

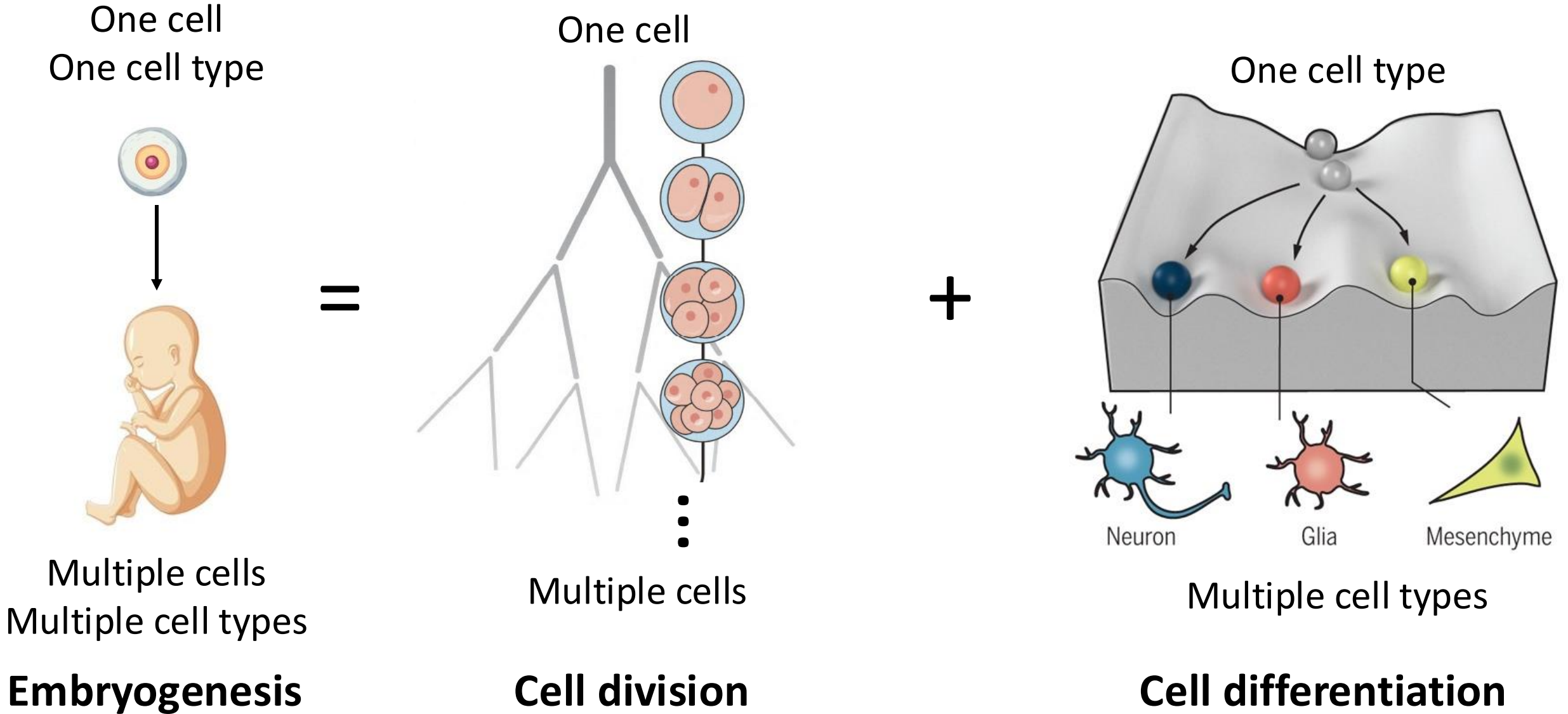
# Inferring cell lineage trees and fate maps from lineage tracing data

Palash Sashittal

Department of Computer Science

Virginia Tech

# Organismal Development

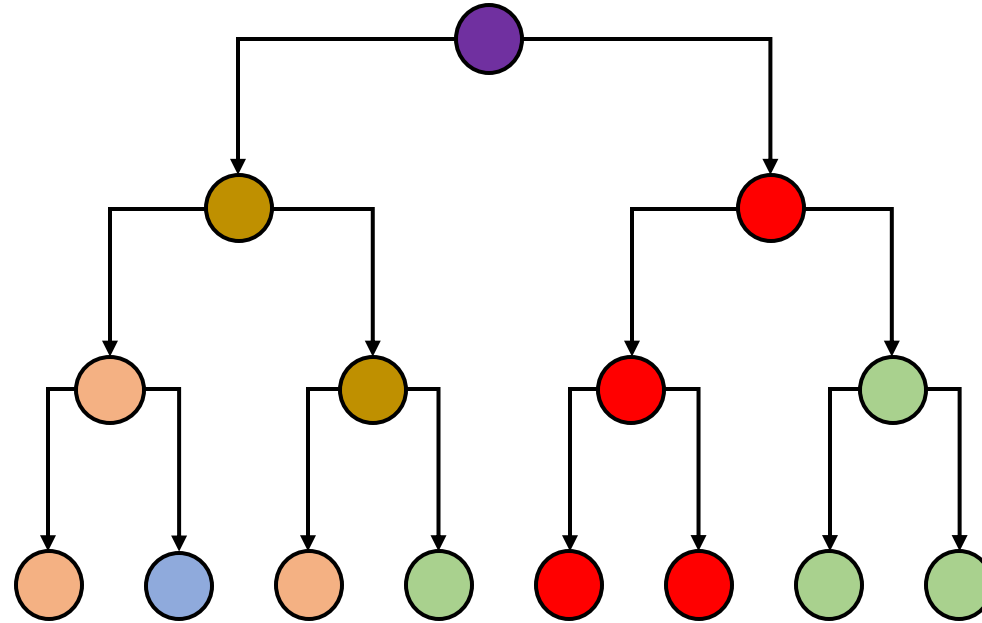


# Organismal Development

One cell  
One cell type

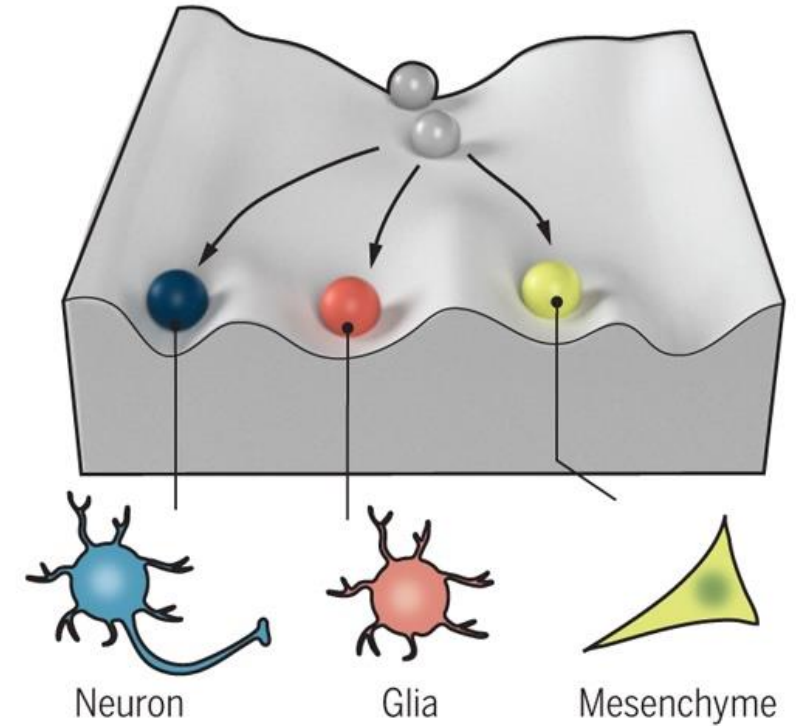


Multiple cells  
Multiple cell types  
**Embryogenesis**



**Cell lineage tree  $T$**   
Rooted tree with leaves  
representing cells in the organism

One cell type



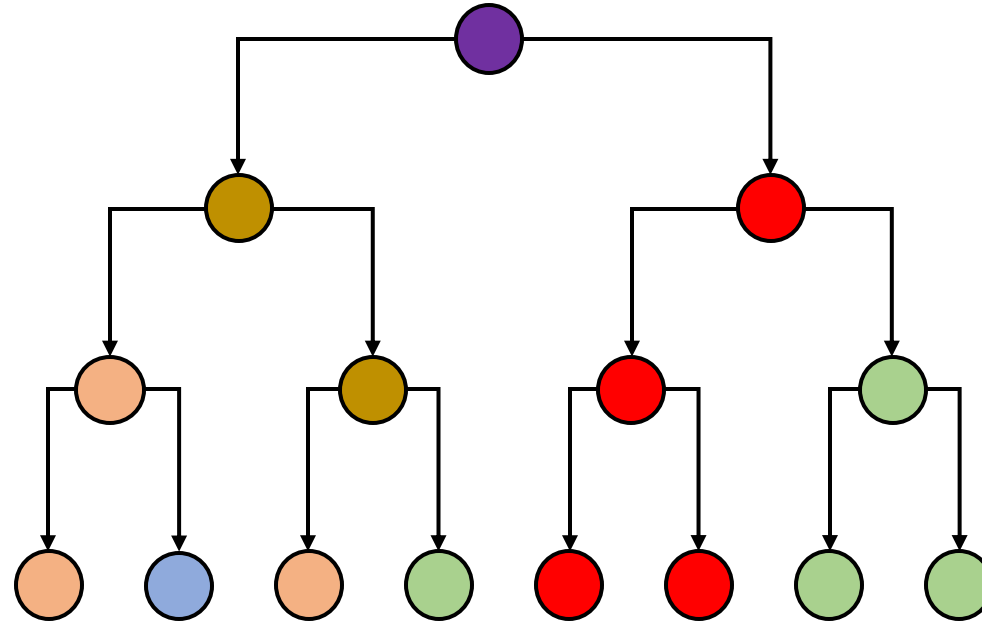
Multiple cell types  
**Cell differentiation**

# Organismal Development

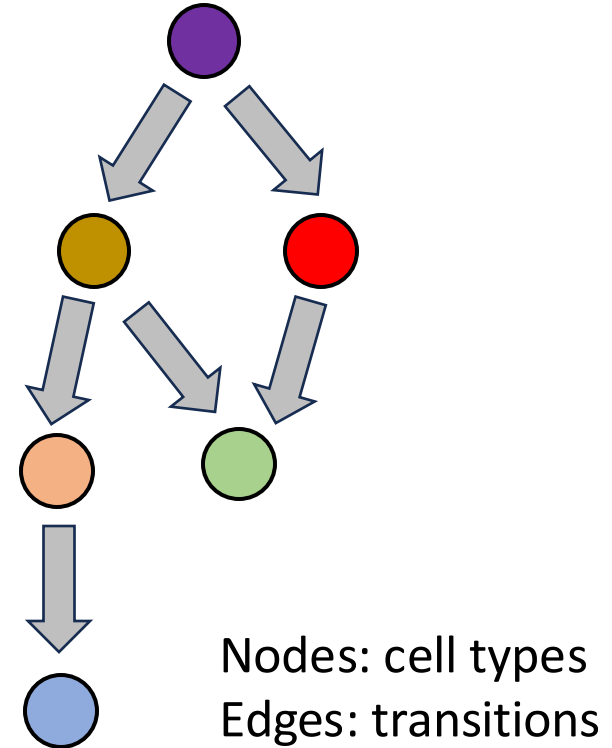
One cell  
One cell type



Multiple cells  
Multiple cell types  
**Embryogenesis**



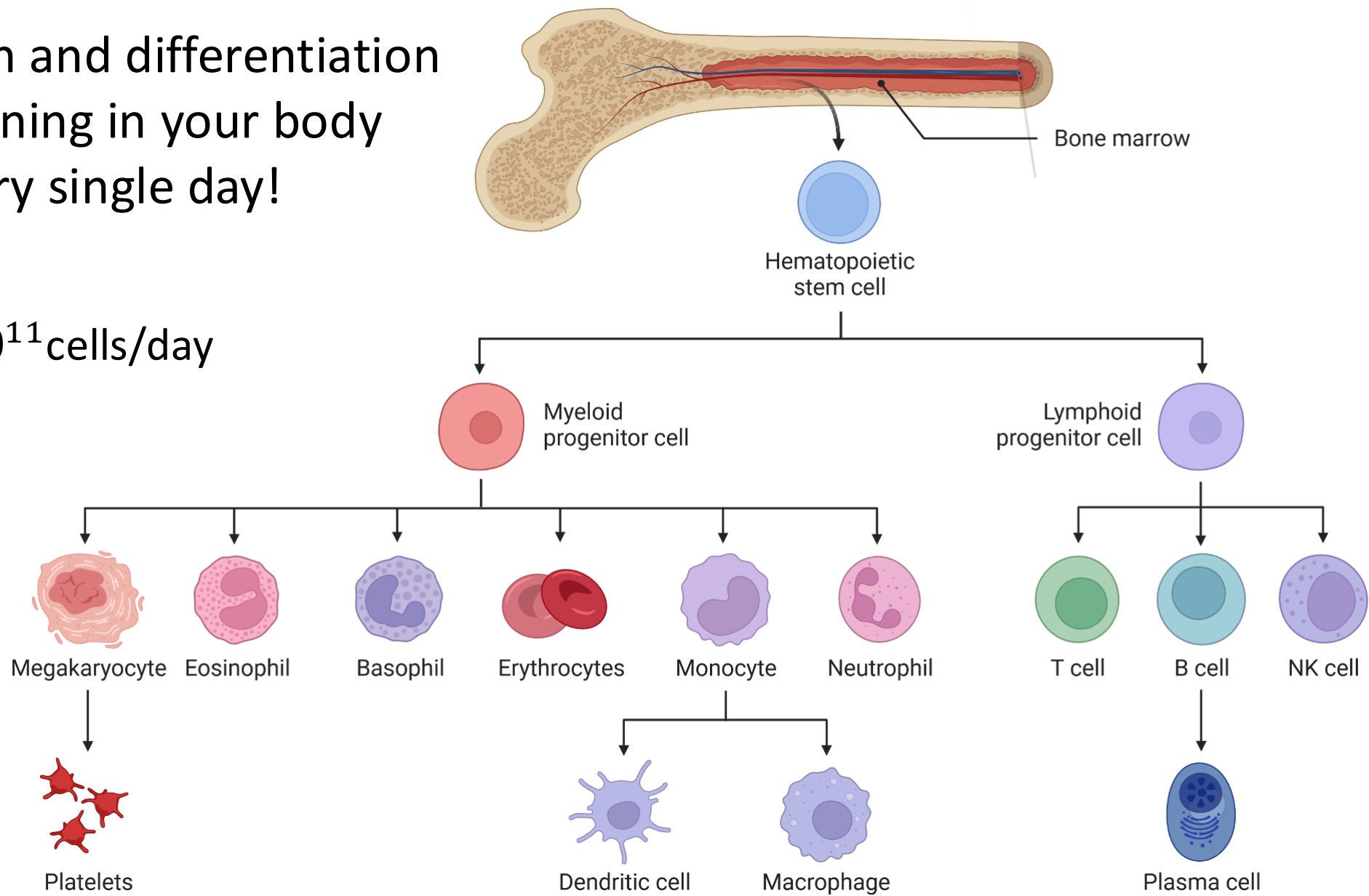
**Cell lineage tree  $T$**   
Rooted tree with leaves  
representing cells in the organism



**Cell differentiation map  $F$**   
Directed graph showing  
cell type transitions

Cell division and differentiation  
is happening in your body  
every single day!

$\sim 5 \times 10^{11}$  cells/day



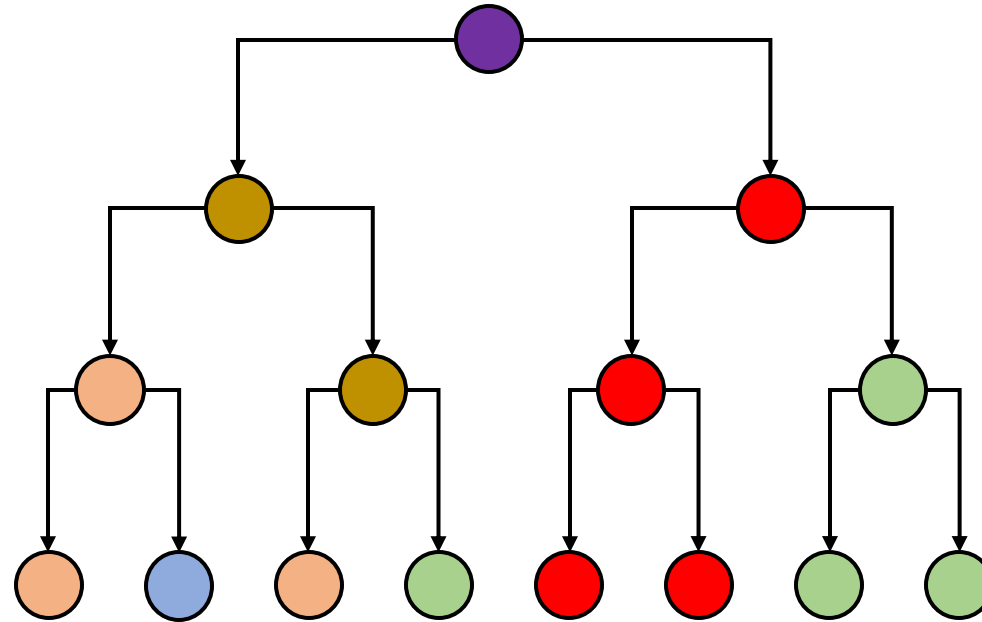
**Human hematopoiesis differentiation map**

# Organismal Development

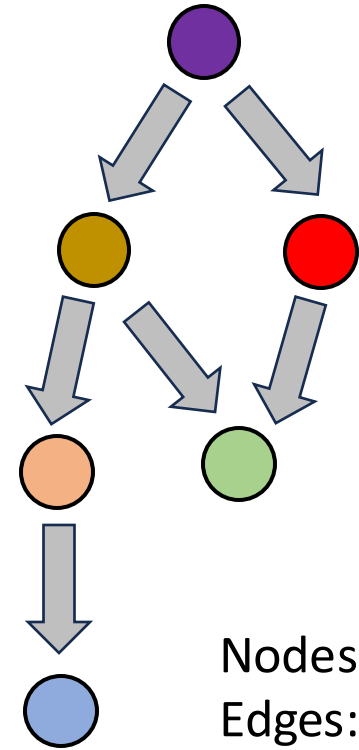
One cell  
One cell type



Multiple cells  
Multiple cell types  
**Embryogenesis**



Cell lineage tree  $T$



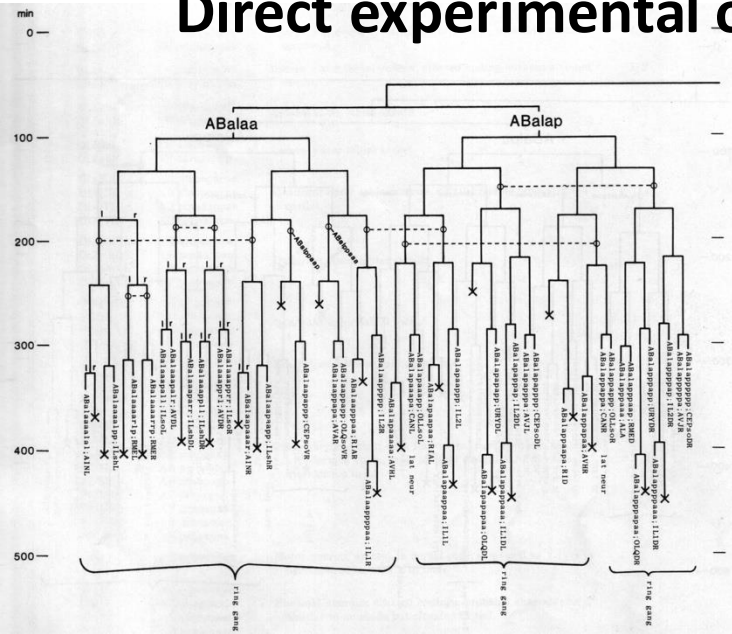
Nodes: cell types  
Edges: transitions

Cell differentiation map  $F$

## Central problem in developmental biology

What is the history of cell division and differentiation during development?

# Direct experimental observations

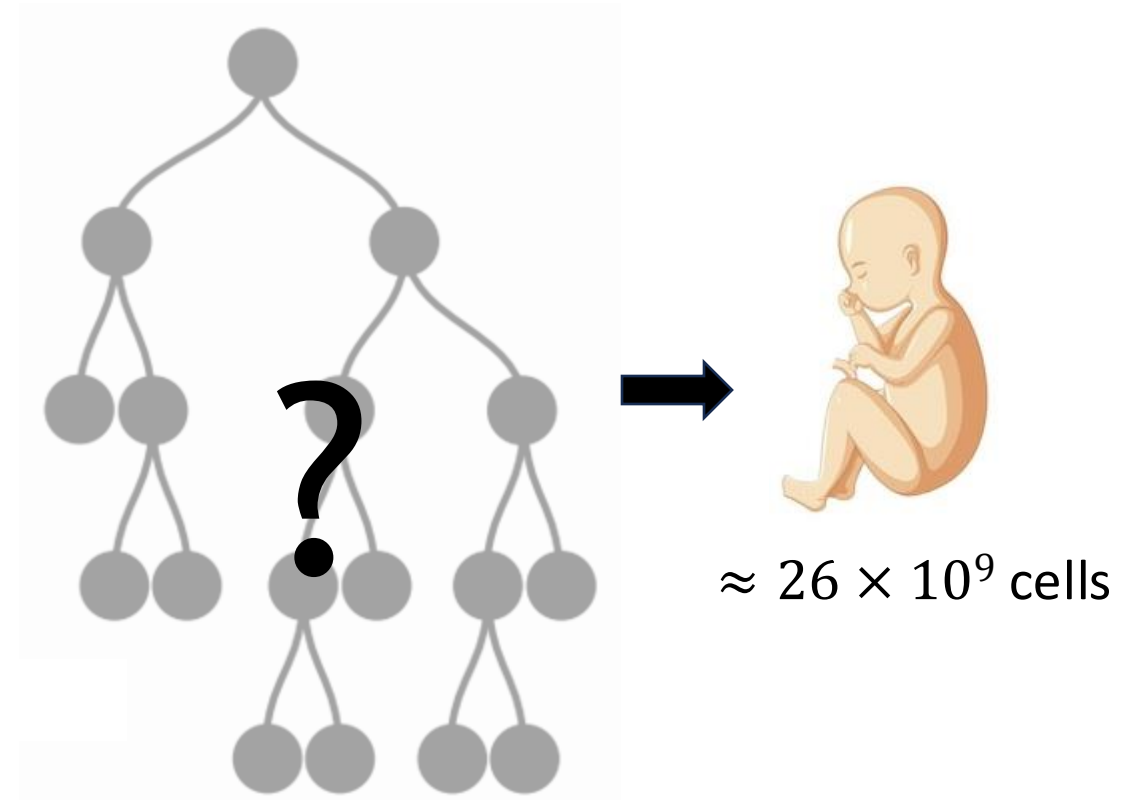


*Caenorhabditis  
elegans*



959 cells

Cell division history and differentiation  
of **every** cell has been mapped!



$\approx 26 \times 10^9$  cells

What is the history of cell division and  
differentiation during mammalian  
development?

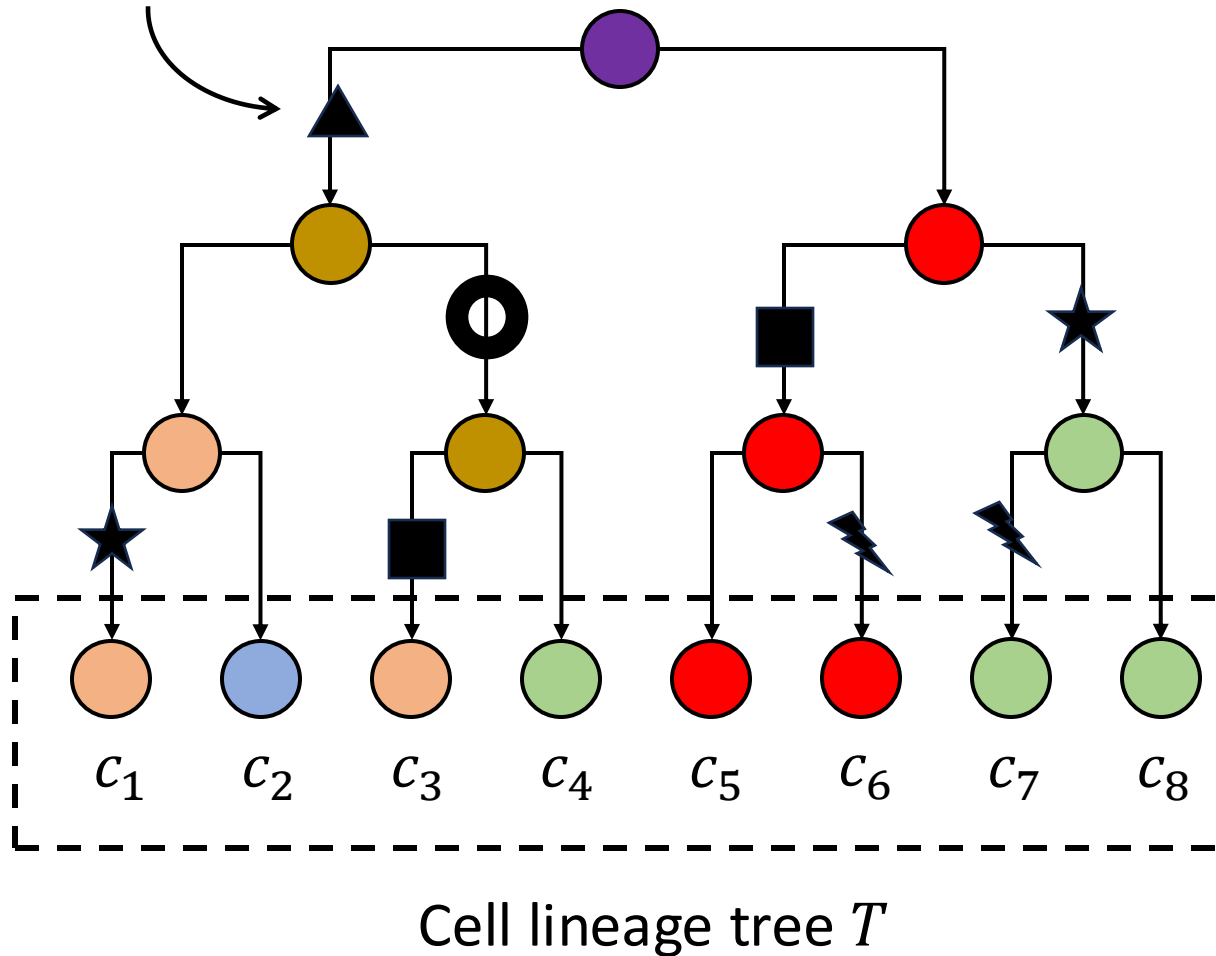


2002 Nobel Prize in  
Physiology or Medicine  
S. Brenner, H. Horvitz  
and J. Sulston

*“for their discoveries concerning genetic regulation  
of organ development and programmed cell death”*

# The Era of Lineage Tracing Technologies

**Artificial mutations** introduced using **genome editing** tools such as **CRISPR-Cas9**



Single-cell  
sequencing



Measurement of mutations and  
cell types of leaves of the tree

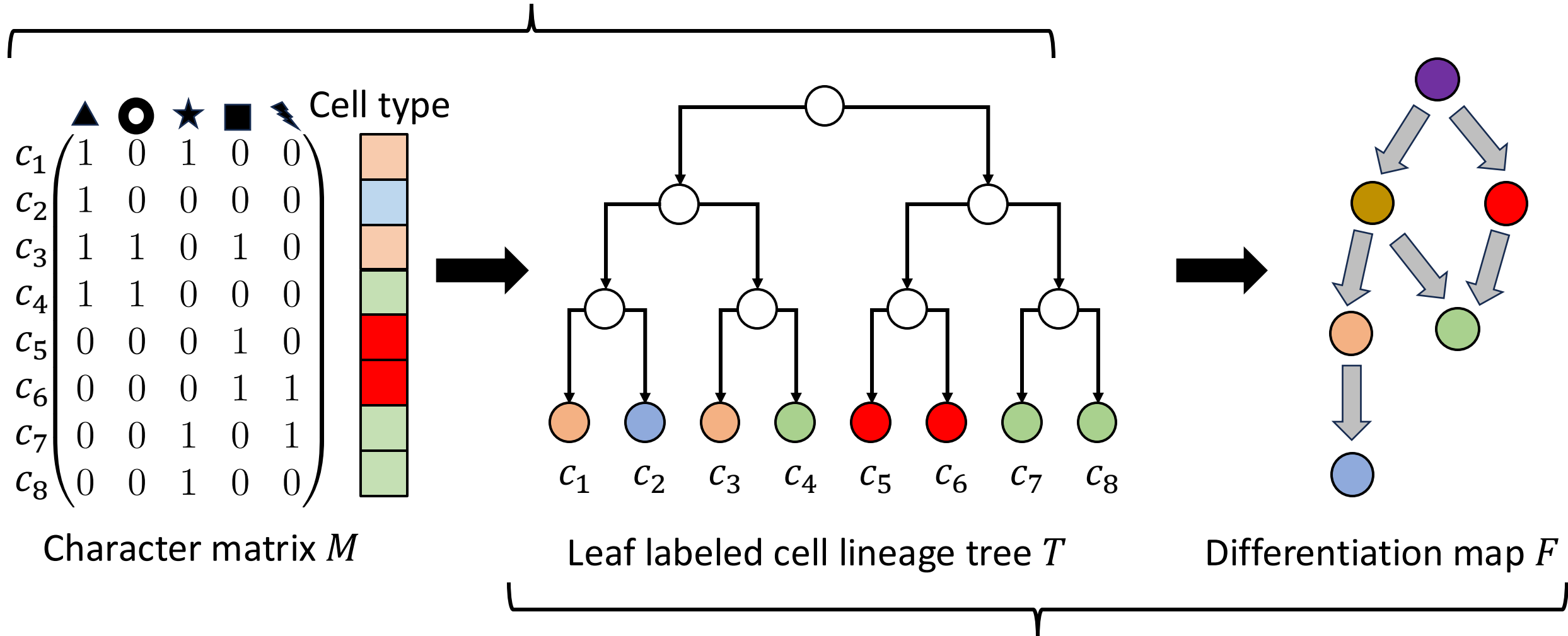
	▲	●	★	■	⚡	Cell type
$c_1$	1	0	1	0	0	Orange
$c_2$	1	0	0	0	0	Blue
$c_3$	1	1	0	1	0	Orange
$c_4$	1	1	0	0	0	Green
$c_5$	0	0	0	1	0	Red
$c_6$	0	0	0	1	1	Red
$c_7$	0	0	1	0	1	Green
$c_8$	0	0	1	0	0	Green

Character matrix  $M$

**Lineage tracing data**



## Problem 1: Cell lineage tracing

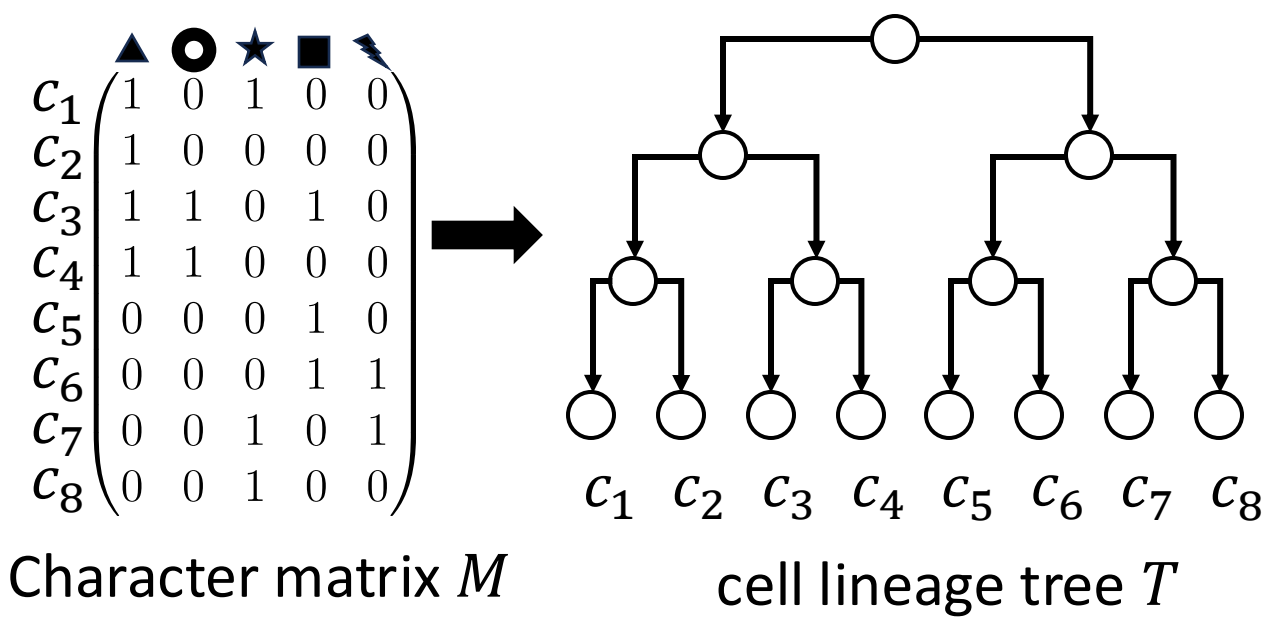


## Problem 2: Cell differentiation mapping

# (1) Cell lineage tracing

- Star homoplasmy model for CRISPR-Cas9 mutations
- **Startle** infers more accurate cell lineage trees than competing methods

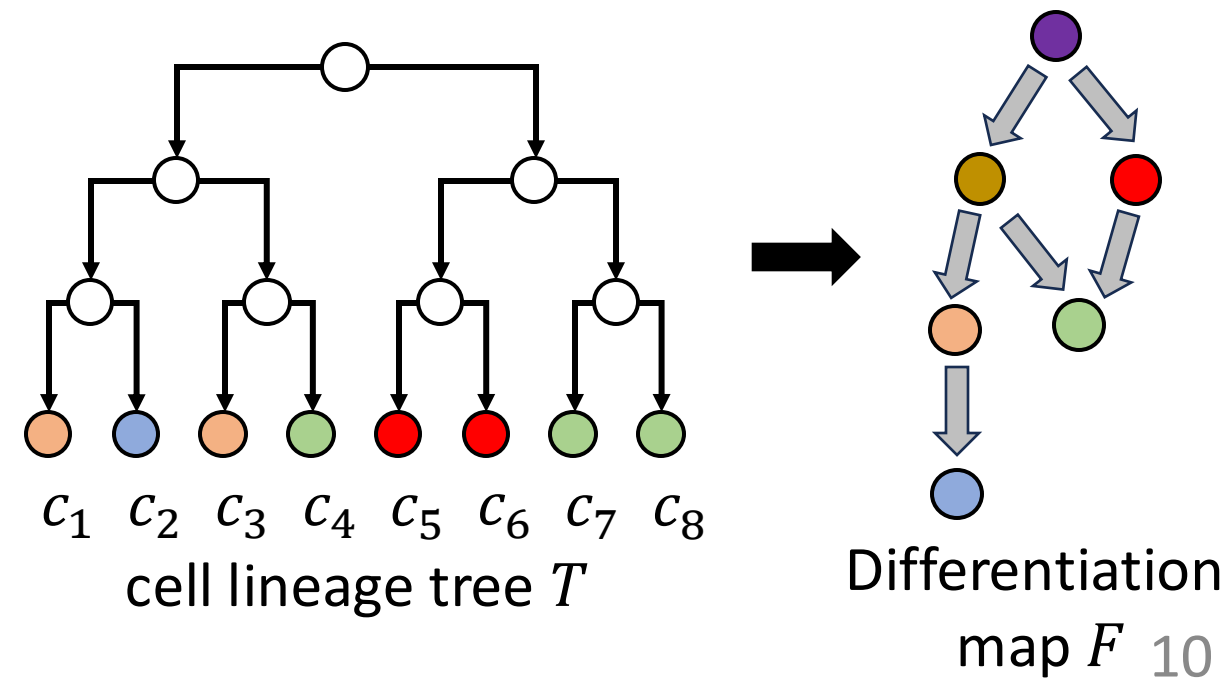
Sashittal\*, Schmidt\* et al., *Cell Systems*, 2023  
Also accepted at RECOMB 2023



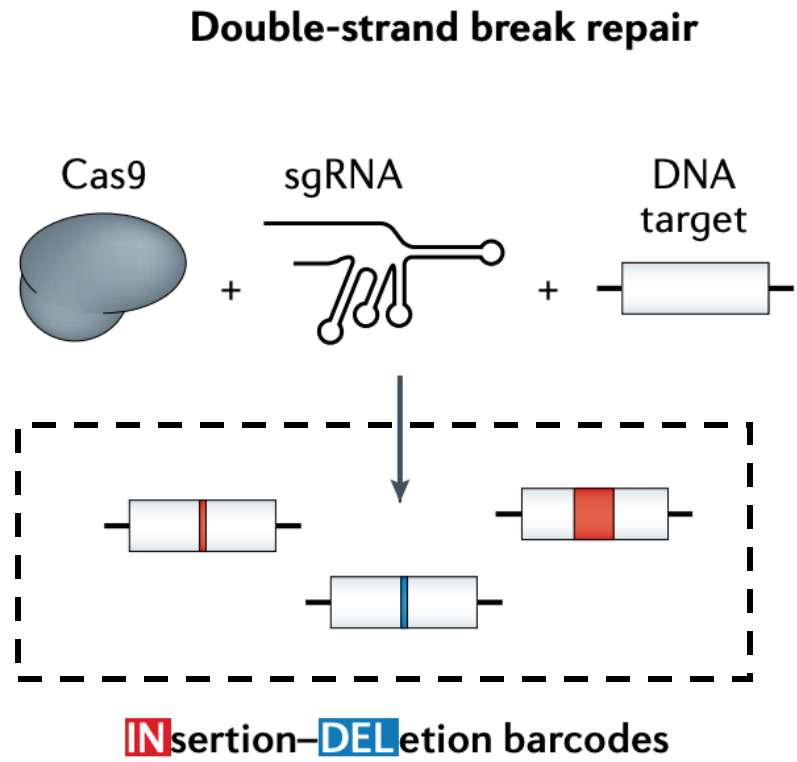
# (2) Cell differentiation mapping

- Formalized the problem of inferring cell differentiation maps from lineage tracing data
- **Carta** balances the trade-off between the complexity and fit of the differentiation map

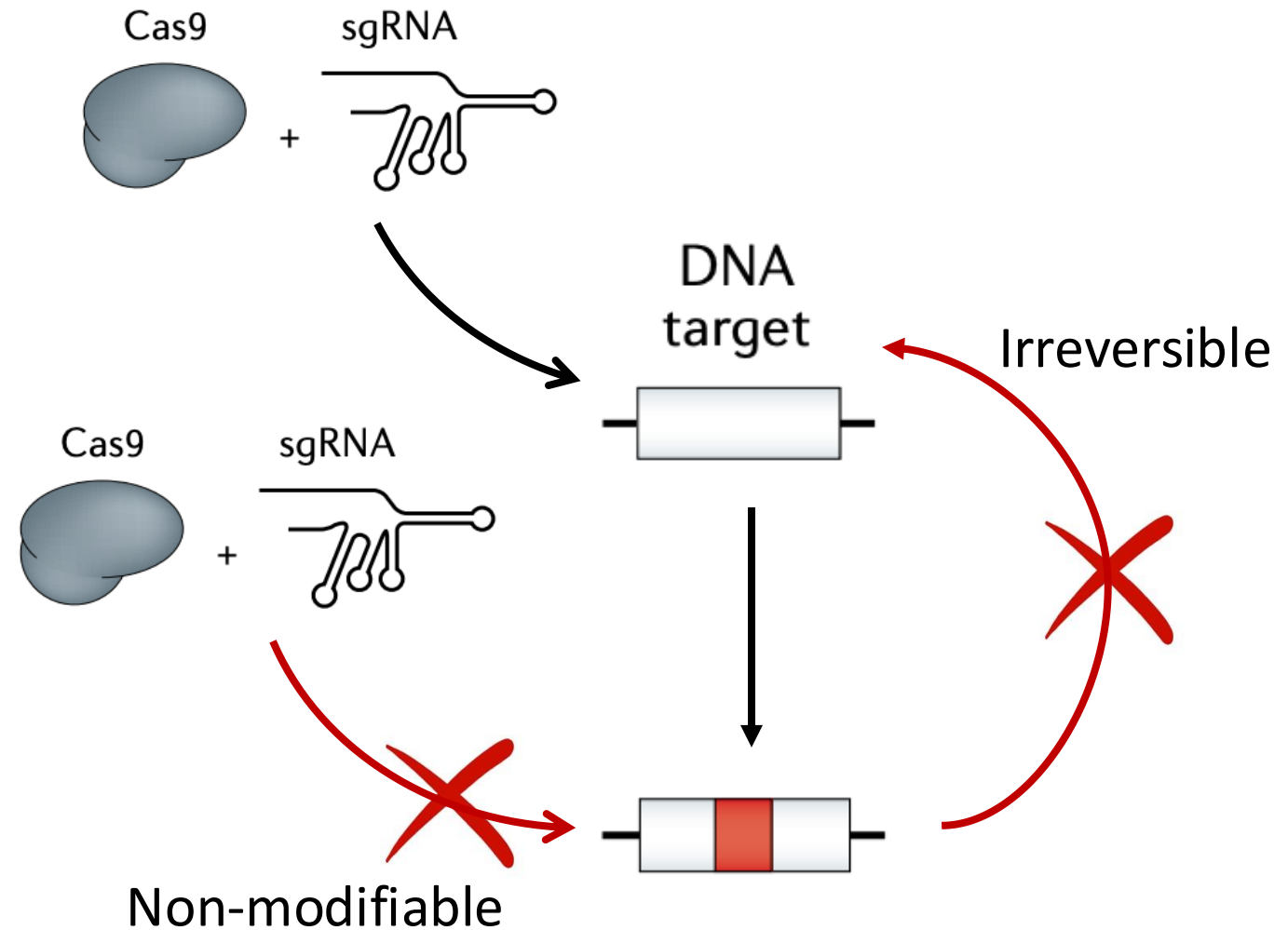
Sashittal\*, Zhang\* et al., *Nature Methods*, 2025  
Also accepted at RECOMB 2025



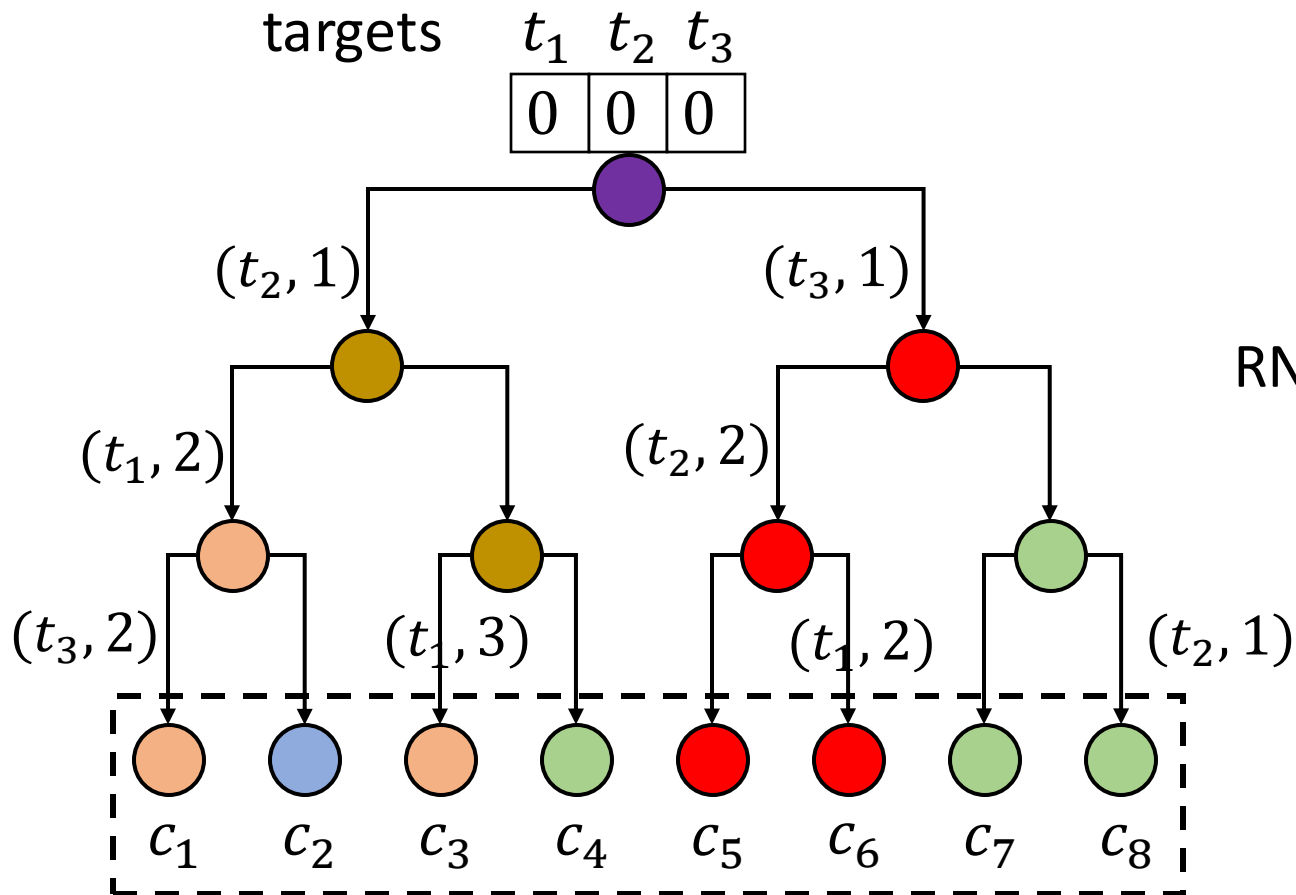
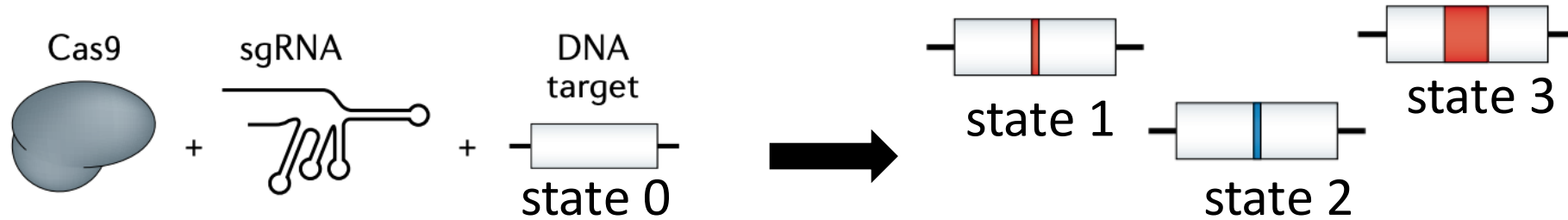
# CRISPR-Cas9-based lineage tracing



- ✓ Irreversible
- ✓ Non-modifiable
- ✓ Multi-state



# CRISPR-Cas9-based lineage tracing



Single-cell  
RNA sequencing



	$t_1$	$t_2$	$t_3$	Cell type
$c_1$	2	1	2	Orange
$c_2$	2	1	0	Blue
$c_3$	3	1	0	Orange
$c_4$	0	1	0	Green
$c_5$	0	2	1	Red
$c_6$	2	2	1	Red
$c_7$	0	0	1	Green
$c_8$	0	1	1	Green

# CRISPR-Cas9-based lineage tracing

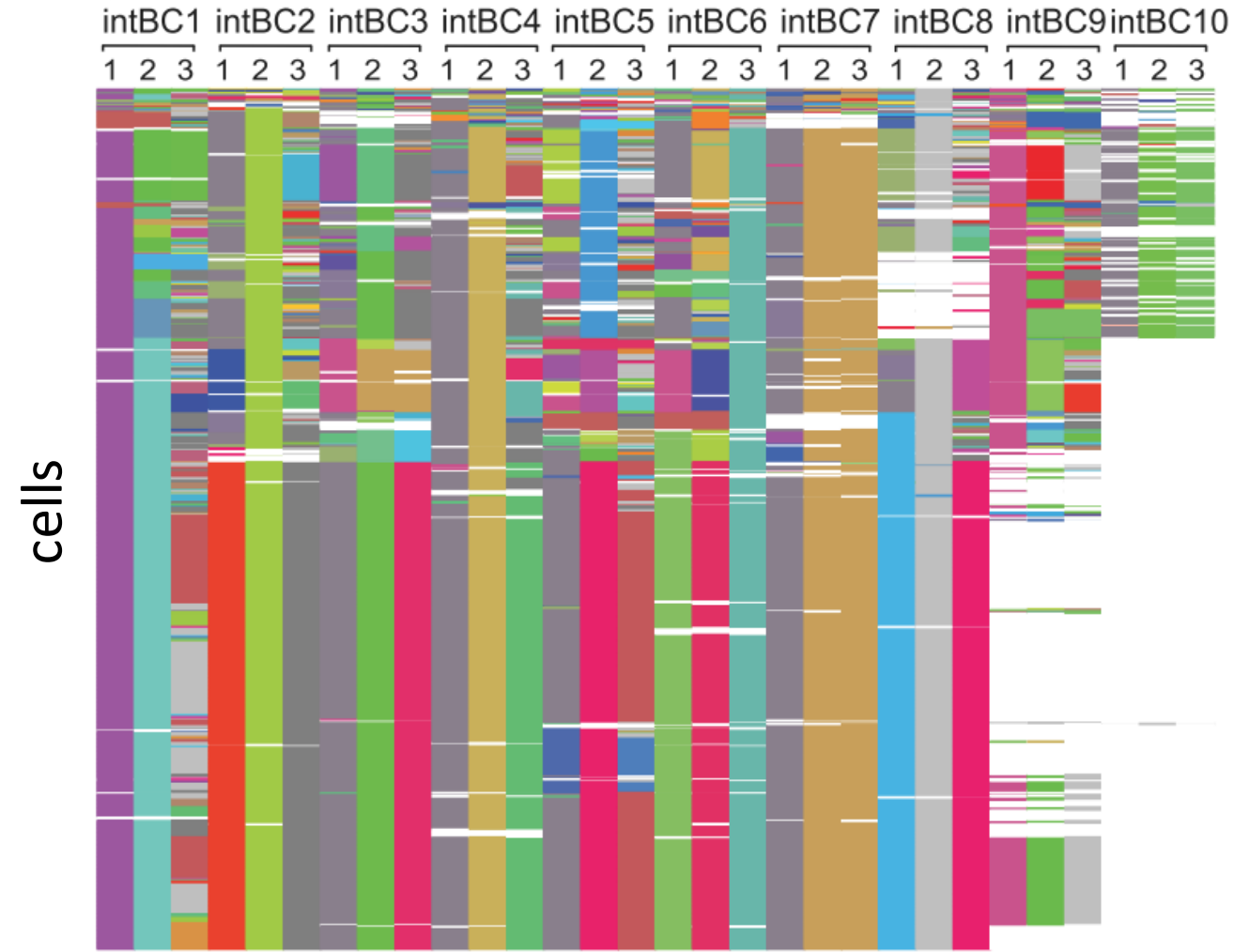
## Challenges in real data

- Large number (50 to 100) of states (indels) for each character (target site)

- Large number (100s to 1000s) of cells

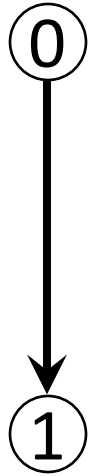
- What is the model for the evolution of CRISPR-Cas9 induced mutations?

Specialized methods have been introduced and benchmarked in a DREAM challenge (Gong et al., 2021, Cell Systems)



# Specialized models for CRISPR-Cas9-based lineage tracing

## Two-state Camin-Sokal model



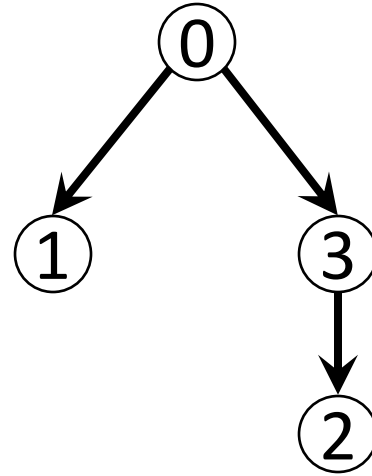
Camin et al., 1965

- ✗ Multi-state
- ✓ Irreversible
- \*✓ Non-modifiable

McKenna et al., *Science* (2016)

Raj et al., *Nature Biotechnology* (2018)

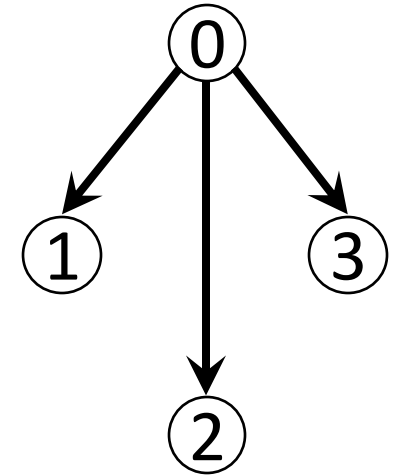
## Multi-state Camin-Sokal model



Felsenstein et al., 2004

- ✓ Multi-state
- ✓ Irreversible
- ✗ Non-modifiable

## Multi-state Star homoplasmy model



Sashittal et al., 2023

- ✓ Multi-state
- ✓ Irreversible
- ✓ Non-modifiable

# Star homoplasy tree inference problem statement

**Star Homoplasy Problem [Sashittal et al., 2023]**

Given character matrix  $M$  and mutation weights  $w$ , find star homoplasy phylogeny  $T$  for  $M$  that minimizes parsimony score  $W(T)$ .

**Theorem [Sashittal et al., 2023]**

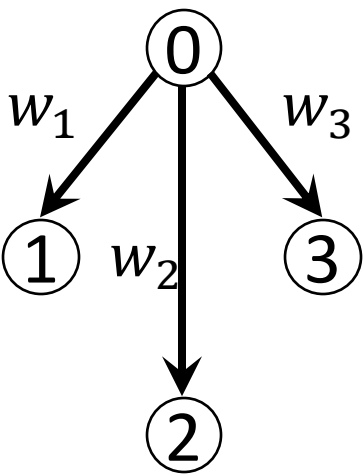
Star homoplasy problem is NP-hard, even when the number  $k$  of homoplasies is fixed and  $k \geq 4$ .

*\*Reduction from Cubic Vertex Cover Problem*

**Input**

	$t_1$	$t_2$	$t_3$
$c_1$	2	1	2
$c_2$	2	1	0
$c_3$	3	1	0
$c_4$	0	1	0
$c_5$	0	2	1
$c_6$	2	2	1
$c_7$	0	0	1
$c_8$	0	1	1

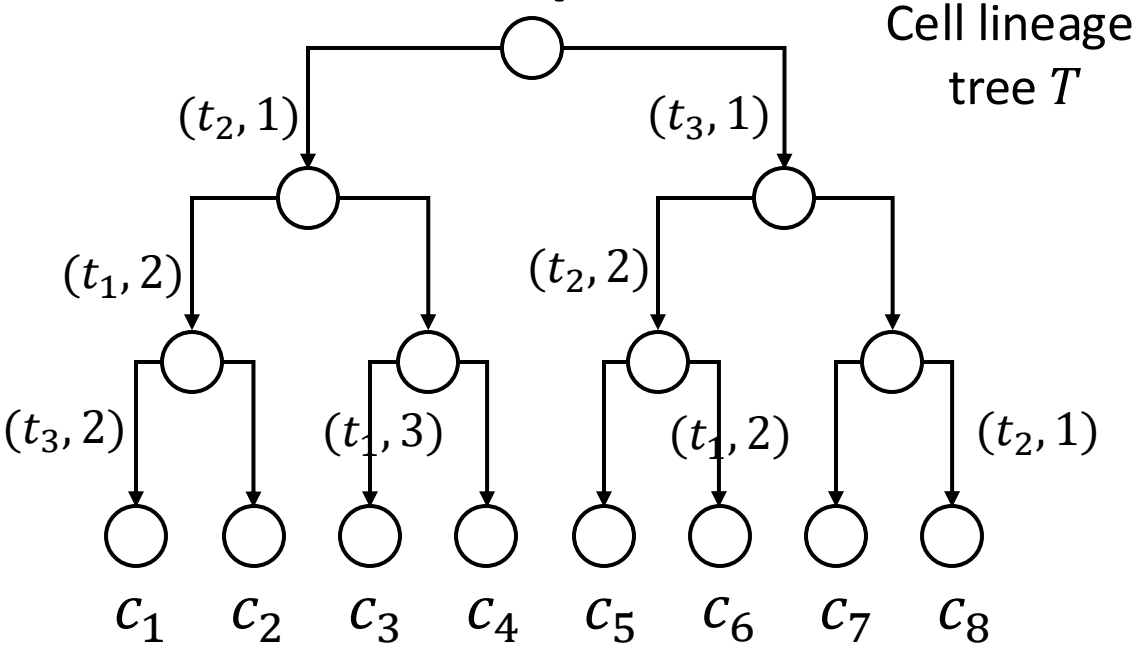
Character matrix  $M$



Weights indicating probability of mutation



**Output**



$$W(T) = 3w_1 + 4w_2 + w_3$$

# Startle performs hill climbing in the space of trees

Search through  
tree space using  
NNI moves

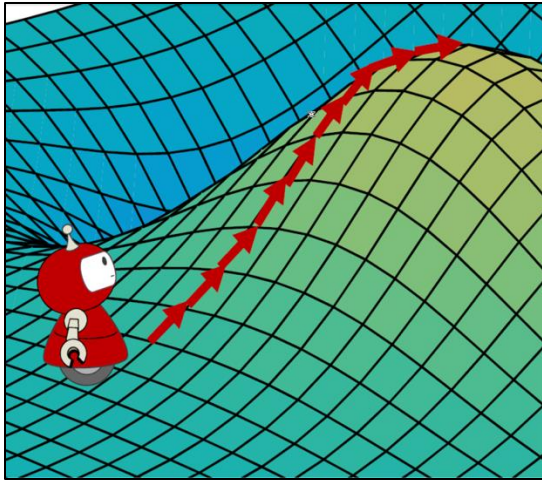
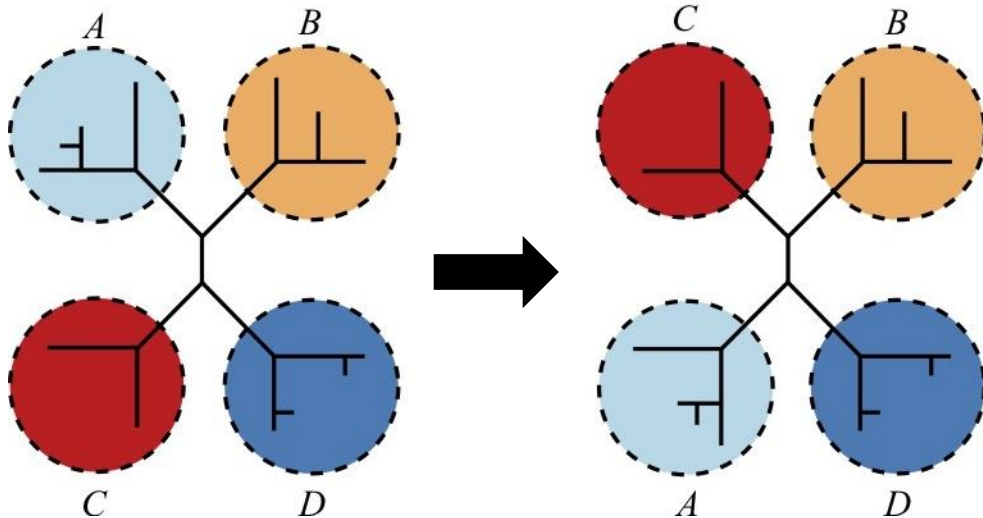


Figure from mathworks.com



Nearest neighbor interchange (NNI)

How do we evaluate a given tree  $T$ ?

**Small Star Homoplasy Problem [Sashittal et al., 2023]**

Given a tree  $T$  for character matrix  $M$  and mutation weights  $w$ , find the minimum parsimony score  $W(T)$ .

**Theorem [Sashittal et al., 2023]:**

Small Star Homoplasy problem can be solved using dynamic programming in  $O(nm)$  time.

**Theorem [Sashittal et al., 2023]:**

We can compute parsimony scores  $W(T')$  for all  $O(n)$  trees  $T'$  in the NNI neighborhood of a tree  $T$  in  $O(nmd)$  time, where  $d$  is the average depth of  $T$ .

\*Naïve implementation will take  $O(n^2m)$  time



# Star tree lineage estimator (Startle)



Henri Schmidt



Benjamin Raphael

## Character matrix

		characters	
		$c_1$	$c_2$
cells	$s_1$	1	0
	$s_2$	1	2
	$s_3$	1	1
	$s_4$	2	1

Startle

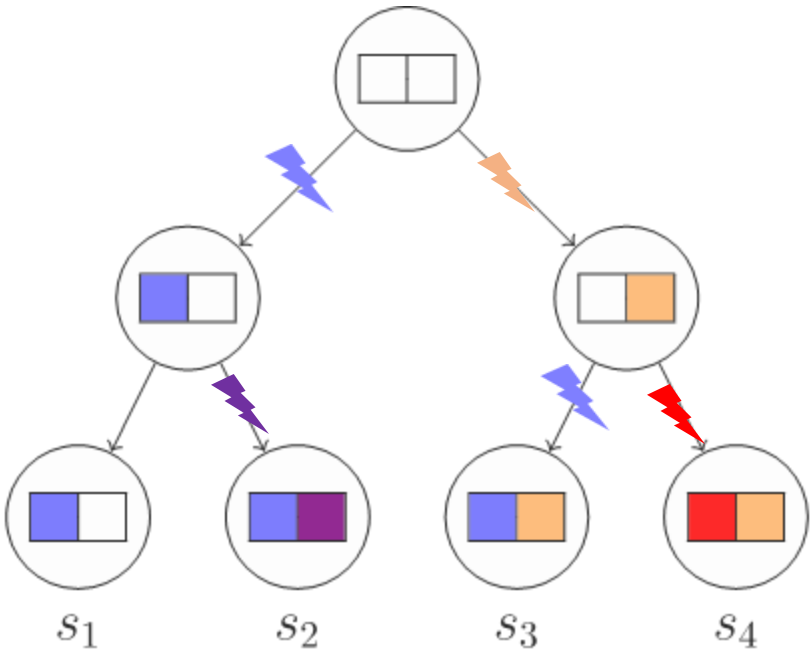


Tree search using nearest  
neighbor interchange  
(NNI) moves

+

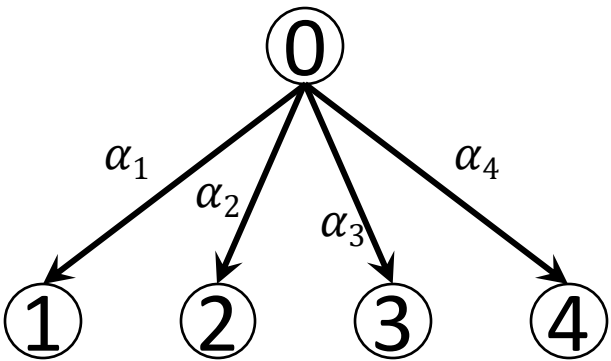
ILP for bounded-  
homoplasy version

## Maximum parsimony star homoplasy phylogeny



## Star homoplasy model

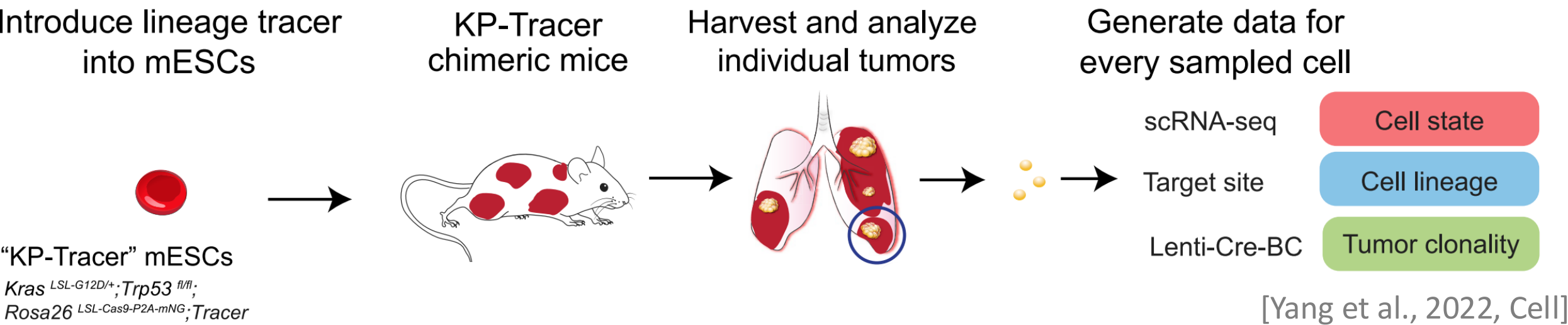
Unmutated state



Mutated states

Sashittal\*, Schmidt\*, et al.  
RECOMB 2023; Cell Systems 2023

# Mouse metastatic lung adenocarcinoma data



**Largest dataset in the study**  
(3724\_NT\_T1\_All):

*n* = 21108 cells across 5 tumors

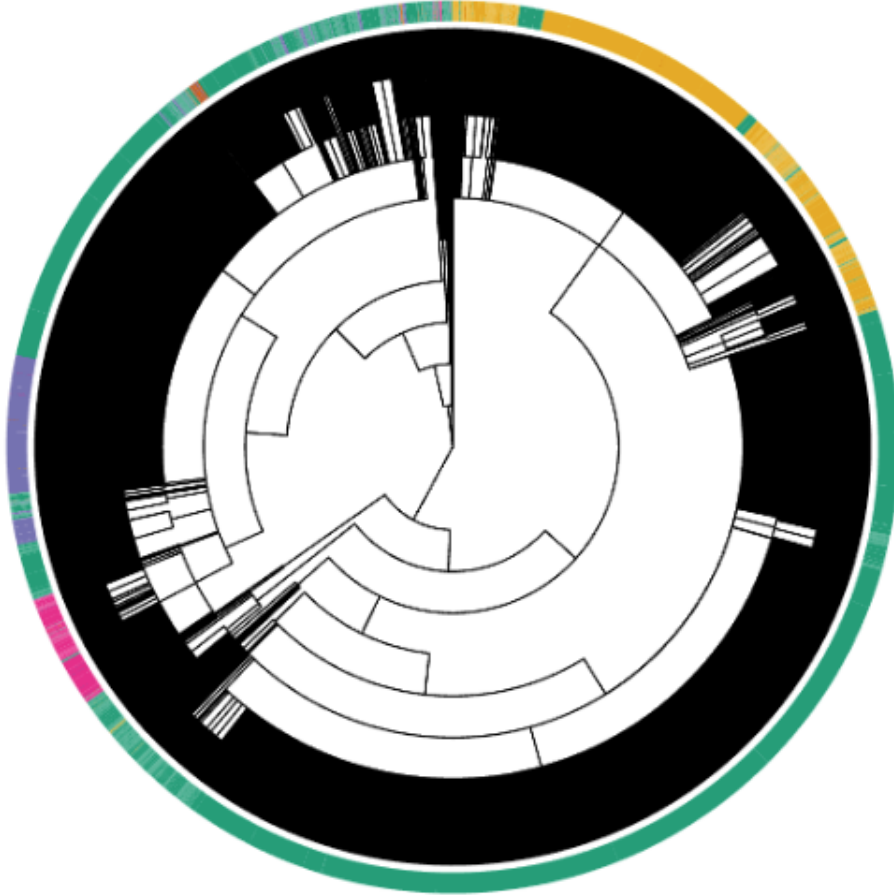
What is the cell lineage tree for these cancer cells?

Tumor	# of cells
Lung	14852
Soft tissue	3891
Liver met 1	90
Liver met 2	1512
Liver met 3	863

# Startle produces more parsimonious trees

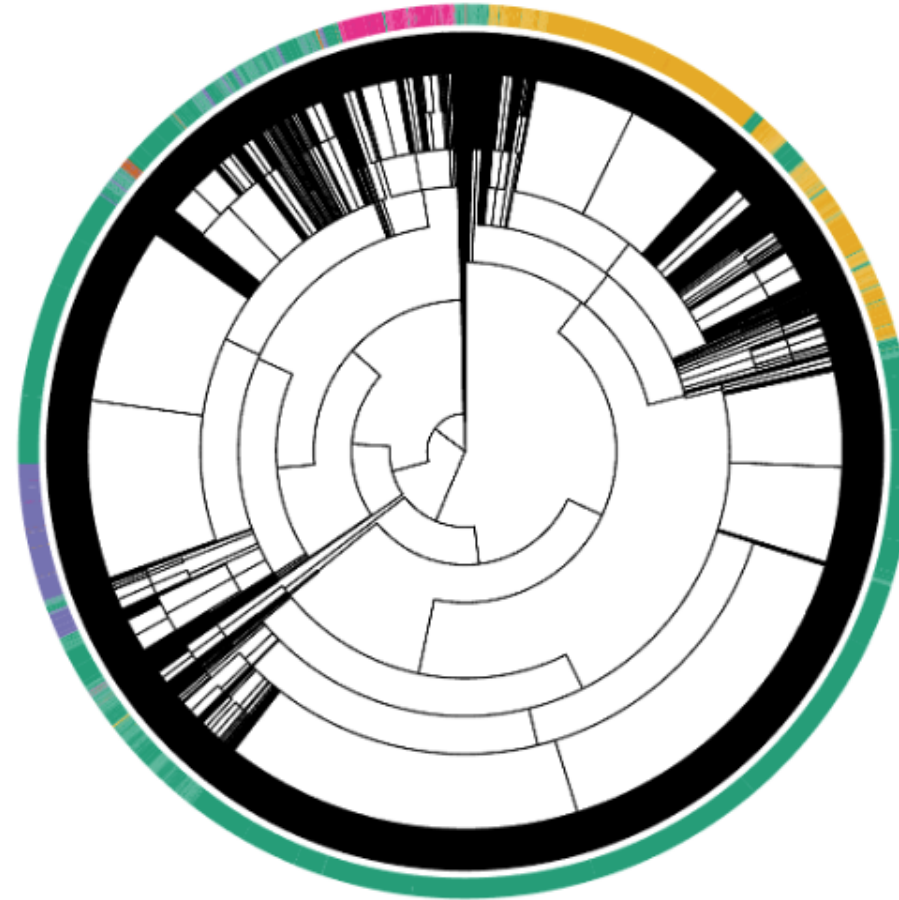
## Published phylogeny

Cassiopeia [Jones et al., 2021, *Genome Biology*]



Parsimony Score = 4827.43

## Startle phylogeny



Parsimony Score = **4715.5**

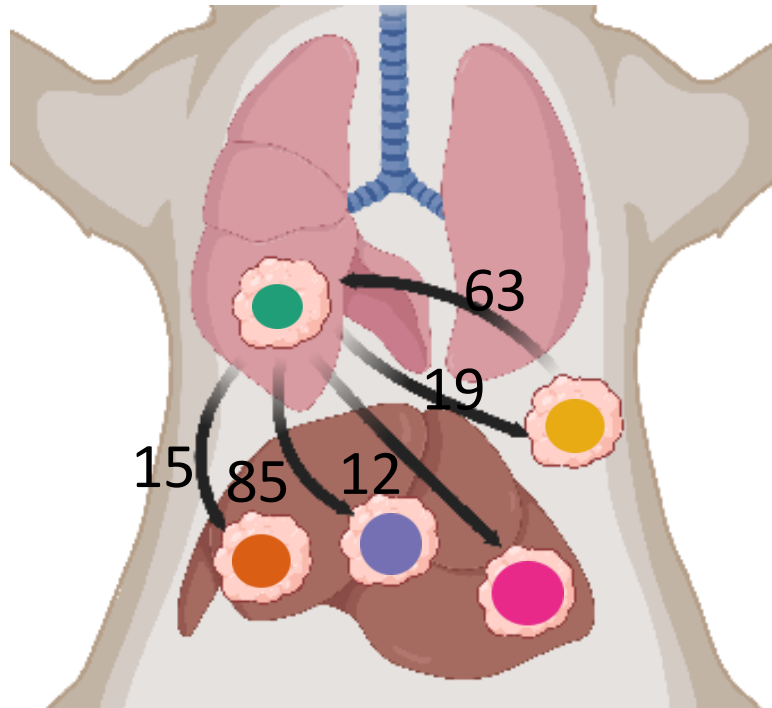
### Anatomical sites (cells)

- Primary tumor (14852)
- Liver met. 1 (90)
- Liver met. 2 (1512)
- Liver met. 3 (863)
- Soft tissue met. (3891)

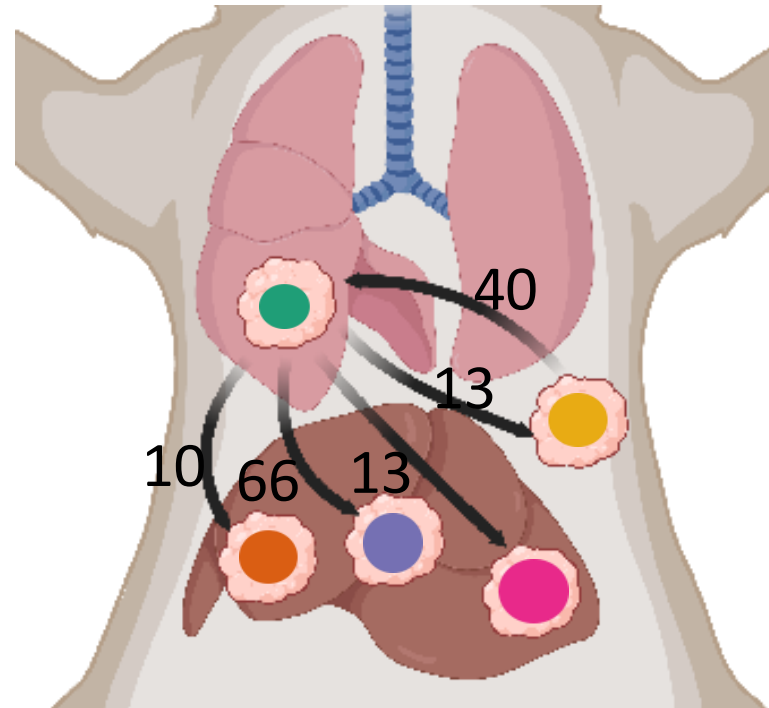
Total cells: 21108






# Startle trees have fewer migrations between anatomical sites

Inferred\* migrations  
from published tree



Inferred\* migrations  
from Startle tree

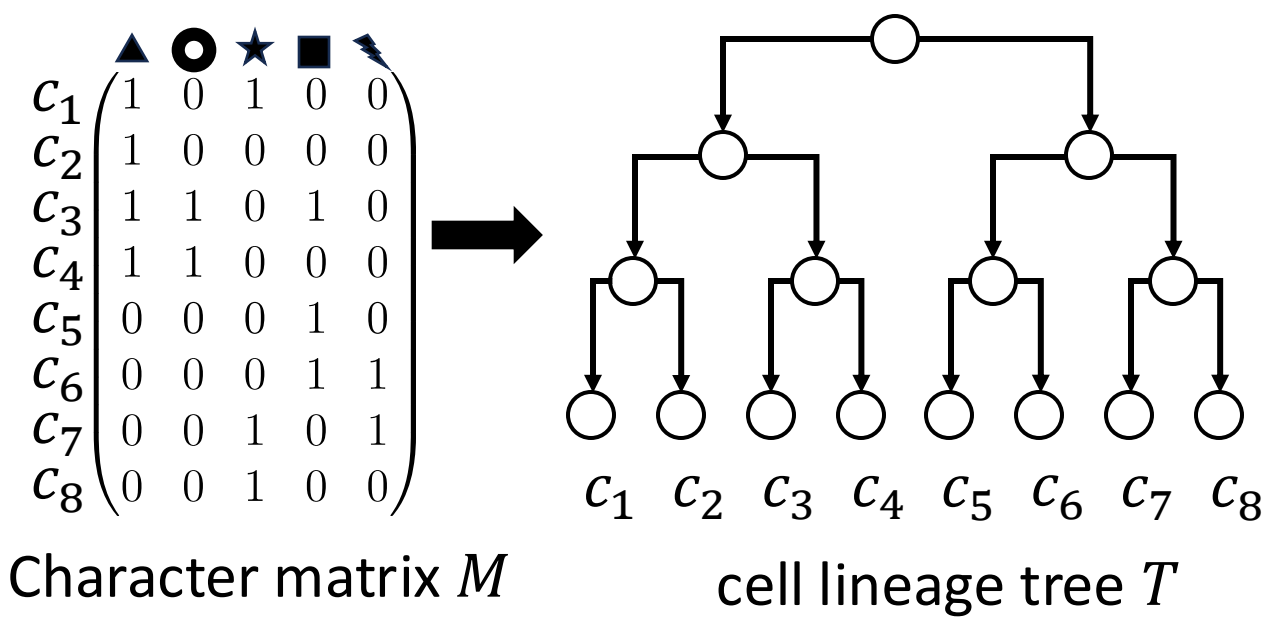


Anatomical sites (cells)	
	Primary tumor (14852)
	Liver met. 1 (90)
	Liver met. 2 (1512)
	Liver met. 3 (863)
	Soft tissue met. (3891)
Total cells: 21108	

# (1) Cell lineage tracing

- Star homoplasmy model for CRISPR-Cas9 mutations
- **Startle** infers more accurate cell lineage trees than competing methods

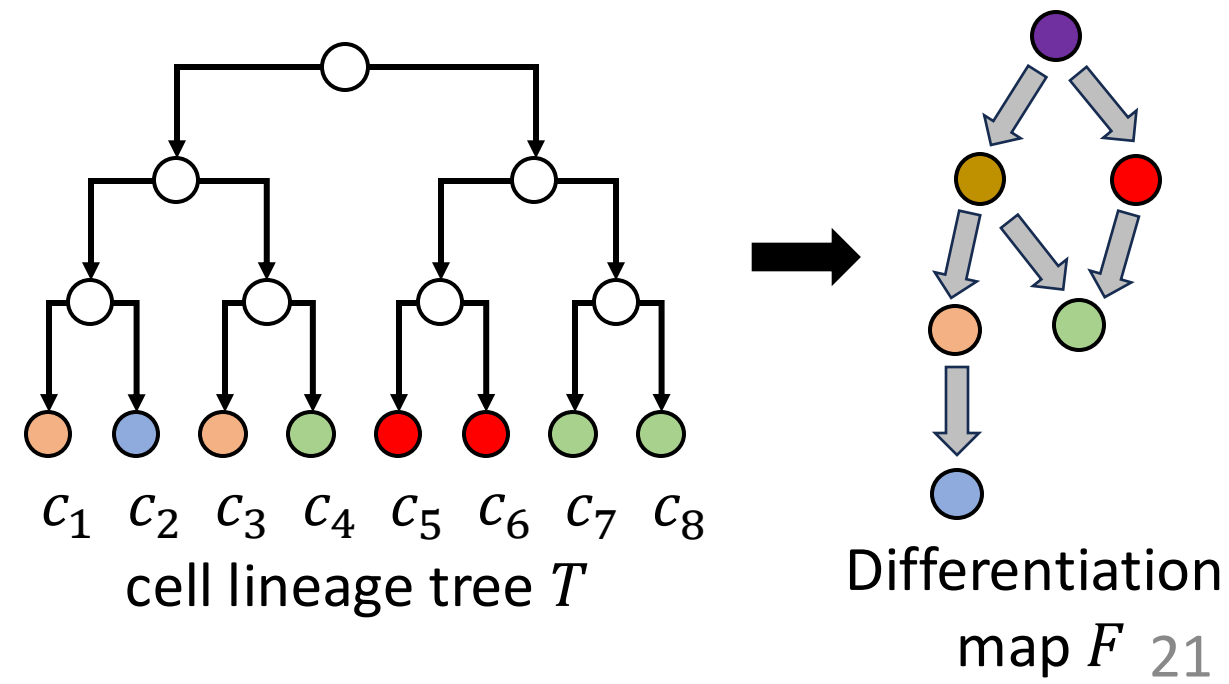
Sashittal\*, Schmidt\* et al., *Cell Systems*, 2023  
Also accepted at RECOMB 2023



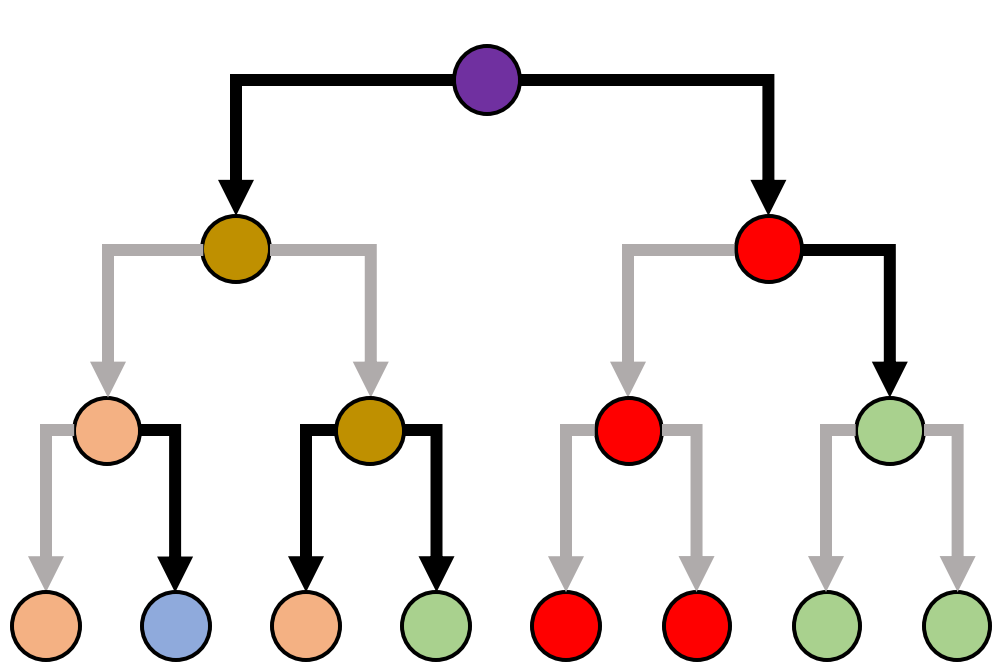
# (2) Cell differentiation mapping

- Formalized the problem of inferring cell differentiation maps from lineage tracing data
- **Carta** balances the trade-off between the complexity and fit of the differentiation map

Sashittal\*, Zhang\* et al., *Nature Methods*, 2025  
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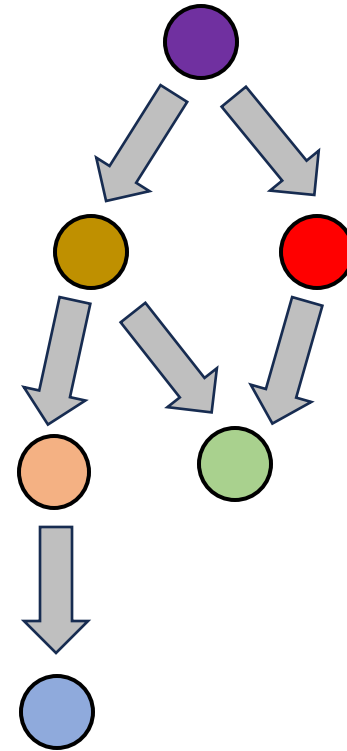


# Ancestral cell types reveal the differentiation map



**Cell lineage tree  $T$**   
with all cell types known

easy!  
➔



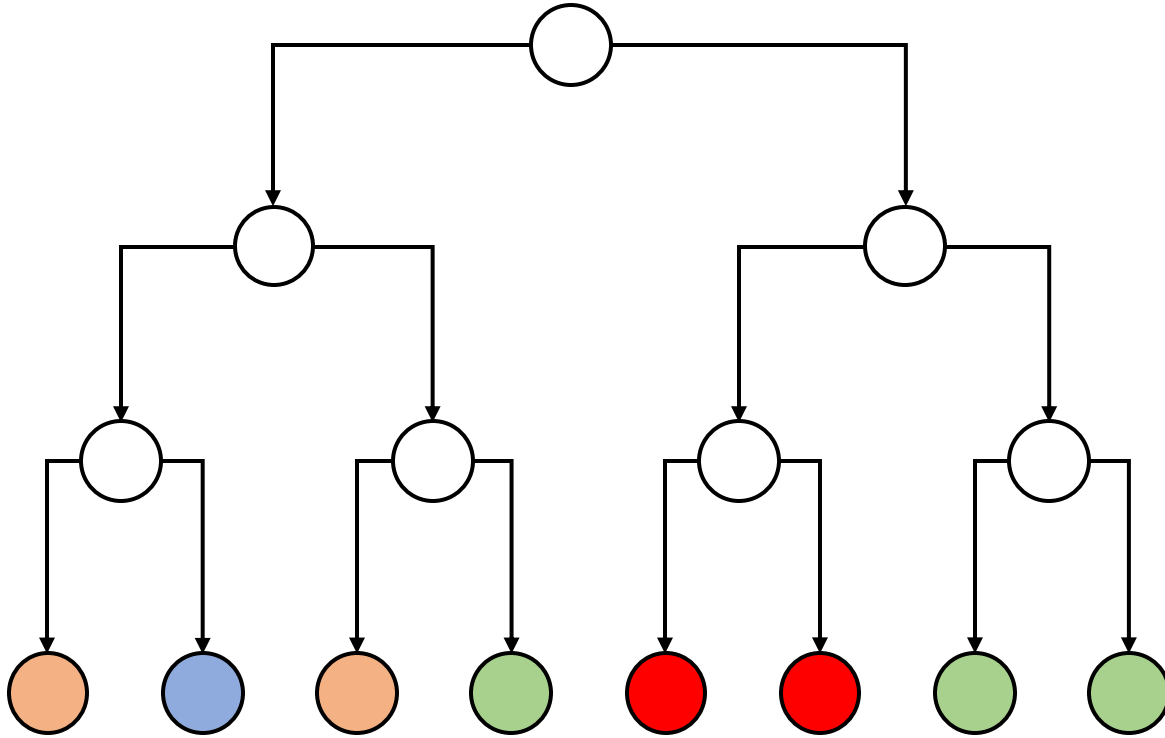
**Cell differentiation map  $F$**

Given ancestral cell types, we can trivially get:

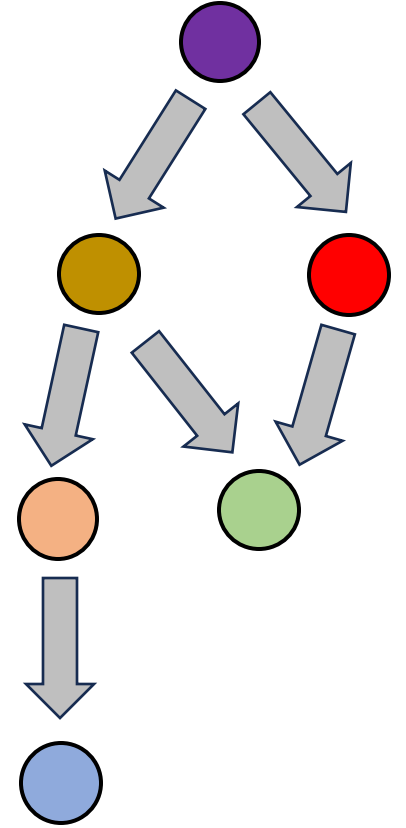
1. Cell types in the differentiation process
2. Transitions between cell types

# Key challenges in cell differentiation mapping

# Cell lineage tree



??



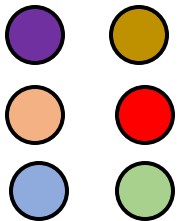
## Key challenges in inferring the type of ancestral cells

1. Which progenitors are not observed at present time?
2. Which of the observed cell types are progenitors?

## Unobserved progenitors

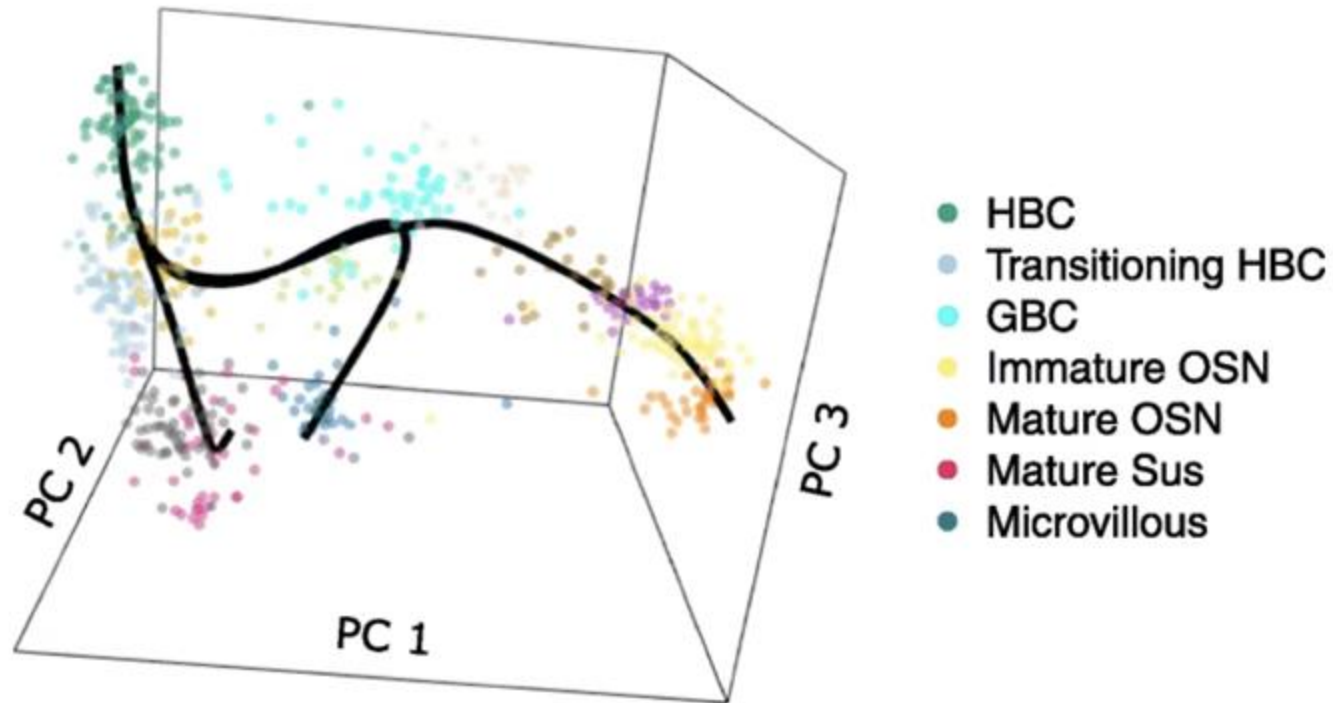
## Observed progenitors

## Terminal cell types



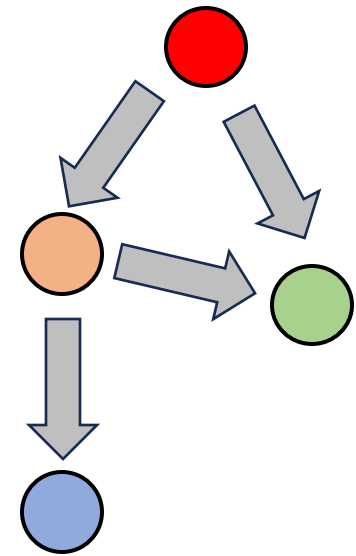
# Cell differentiation mapping

scRNA-seq data from one or more timepoints  
(with or without lineage information)



Principal curves or ridge estimation

All progenitors  
are observed



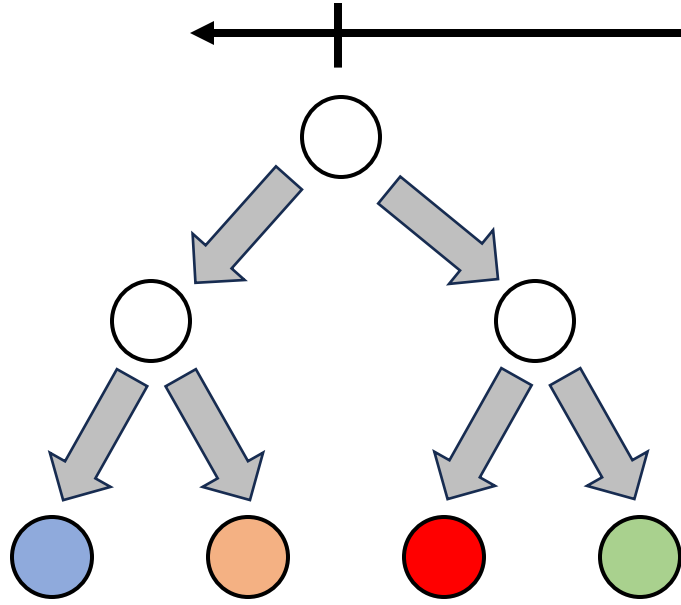
Trajectory inference methods

Trapnell et al., 2014, Nat. Biotech.;  
Haghverdi et al., 2016, Nat. Methods;  
Manno et al., 2018, Nature; Qiu et al.,  
2017a, Nat. Methods; Setty et al., 2016,  
Nat. Biotech and many more ....



# Cell differentiation mapping

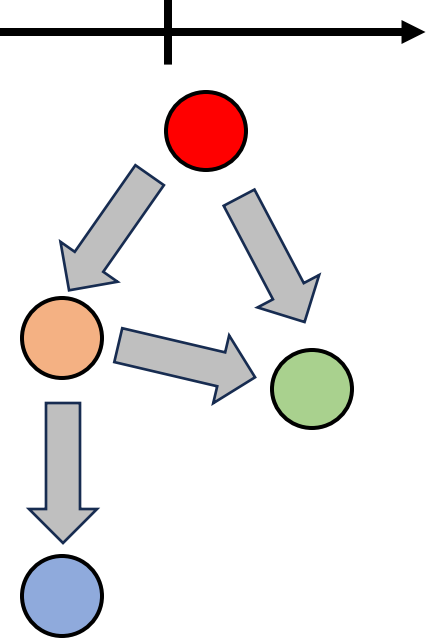
None of the progenitors  
are observed



Distance-based heuristics  
to infer tree-structured  
differentiation maps

Chan et al., 2019, Nature.; Yang et al.,  
2022, Cell; Kahlor et al., 2022, Cell

All progenitors  
are observed



Trajectory inference methods

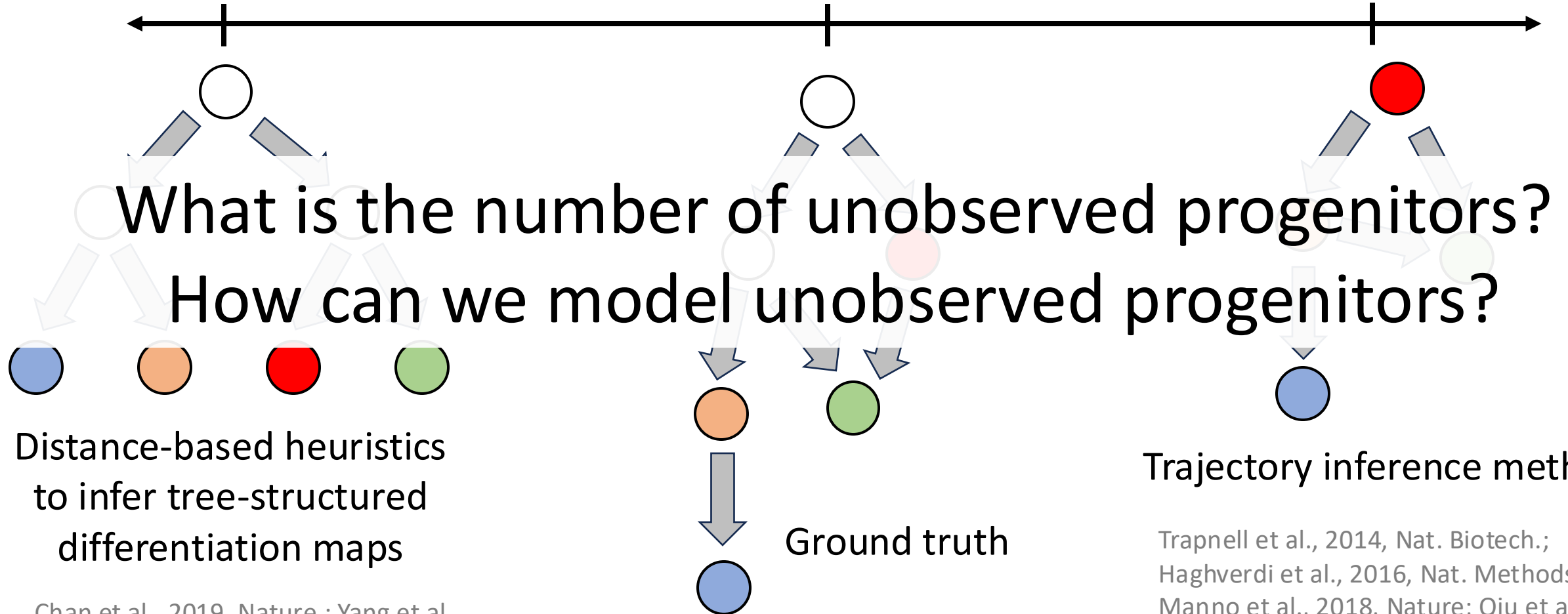
Trapnell et al., 2014, Nat. Biotech.;  
Haghverdi et al., 2016, Nat. Methods;  
Manno et al., 2018, Nature; Qiu et al.,  
2017a, Nat. Methods; Setty et al., 2016,  
Nat. Biotech and many more ....

# Cell differentiation mapping

None of the progenitors  
are observed

Early progenitors are not observed  
Late progenitors are observed

All progenitors  
are observed



Distance-based heuristics  
to infer tree-structured  
differentiation maps

Chan et al., 2019, Nature.; Yang et al.,  
2022, Cell; Kahlor et al., 2022, Cell

Ground truth

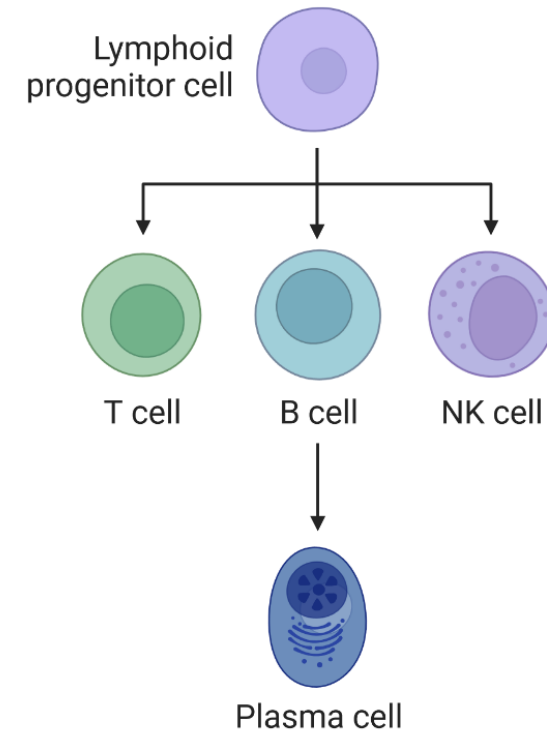
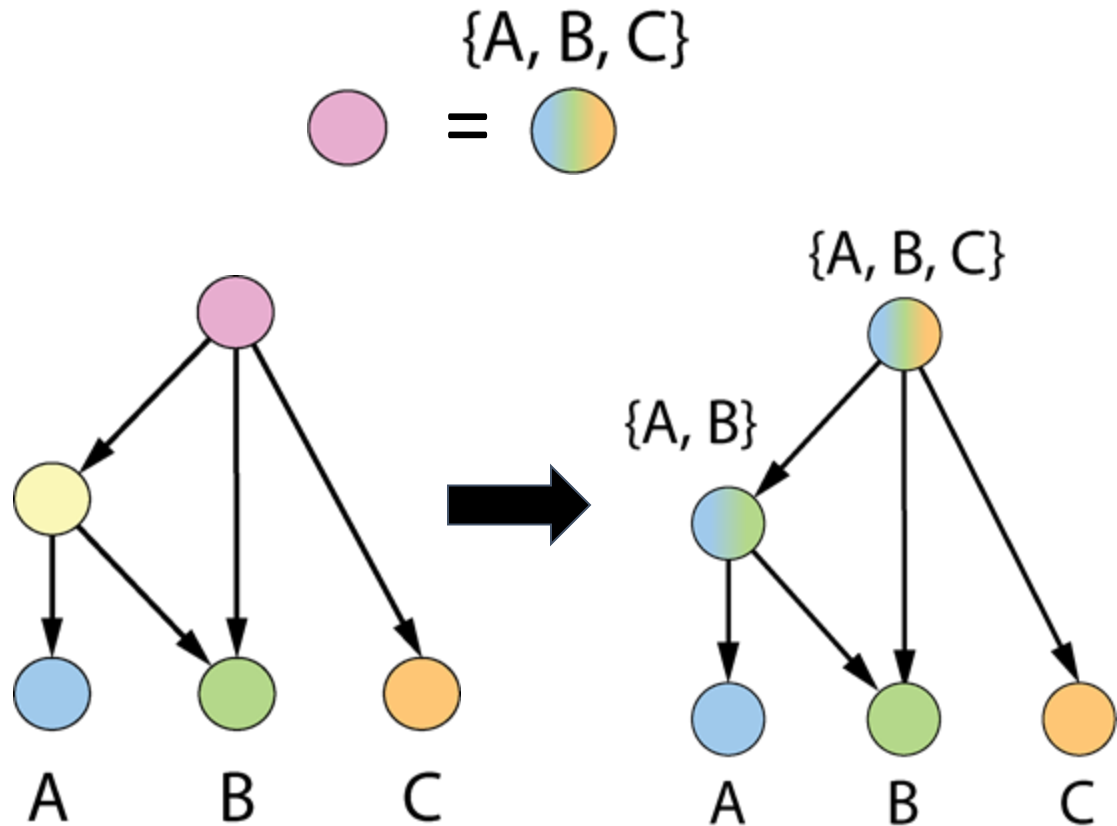
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Haghverdi et al., 2016, Nat. Methods;  
Manno et al., 2018, Nature; Qiu et al.,  
2017a, Nat. Methods; Setty et al., 2016,  
Nat. Biotech and many more ....

# Modeling unobserved progenitors: Potency Set

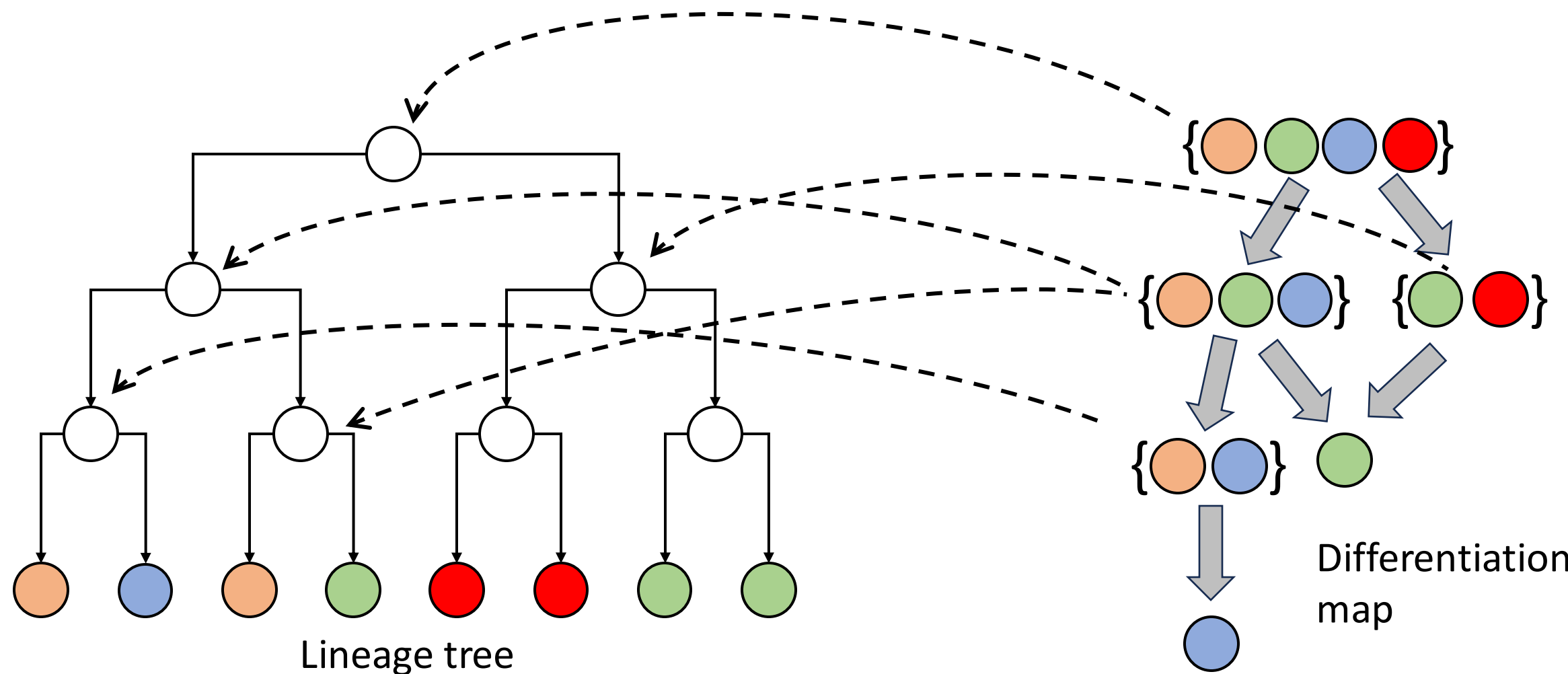
**Definition:** *potency* set  $S = \{\text{cell types that their descendants can differentiate into}\}$

**Formalizes how developmental biologists describe progenitors**



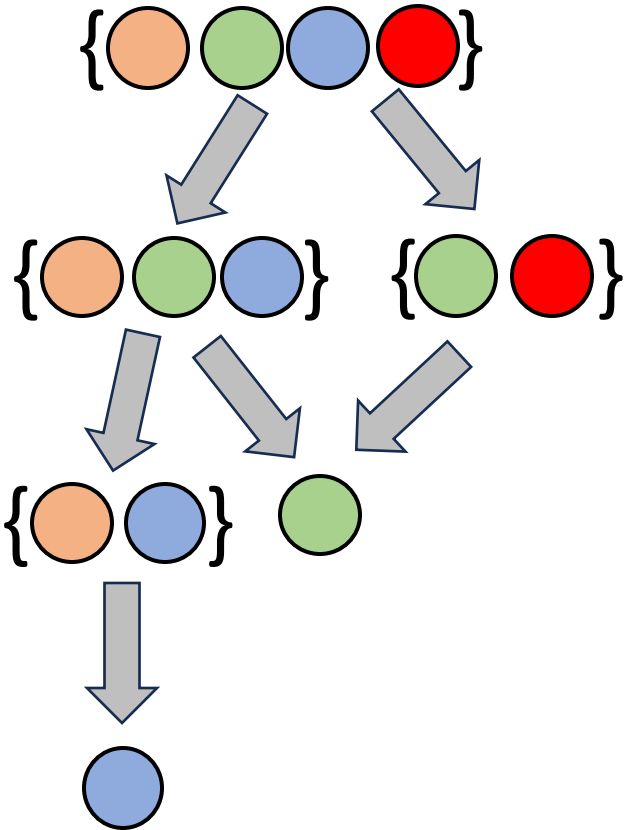
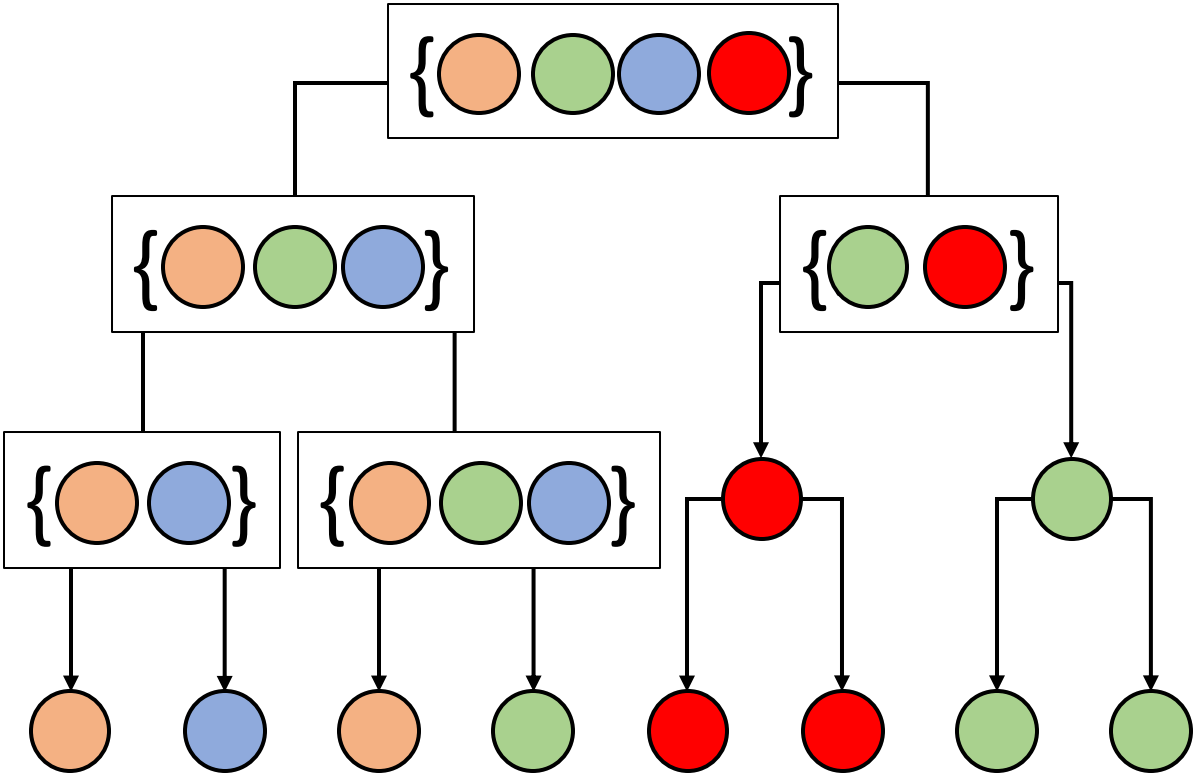
Lymphoid progenitor cells  
differentiates into lymphoid cells

# Cell differentiation map labels ancestors in cell lineage tree



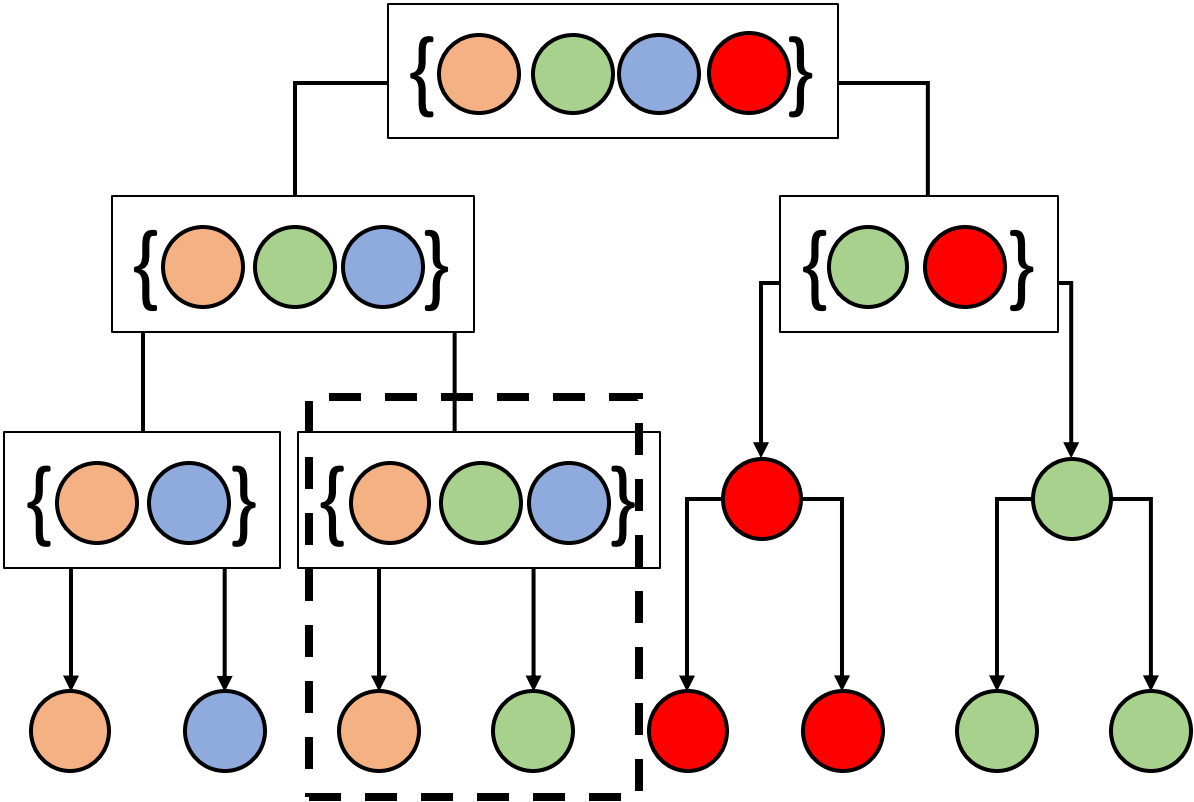
How well does the cell differentiation map fit the data?

# Cell differentiation map labels ancestors in cell lineage tree



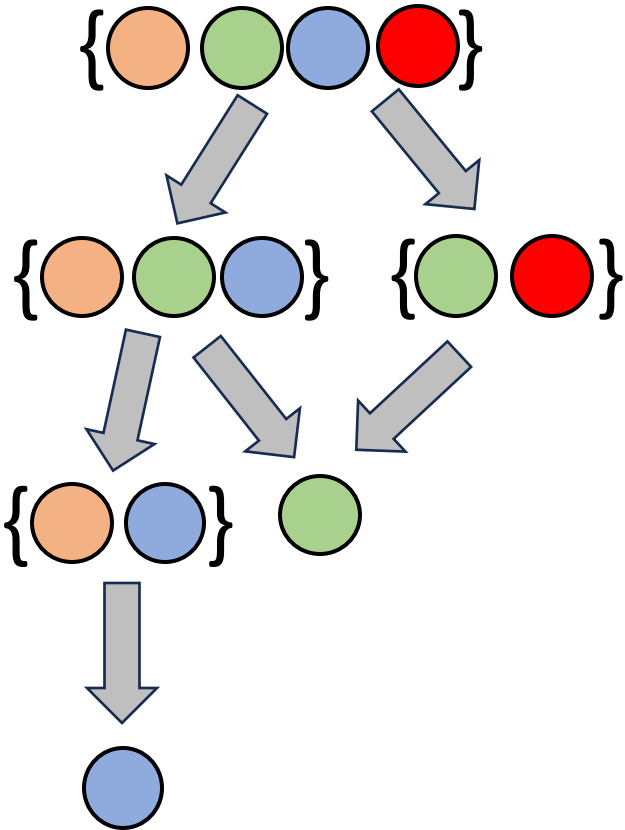
What is the mapping that best fits the data?

# Cell differentiation map labels ancestors in cell lineage tree

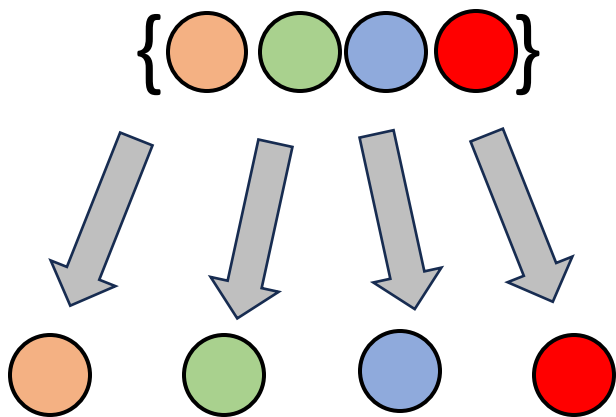


Discrepancy  
between observed  
potency and labeling

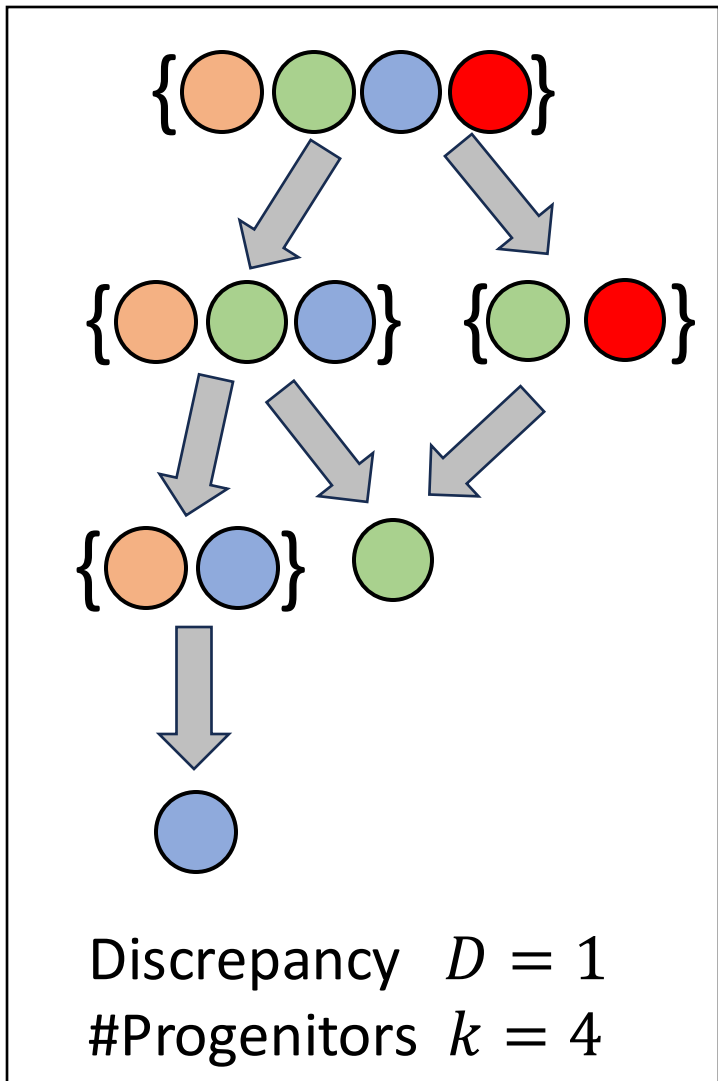
Discrepancy  $D = 1$   
#Progenitors  $k = 4$



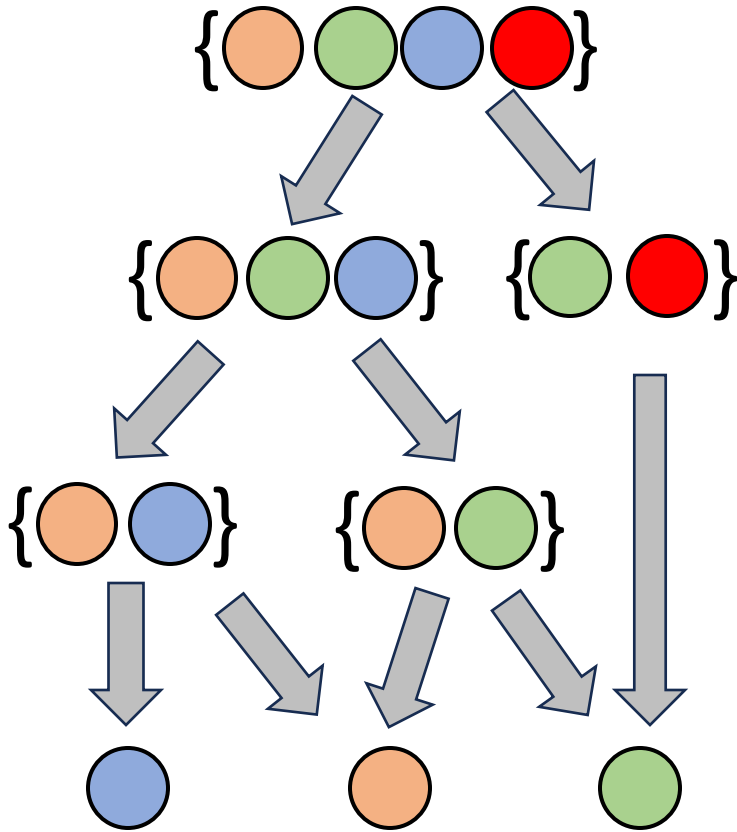
# Characterization of progenitors and cell differentiation map



Discrepancy  $D = 7$   
#Progenitors  $k = 1$



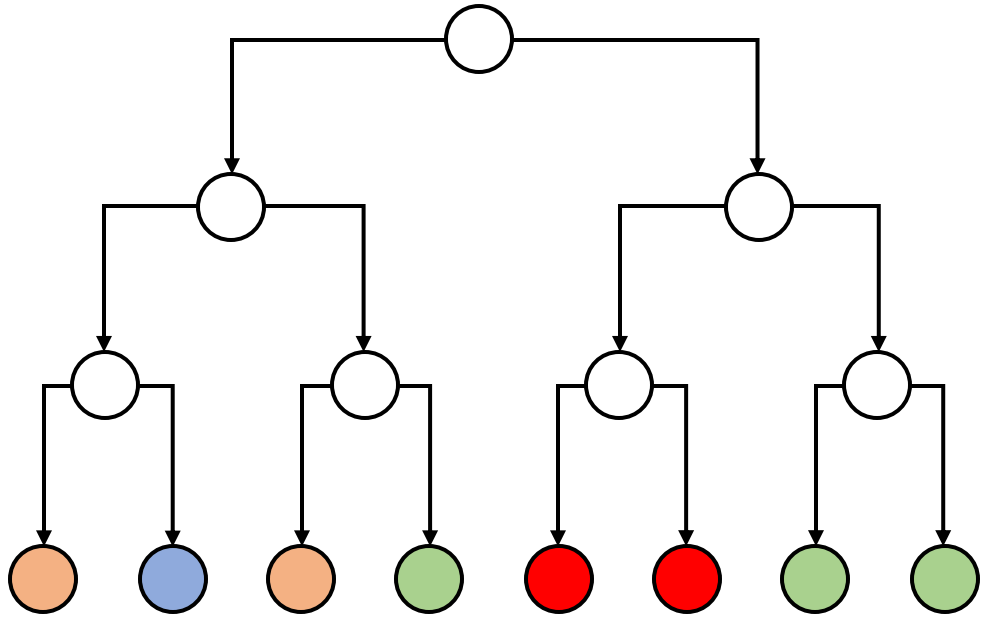
Discrepancy  $D = 1$   
#Progenitors  $k = 4$



Discrepancy  $D = 0$   
#Progenitors  $k = 5$

# Cell differentiation mapping problem

## Input



Leaf labeled cell lineage tree  $T$

$n$  cells,  $m$  cell types

Typically,  $n \gg m$

## Cell Differentiation Mapping (CDM) [Sashittal et al., 2025]

Given a leaf labeled cell lineage tree  $T$  and integer  $k$ , find a cell differentiation map  $F$  with  $k$  progenitors that minimizes discrepancy  $D(T, F)$ .

### Theorem [Sashittal et al., 2025]:

Decision version of CDM Problem is NP-hard.

### Theorem [Sashittal et al., 2025]:

Counting sets of  $k$  progenitors with minimum discrepancy is #P-hard

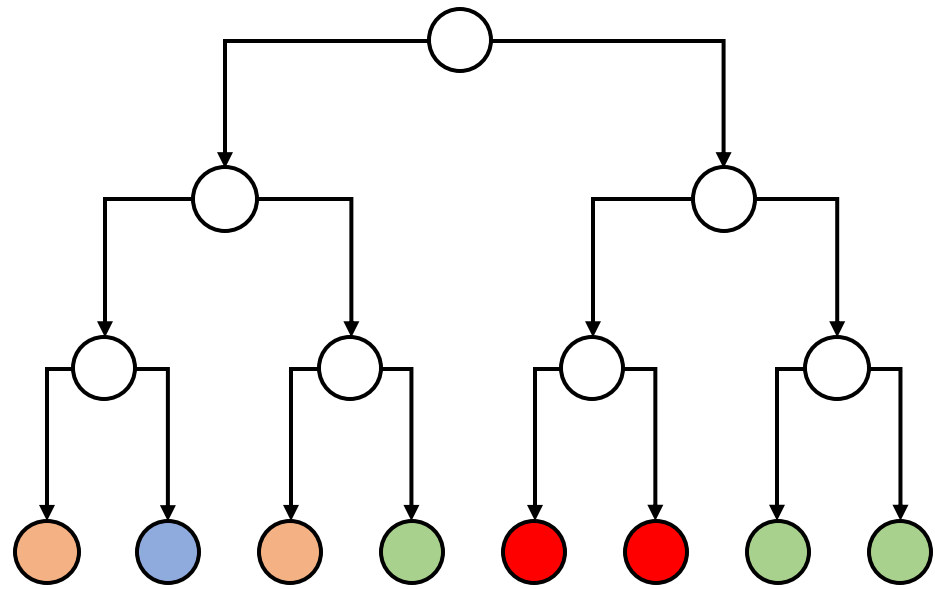
Reduction from *Vertex Cover Problem*

### Theorem [Sashittal et al., 2025]:

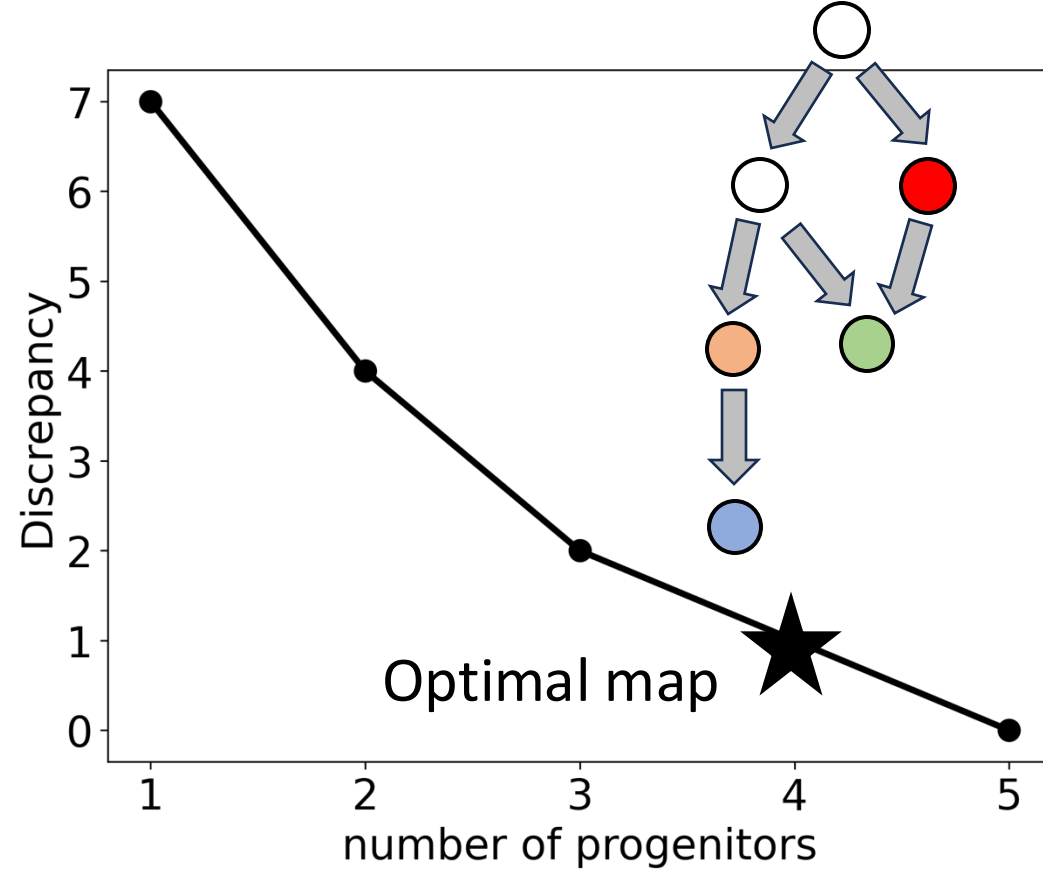
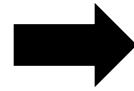
Cell differentiation tree problem is fixed parameter tractable (FPT) in the number  $m$  of cell types.



# CARTA reveals the trade-off between discrepancy and the number of progenitors



Leaf labeled cell lineage tree(s)



We provide a systematic way to test the number of progenitors in the cell differentiation map



Richard Zhang



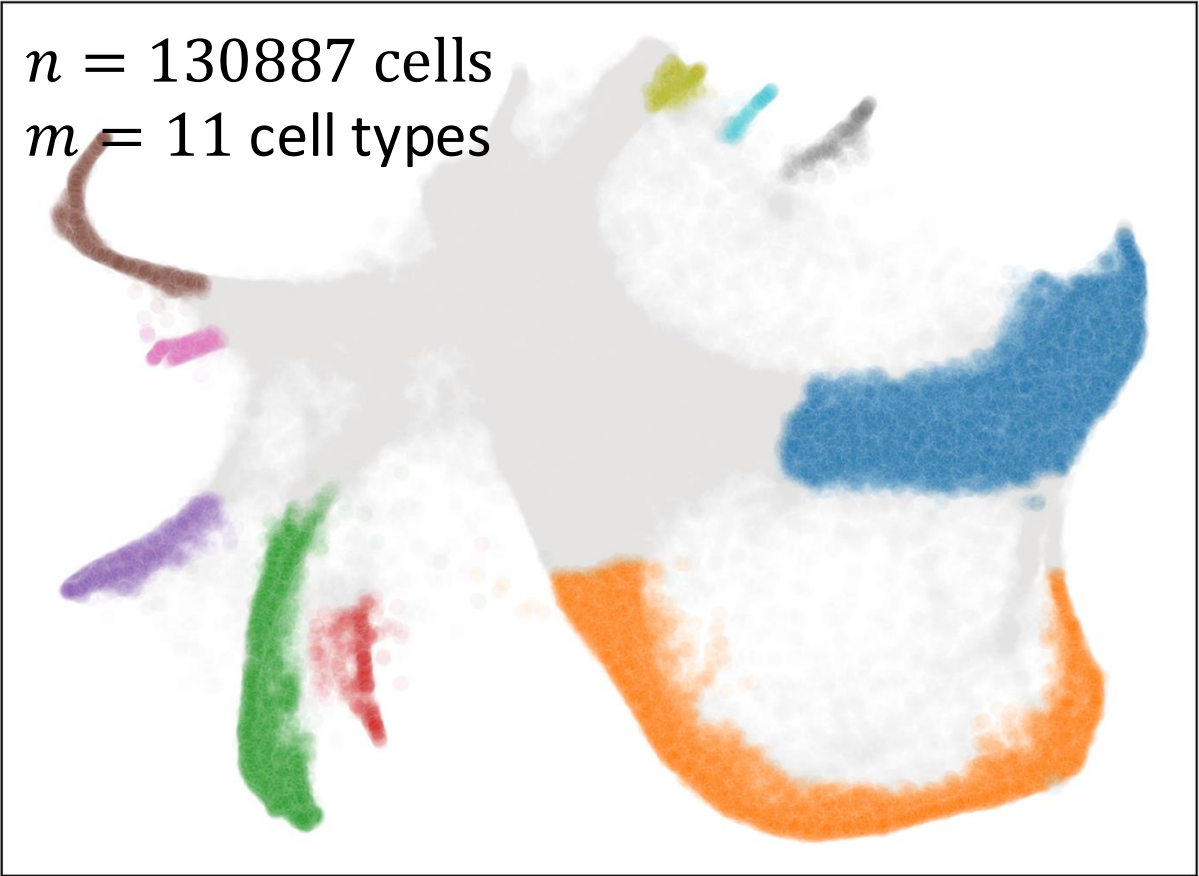
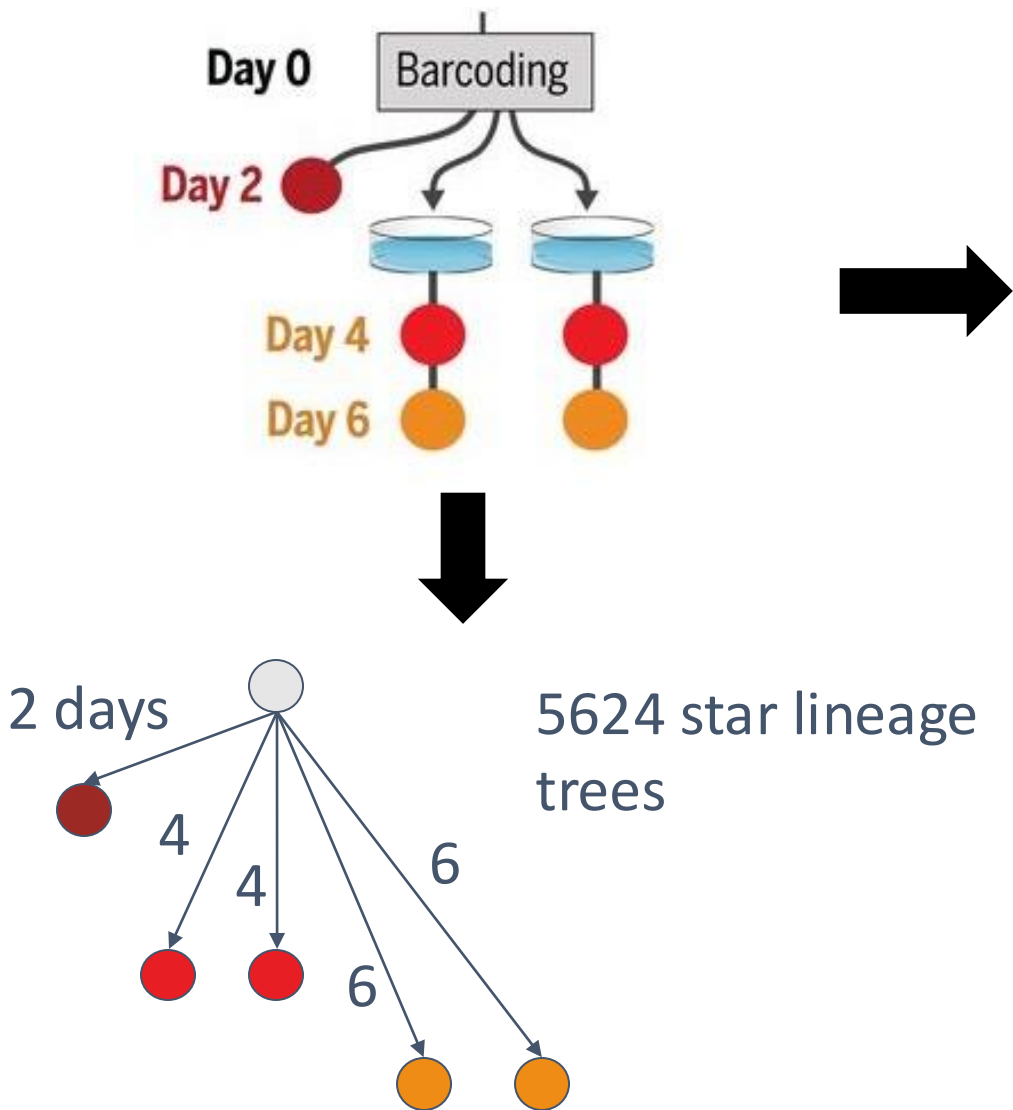
Michelle Chan



Benjamin Raphael

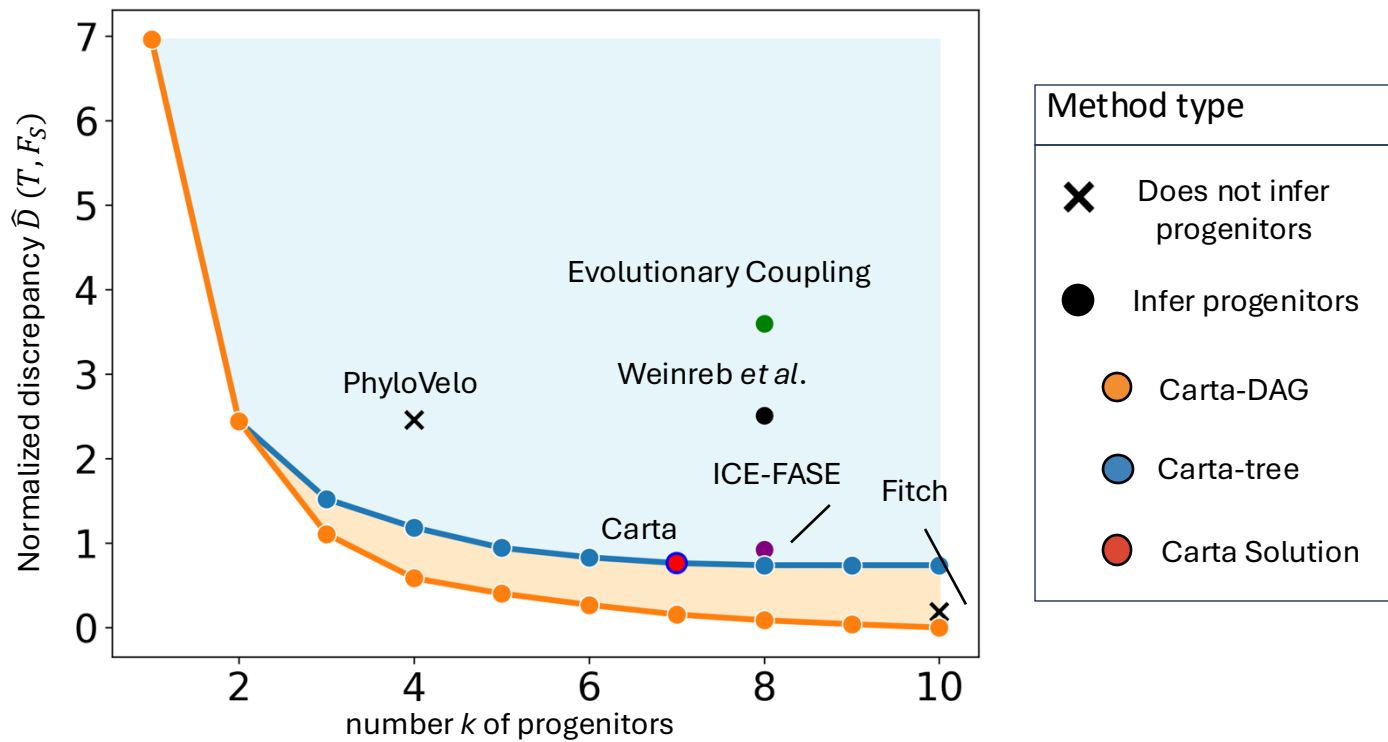
# Mapping differentiation in mouse hematopoiesis

Mouse hematopoietic progenitor cells

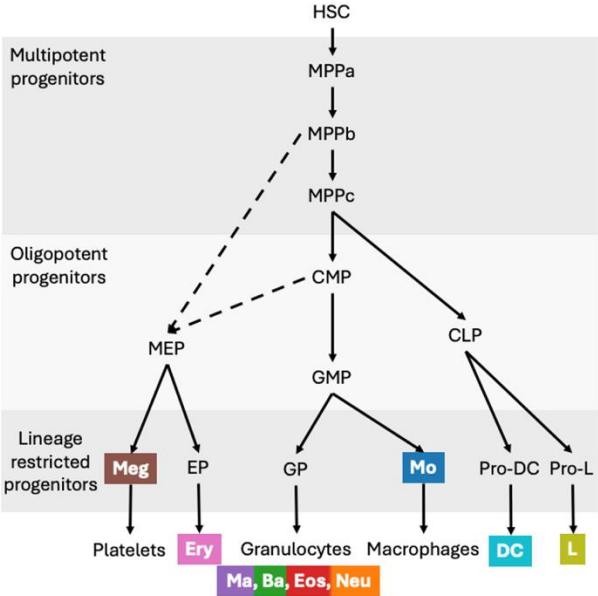


- |                  |                                       |
|------------------|---------------------------------------|
| Undifferentiated | Megakaryocytes (Me)                   |
| Monocytes (M)    | Ccr7+ migratory dendritic cells (CDC) |
| Neutrophils (N)  | Lymphoid (L)                          |
| Basophils (B)    | Eosinophil (Eo)                       |
| Mast cells (Ma)  | Plasmacytoid dendritic cells (PDC)    |
| Erythrocytes (E) |                                       |

# Carta obtains more accurate cell differentiation map

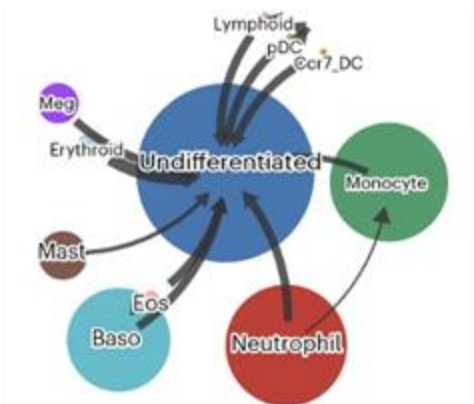


## Hematopoietic differentiation map (Seita and Weissman, 2010)



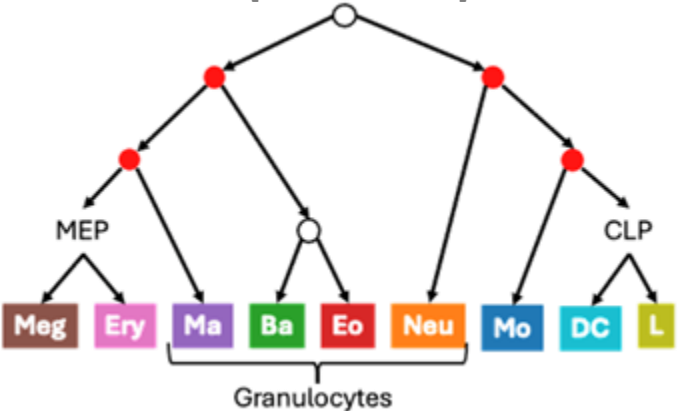
## PhyloVelo

[Wang et al., Nature Biotech. 2023]

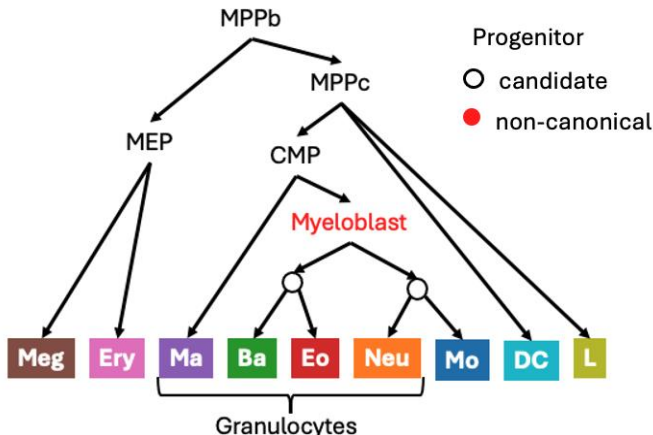


## Weinreb et al.

[Science 2020]

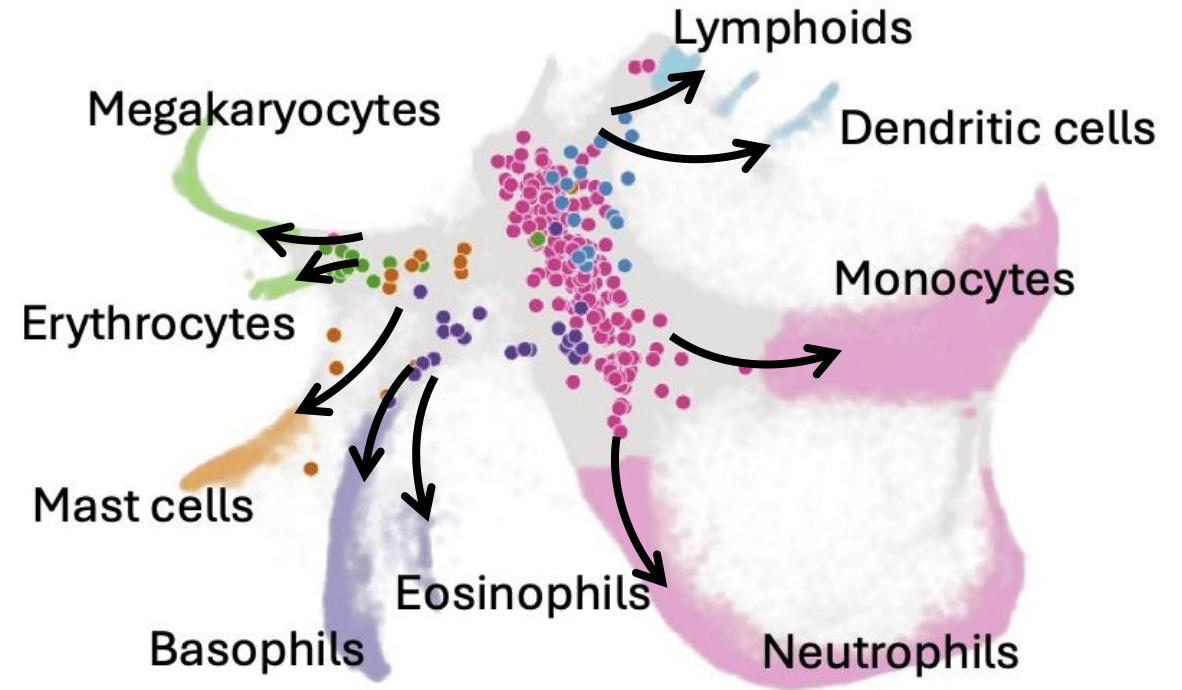
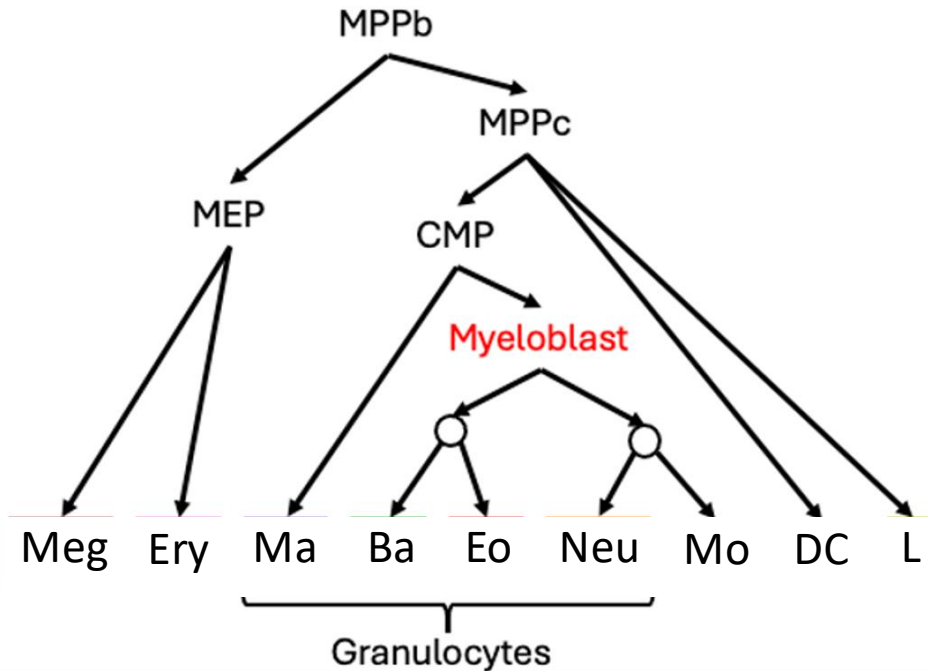


## CARTA



# Carta predicted cell fates align with gene expression

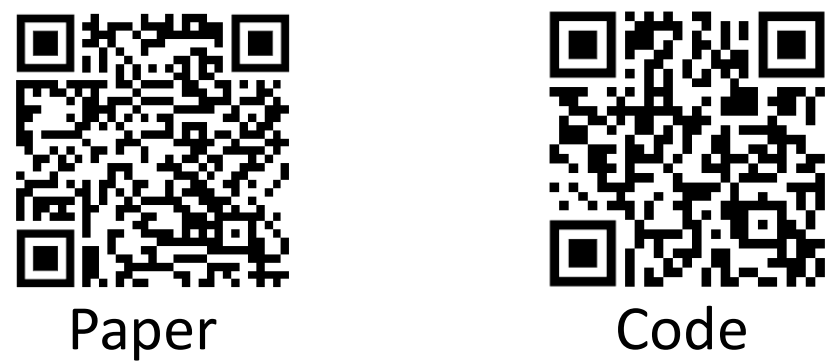
## CARTA



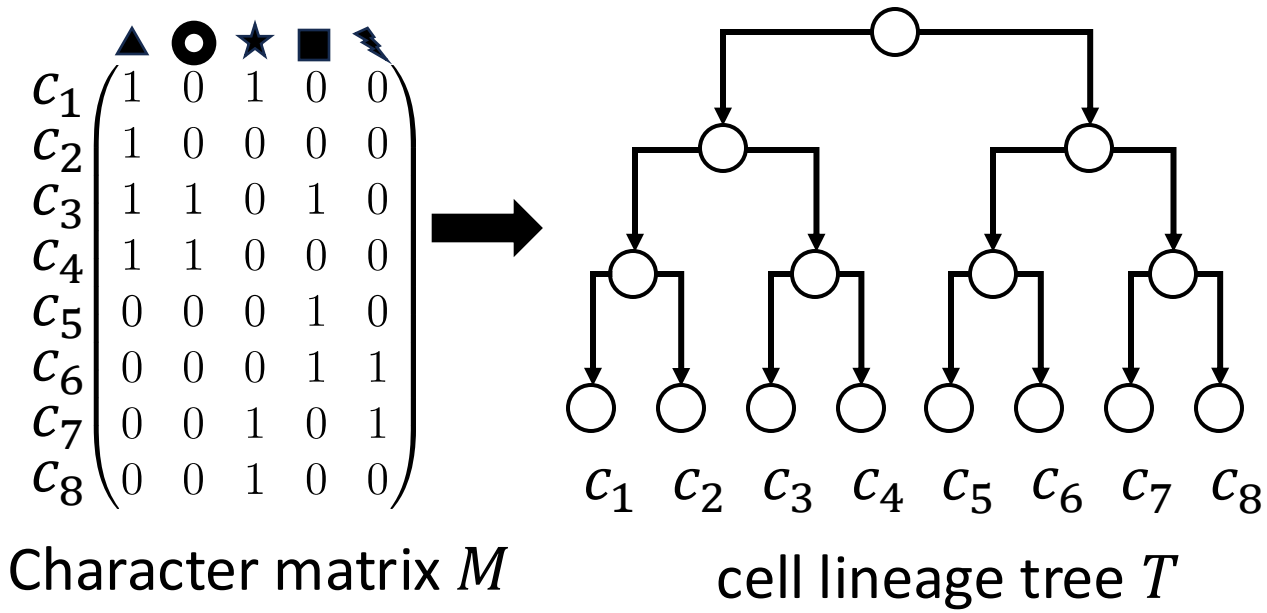
Good agreement of gene expression  
with potency inferred by CARTA



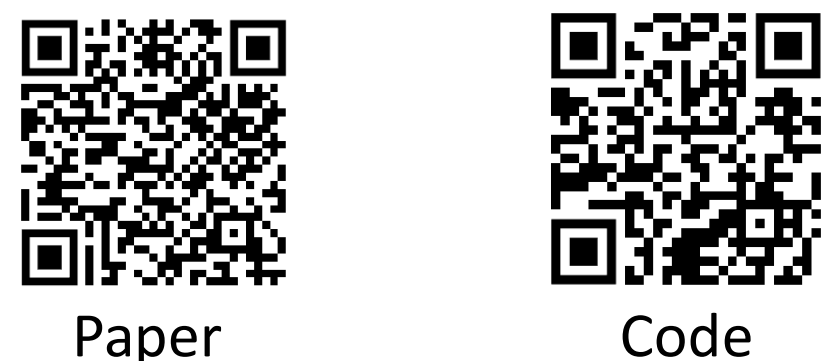
# (1) Cell lineage tracing using Startle



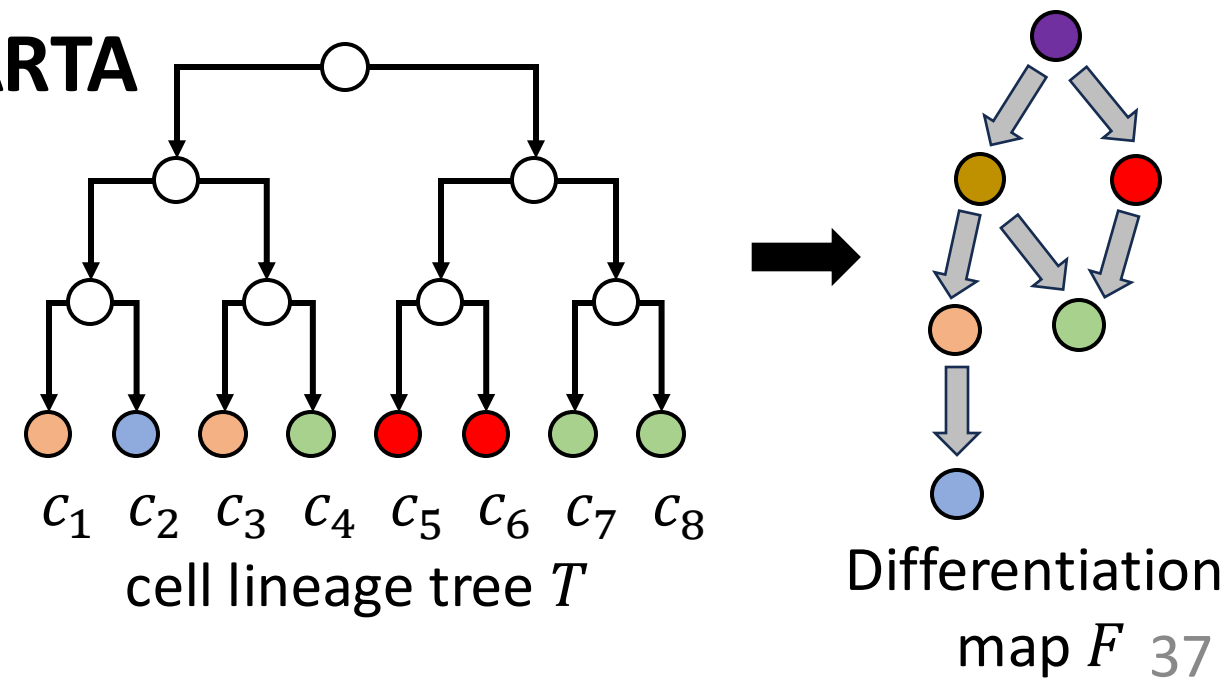
Sashittal\*, Schmidt\* et al., *Cell Systems*, 2023  
Also accepted at RECOMB 2023



# (2) Differentiation mapping using CARTA



Sashittal\*, Zhang\* et al., *Nature Methods*, 2025  
Also accepted at RECOMB 2025



**BACKUP**