup> ✉

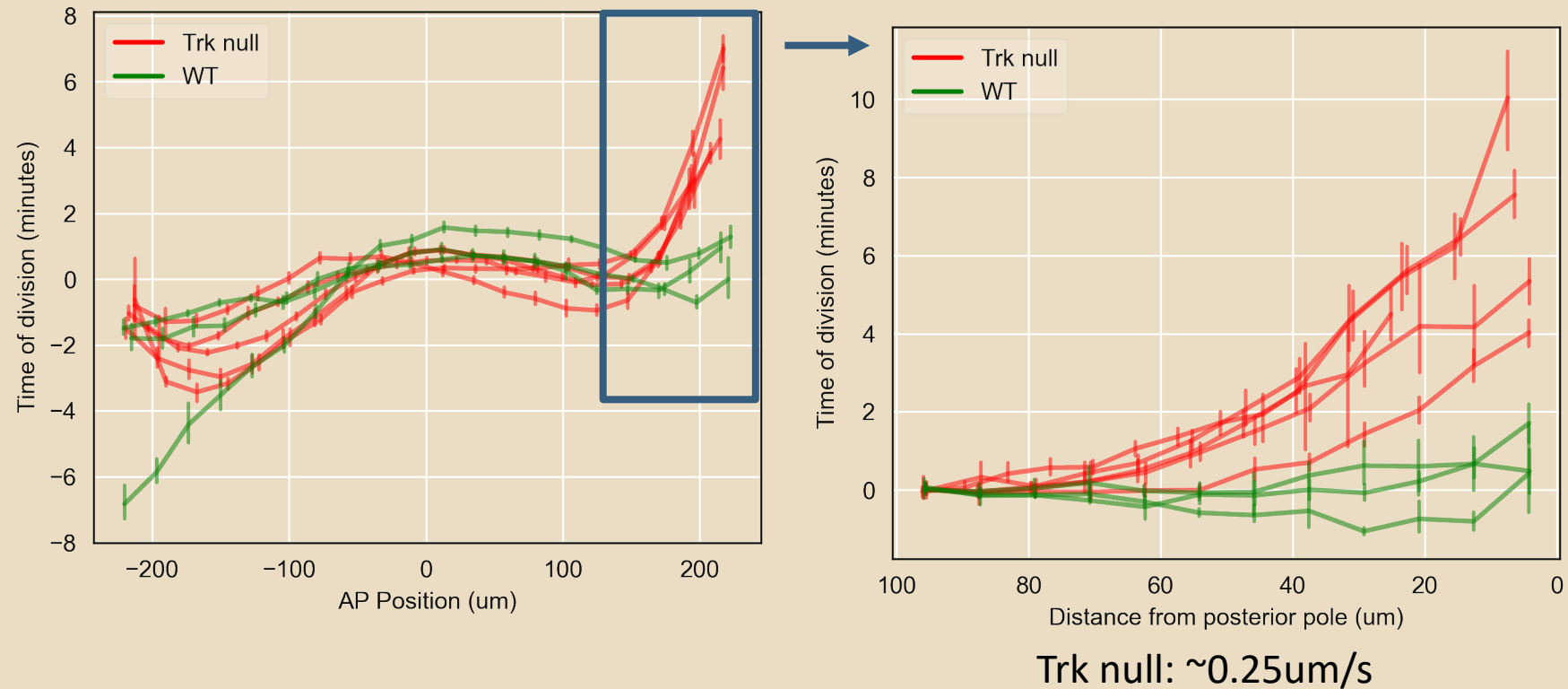




**In Trk null embryos, mitotic waves travel slowly to the posterior**

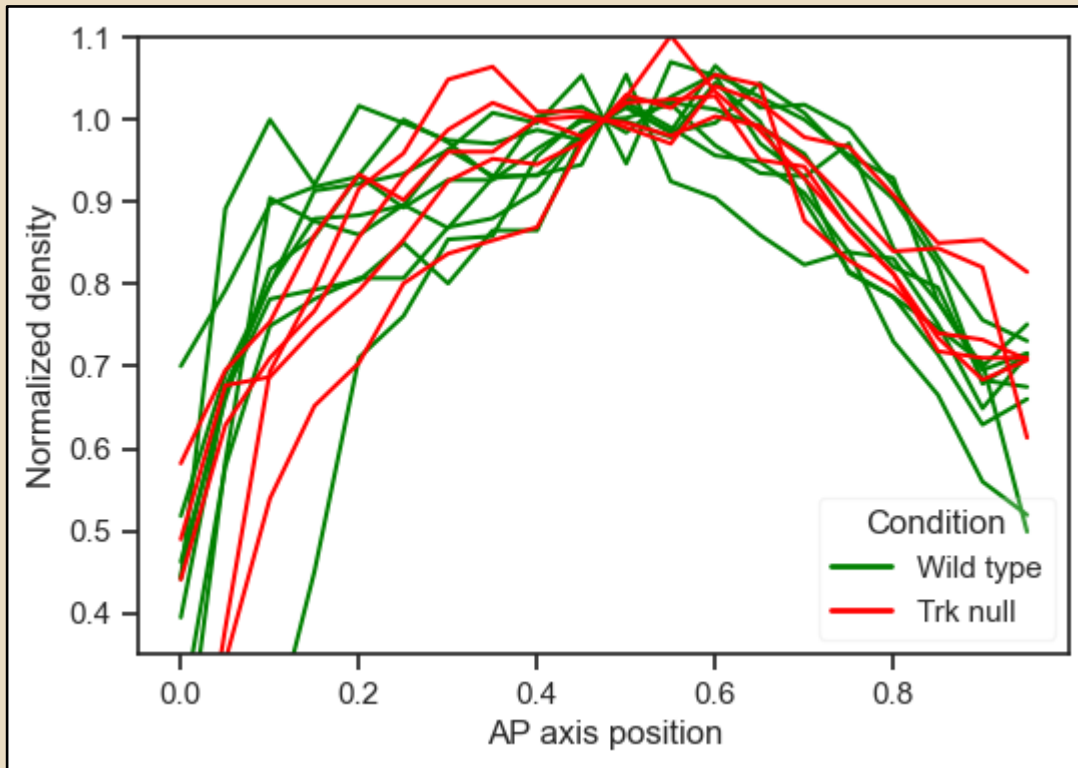


# In Trk null embryos, mitotic waves travel slowly to the posterior

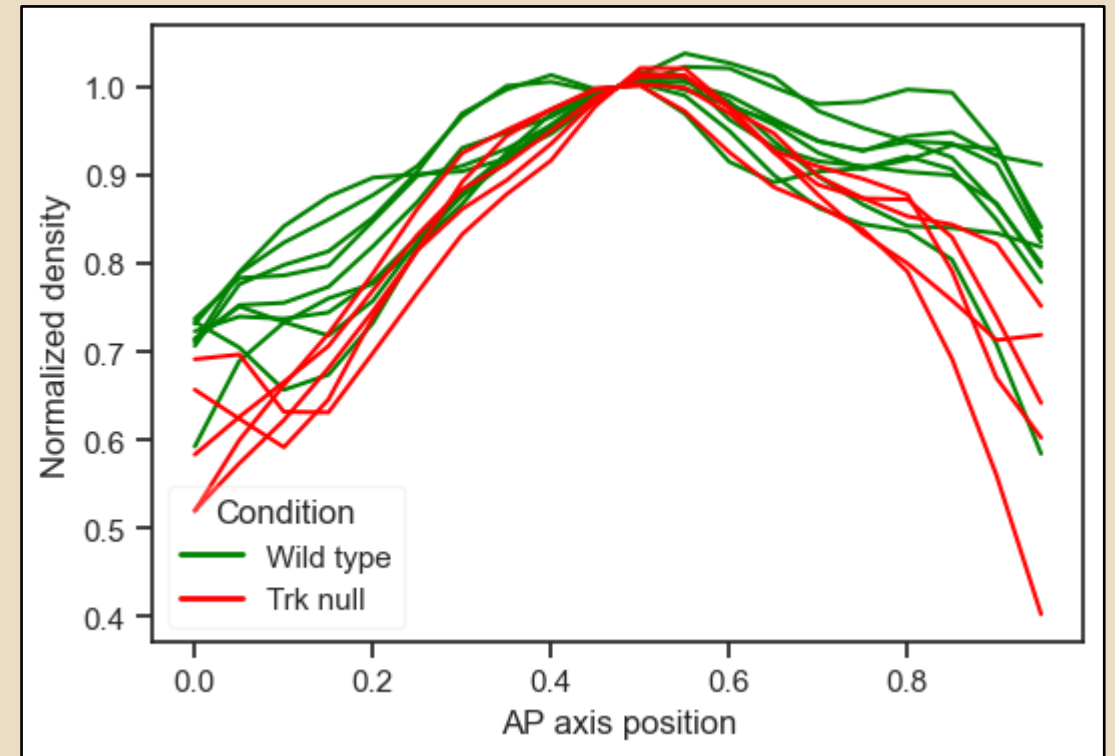


# Trk null embryos produce a less uniform distribution of nuclei

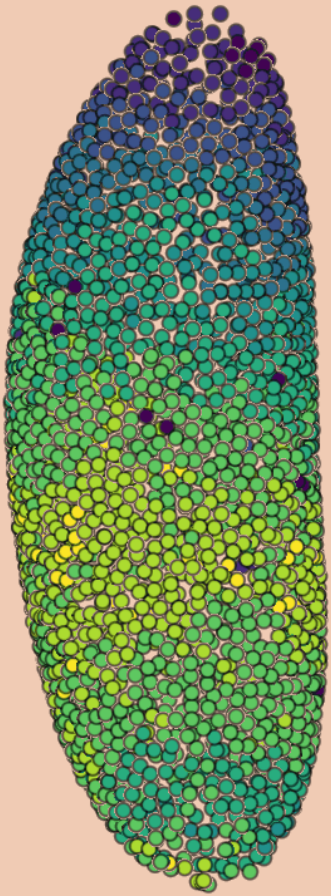
Initial Density Profiles



Final Density Profiles



# Conclusions



- I. Individual nuclear tracking enables precise description of blastoderm formation**
- II. Mitotic waves produce thousands of new nuclei while simultaneously organizing density at an embryo scale**



# Acknowledgements

Liu Yang  
Hayden Nunley  
Stanley Nicholson  
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Stas Shvartsman  
Jared Toettcher

